PROBIT ANALYSIS

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A STATISTICAL TREATMENT OF THE SIGMOID RESPONSE CURVE

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FOREWORD

IN our work at Rothamsted on insecticides, their action and relative potency, we have been so dependent and have made so many calls on the skill and patience of the Statistical Department and in particular, in recent years, of Mr Finney, that it is a pleasure to learn that the statistical methods and techniques which he has placed so willingly at our disposal for the solution of our various problems are now to be expressed in a more permanent form and to be given to a wider field of workers. If this book receives its due, the investigators of toxicological problems throughout the world will find it a standby.

To many of us, engaged on the practical issues of devising the experimental material and methods to be employed in our laboratory tests, there will be much in these pages that has to be taken upon trust. They are meant to act primarily as an aid in computation, but there is a profound thread of reasoning running through them all, giving a coherence to the several chapters. One cannot but feel, therefore, that the more mathematical readers will find the book suggestive and stimulating.

Twenty-five or more years ago, when I entered the field of research from which Mr Finney takes so many examples for detailed computational study, the very whisper of the need for statistical analysis, falling upon the ears of the biological expert, was enough to bring down a storm of denial upon one's head. Although there may be some small residue of such a reaction still in existence, it now only persists in obscure nooks and crannies of the world of biological research. Much of this change is due to the school of statisticians founded by R. A. Fisher at Rothamsted, of which Mr Finney has been a distinguished member. They always showed a willingness to enter into one's experimental difficulties and an understanding of the limitations imposed by time and space upon the amount of work it was possible to do. The measure of the thought given to these matters can be gauged by the fact that some of the most important tables, given by Mr Finney in the following

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pages, were in process of computation in actual anticipation of problems which we later carried to him. This brings up the now rather hackneyed, but none-the-less important, matter of the prior consultation with a competent statistician before designing experiments. Repeatedly is such a course justified by events, in that the plan can not only be often simplified, but so designed as to yield more information with little, if any, more labour.

Another issue, raised by such text-books as this, concerns the extent to which a training in statistics should enter into the curriculum of the biological and biochemical student. I have found, relatively late in life, how hard a task it is to pick up, during busy years, the requisite amount of basic statistical knowledge to follow the arguments set out in recent text-books written for one's own benefit. I therefore feel that more attention might be given to such a matter by the academic powersthat-be. The engineer may not need to know all there is to be scientifically known about the composition and manufacture of his tools and materials, but at least he should know enough to use them rightly. So too the quantitative biological investigator.

Mr Finney's text-book has a very long and dignified ancestry ranging back to early Egyptian and Mesopotamian times, when texts of mathematics were used for the training of scribes, the professional computers of those distant days. But this is a forward-looking book and is primarily meant for experimentalists. Enlarged editions may well follow this, the first, as the subject grows and new problems arise; and that there are many just round the corner anyone who has discussed these matters with the author is very well aware. Having had the benefit of his personal advice and help throughout the last six years I cannot but wish this book Godspeed on its helpful mission to others.

F. TATTERSFIELD

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PREFACE TO FIRST EDITION

FROM the theory of probability, originally investigated in order to explain nothing more important than the results of games of chance, has developed the science of applied statistics. Over one hundred years ago Laplace wrote that

... la théorie des probabilités n'est au fond, que le bon sens réduit au calcul: elle fait apprécier avec exactitude, ce que les esprits justes sentent par une sorte d'instinct, sans qu'ils puissent souvent s'en rendre compte,

and these words might equally well be written of statistics to-day. In many fields of scientific research, and especially in the biological sciences, numerical studies are complicated by the inherent variability of the material under investigation, and conclusions must be based on averages derived from series of observations. The estimation of these averages and the assessment of their reliability are statistical operations, in the performance of which the experimenter inevitably employs a statistical technique even though he himself may not always recognize this fact. The operations may be simple or complex, depending upon the circumstances; if, however, they cease to be 'le bon sens réduit au calcul', they can no longer be expected to contribute to the understanding of the problem under investigation.

The recent rapid advances in the application of rigorous statistical methods to biological data began with the publication, in 1925, of R. A. Fisher's *Statistical Methods for Research Workers*. Not only did Fisher develop exact methods for the analysis of data from small samples to replace the older approximations from large-sample theory, but he also introduced new and powerful techniques for making the most efficient use of experimental results. Of equal importance to the growth of the present-day philosophy of experimentation was Fisher's suggestion that the statistician should be consulted during the planning of an experiment and not only when statistical analysis of the results is required, as his advice on experimental design

PREFACE TO FIRST EDITION

may greatly increase the value of the results eventually obtained. Since Fisher's book first appeared, the principles of experimental design and the methods of statistical analysis have been extended so rapidly as to make it increasingly difficult for any but the professional statistician to be familiar with the variety of methods needed in biological problems.

Many books since Fisher's have been written with the aim of surveying a wide field of biological statistics, but these can give only an outline of some important topics. There is to-day a need for books in which the specialized statistical methods appropriate to certain branches of science will be discussed in sufficient detail to enable biologists to appreciate them and apply them to their own problems.

One subject requiring fuller discussion than can reasonably be expected in any general text-book of statistics is the method of probit analysis, for the development of which J. H. Gaddum and C. I. Bliss are largely responsible. This method is widely used for the analysis of data from toxicity tests for the assay of insecticides and fungicides, and also of data from other types of assay dependent upon a quantal response. In this book I have tried to give a systematic account of the theory and practice of probit analysis, including as much as possible of the most recent extensions and refinements, in such a form that it may be understood by biologists, chemists, and others who have some knowledge of elementary statistical procedure; at the same time, I have endeavoured to satisfy the mathematical statistician by showing the theoretical background of the method. The less mathematically minded reader will no doubt be content to omit, or at most to read cursorily, Appendix II and other sections concerned with the mathematical basis of the technique. Full understanding and appreciation of statistical methods can be gained only by experience in their use, but careful study of the numerical examples should enable many who were previously unfamiliar with probit analysis to apply it satisfactorily to their own data.

This book has been written as a result of several years of close collaboration with members of the Insecticides Department

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at Rothamsted Experimental Station, especially with Dr F. Tattersfield, Dr C. Potter, and, until he left Rothamsted, Dr J. T. Martin. I wish to express my gratitude to them for discussing with me a wide variety of their problems, for advising me on the experimental aspects of their results, and for the generosity with which they have permitted me to use their data both in this book and in earlier publications. I am also very grateful to my colleagues in the Statistical Department at Rothamsted for much helpful discussion, and particularly to Dr F. Yates for his detailed and constructive criticism in the preparation of my book. Others to whom my thanks are due include Miss G. M. Ellinger for assistance in German translation. Dr C. G. Butler for permission to use the numerical data of Ex. 33, Dr A. E. Dimond and Dr J. G. Horsfall for giving me very full information on the results discussed in § 41 and for permission to use their data, Professor G. H. Thomson for assistance in tracing the history of the probit method, the Editors of the Annals of Applied Biology for permission to reproduce the first half of Table II, Professor R. A. Fisher, Dr F. Yates, and Messrs Oliver and Boyd, Ltd. for permission to reproduce Tables I, VI and VII from their book Statistical Tables for Biological, Agricultural and Medical Research, and my father, Robt. G. S. Finney, for very considerable help in the correction of proofs.

D. J. FINNEY

RQTHAMSTED EXPERIMENTAL STATION

August 1945

PREFACE TO SECOND EDITION

In the four years that have elapsed since the first edition of this book was completed, the theory of the analysis of quantal response data has been extended and new applications of probit analysis and related methods have been made. To discuss these

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in logical order would entail considerable rearrangement of the book, a course that was impracticable for this edition. Instead, I have adopted the compromise of correcting a small number of minor errors, of rewriting §§ 15, 28, parts of §§ 45 and 47, and Appendix II, and of adding a new chapter on Recent Developments. Table II has been extended to 90 % natural mortality, and additional information has been incorporated into Table V; the new Tables VIII and IX aid the analysis of problems discussed in Chapter 11.

I am aware of three major shortcomings that could not be removed without a more radical revision of the book. I have not discussed as fully as I should now choose to do alternative methods for the analysis of quantal response data, especially that based upon the angle transformation (\S 15) and approximations such as Kärber's (\S 13, 49). My examples give the impression that probits are useful primarily for data from insecticide tests, whereas in fact they find application in many other branches of science. I may seem to imply that probit analysis is a statistical technique co-extensive with the analysis of biological assays; in fact, only one class of biological assays has any need of probits or related techniques, and probit analysis is used in the statistical examination of many experiments that would not generally be regarded as assays.

I am indebted to all who, privately and in reviews, have told me of errors in the first edition or have made suggestions which have helped the preparation of this edition. I am especially grateful to Mr P. S. Hewlett and Mr R. L. Plackett, for sending me a draft of a new paper and allowing me to comment on it in § 53, and to the Editors of *Biometrika* for permission to include Table VIII.

D. J. FINNEY

OXFORD

December 1949

Chapter 1

INTRODUCTORY

1. BIOLOGICAL ASSAY

THE term *biological assay*, in its widest sense, should be understood to mean the measurement of the potency of any stimulus, physical, chemical or biological, physiological or psychological, by means of the reactions which it produces in living matter. The biological method of measuring the stimulus is adopted either for lack of any alternative, or because an exact physical or chemical measurement of stimulus intensity may need translation into biological units before it can be put to practical use.

Biological assay is most commonly considered as referring to the assessment of the potency of vitamins, hormones, toxicants and drugs of all types by means of the responses produced when loses of these are given to suitable experimental animals. Estination of the potency of a natural product, such as a drug extracted from plant material, in producing a biological effect of a certain type, is often impossible or impracticable by chemical analysis. Even if the chemical constitution of the material is known or determinable, there may be little knowledge of the magnitude of the effect which the constituents will produce, a difficulty not confined to natural products but occurring also with many manufactured compounds, such as insecticides, which are made to precise chemical specifications yet which are of unknown biological activity. The material must in fact be tested and standardized by methods appropriate to its future use.

For example, vitamin assays may be made in terms of weight changes or other physical measurements observed in rats, the effects of different doses of the preparation to be assayed being compared with the effects of a standard in order to estimate the relative potency of the test preparation and the standard. Insulin may be assayed in terms of the fall in blood sugar in injected rabbits, and digitalis by the mortality amongst injected cats. Again, the potency of insecticides may be assessed by means of

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the mortality in batches of treated insects, and that of fungicides by the proportion of treated spores failing to germinate. Another form of assay procedure which is sometimes useful depends on measurement of the time required for the production of a specified effect instead of measurement of the magnitude of the effect produced. In an interesting and informative article, which should be read by all who are seriously concerned with this type of investigation, Bliss and Cattell (1943) have reviewed nearly 300 recently published papers on the theory and practice of biological assay, with especial reference to vitamin, hormone, and drug assay. The texts of Burn (1950) and Coward (1938) may also be consulted, though the statistical methods there advocated do not fully exploit modern developments.

One type of assay which has been found valuable in many different fields, but especially in toxicological studies, is that dependent upon the quantal, or all-or-nothing, response. Though quantitative measurement of a response is always to be preferred when available, there are certain responses which permit of no graduation and which can only be expressed as 'occurring' or 'not-occurring'. The most obvious example of this kind of response is death; although workers with insects have often found difficulty in deciding precisely when an insect is dead (Tattersfield et al. 1925), in many investigations the only practical interest lies in whether or not a test insect is dead, or perhaps in whether or not it has reached a degree of inactivity such as is thought certain to be followed by early death. In fungicidal investigations, failure of a spore to germinate is a quantal response of similar importance. In studies of drug potency, the response may be the cur of some particular morbid condition, no possibility of partial cur being under consideration.

This book is concerned chiefly with the statistical techniqu needed in the analysis of quantal response data, rather than wi the general theory of biological assay. Though most of the d cussions are presented in terms of tests of the potencies of secticides, the methods are applicable to many other types data, and are not peculiar to those relating to assays in strict sense.

2. VARIABILITY OF RESPONSES

One feature possessed by all biological assays is the variability in the reaction of the test subjects and the consequent impossibility of reproducing at will the same result in successive trials, however carefully the experimental conditions are controlled. Though similar variability may be encountered in assays based only on purely physical or chemical measurements, it is generally then of far less practical importance. The contrast between the physical approach and the biological may be seen from a consideration of two methods for the estimation of the ratio of two unknown weights. The physical method is to balance each in turn against a set of standardized weights, and to take as the required estimate the ratio of their magnitudes. There may be technical difficulties in carrying out the operations of weighing to very high accuracy, and both the quality of the balance and the competence of the operator are important factors, but for most practical purposes the reproducibility of the results is not called in question; one measurement on each weight will usually suffice to determine the ratio with an accuracy far beyond that obtainable in any biological assay.

The physical assay of the ratio is here so simple that no alternative method is needed. For the sake of the illustration it may be compared with a biological technique, using quantal responses, in which the weights are dropped from a fixed height on to the heads of live rats. Data for the assay are provided by the records of death or survival. That the first weight, at its first trial, killed a rat, while the second weight did not, would not show with any certainty that the first was the heavier, still less would it give any clue to their ratio; the effect would be influenced not only by the weight dropped, but also by the age, sex, size and physical condition of the rat, and other biological and environmental factors (as well as, of course, the shape and elasticity of the weights, which will here be assumed the same for both). If batches of rats, chosen at random from the stock available, were tested with each weight, the proportionate effect of variation in susceptibility from rat to rat would be reduced with increasing size of sample, and

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the weights could be compared in terms of the two mortality rates. Variability could be still further controlled, though never entirely eliminated, by using a specially bred strain of rats, and selecting batches homogeneous for sex, age and other relevant factors. When every test is made from the same arbitrary height, this assay cannot discriminate between weights too light to cause any deaths or between weights so heavy as to kill every rat. This difficulty can be overcome by making tests from a series of different heights and obtaining a range of mortalities for each weight. The weights are then compared in terms of equivalent heights, or heights estimated to give the same (say 50 %) mortality. The height scale thus provides a basis for the biological comparison of any number of weights, but, without experimental or theoretical knowledge of the law relating mortality to height and the physical measure of weight, the results of the biological assay cannot be transformed to purely physical terms.

This example has been discussed in some detail, as, in spite of its absurdity, it illustrates the necessity for a careful consideration of variability in any biological assay technique. To some extent the quantal nature of the responses is a complication, but quantitative responses by no means provide an escape from the problem. Equal doses of insulin will not produce equal effects on the blood sugar of different rabbits, or even on the blood sugar of the same rabbit at different times. Consequently, though two insulin preparations could be compared in terms of the magnitudes of the changes in blood sugar produced in two rabbits, only repetition of the tests on several rabbits for each preparation can give an estimate of the relative potency sufficiently precise to be of any practical value.

Biological aspects of, and reasons for, variability in test organisms of many kinds have been discussed by Clark (1933, especially Chapter VI), and his remarks on individual variations in response deserve careful reading. The occurrence of this variability introduces considerations other than those of biology; when there is a large natural variability of response amongst the test subjects, the analysis of numerical data for the estimation of the effects of applied treatments can only be effected satisfactorily with the aid of exact statistical techniques.

3. STATISTICAL METHODS

The development of statistical techniques for the analysis of biological data of all types has proceeded with great rapidity in recent years. In many fields of research on biological topics, experimental and observational results can only be used to the best advantage by subjecting them to precise and critical statistical examination. When a programme of biological research involves the collection of numerical data, the problem of interpreting these is almost inevitably one of statistics. The choice is not, as the biologist sometimes imagines, of whether his figures shall be 'statistically analysed' or not, but rather of whether the analysis shall be theoretically sound and able to extract all the relevant information from the material, or inadequate and possibly unsound. Even the simplest and most straightforward averaging of results is essentially a statistical process; the analysis appropriate to any body of data is determined by the inherent properties of those data, not by the whim of the statistician. It is unfortunate, to say the least, that good experimental work should ever be followed by a statistical treatment of the results so unsatisfactory that the conclusions are incomplete, unreliable, or even actively misleading.

The function of the statistician in biological investigations is to supply that critical and objective judgement of numerical material which is a product of his specialized training and experience. An important aspect of his work is co-operation in the planning of an experimental programme so that, taking into account all relevant information already available, it is designed to give results of maximum utility and precision. The assistance of a competent statistician from the beginning of the programme will often substantially increase the value of the results obtained from a given amount of experimental time and labour, in respect of both their scope and their reliability, whereas the conclusions may be much less satisfactory if the statistician is only consulted after the completion of the experimental work.

Nevertheless, the methods of analysis used by the statistician are not esoteric mysteries, but are simply instruments for

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discovering the most important features of numerical data. The computational procedures appropriate to many types of data have been so far standardized that they can be applied by a biologist who has some understanding of their purposes, even though he may know little of their theoretical foundations. The blind application of formulae is a danger which should be avoided, for not infrequently the formulae may be used quite inappropriately; on the other hand, the anxiety of many biologists to learn enough of statistical methods in order to be able to analyse their own data without complete dependence on the assistance of a statistician is witnessed by the recent spate of books designed to instruct the non-mathematician in statistical technique.

The statistical treatment of quantal assay data has been much aided by the development of *probit analysis*. This method, which is usually attributed to Gaddum (1933) and Bliss (1934*a*, *b*; 1935*a*, *b*) though it has, in fact, a much longer history (§ 14), has now been widely adopted as the standard method of reducing the data to simple terms.

4. SUMMARY OF CONTENTS

This book is written with the intention of introducing the probit method to many who have previously not ventured to use it, and of presenting some of its more recent developments to those who are already familiar with it. In the first few chapters the technique is shown in its simplest form, stripped of all but the essentials. It is hoped that these chapters, at least, will be capable of appreciation and use by many whose knowledge of other branches of statistics is small. Even for this purpose, however, a slight acquaintance with modern statistical thought and terminology is necessary, and although notes on various tests and distributions will be found in the appropriate sections these can do little more than give references and hints on particular applications. The reader is strongly recommended to familiarize himself with the relevant portions of R. A. Fisher's Statistical Methods for Research Workers (1944), especially the sections dealing with the normal, t, and χ^2 distributions, and with regression. K. Mather's Statistical

SUMMARY OF CONTENTS

Analysis in Biology (1943) provides a valuable introduction for those who find Fisher's book too difficult.

The numerical examples in subsequent chapters have been carefully chosen to illustrate many points of procedure and to show the application of the method to a variety of toxicological data. Though the computational work required is sometimes laborious, it is not as heavy as some accounts have made it appear; in an appendix is given a detailed description of a systematic arrangement of the computations for the simplest type of problem, and this arrangement may easily be extended to suit more complex data. A second appendix gives a brief outline of the mathematical theory of the probit method. The book is completed by a series of tables which lessen considerably the computing time and labour required for probit analysis.

Chapter 2

QUANTAL RESPONSES AND THE DOSAGE-RESPONSE CURVE

5. THE FREQUENCY DISTRIBUTION OF TOLERANCE

In all biological assays there are two components to be considered, the *stimulus* (for example, a vitamin, a drug, a physical force, or a mental test) and the *subject* (for example, an animal, a plant, a piece of tissue, or a single cell). The stimulus is applied to the subject at an intensity specified in units of concentration, weight, time, or other appropriate measure and under environmental conditions as carefully controlled as is practicable, as a result of which a *response* is produced by the subject. Different stimuli are then compared in terms of the magnitudes of the responses they produce, or, more commonly and usefully, in terms of the intensities required to produce equal responses.

When the characteristic response is quantal, its occurrence or non-occurrence will depend upon the intensity of the stimulus applied. For any one subject, under controlled conditions, there will be a certain level of intensity below which the response does not occur and above which the response occurs; in psychology such a value is designated the threshold or limen, but in pharmacology and toxicology the term tolerance seems more appropriate. This tolerance value will vary from one member to another of the population used, frequently between quite wide limits. When the characteristic response is quantitative, the stimulus intensity needed to produce a response of any given magnitude will show similar variation between individuals. In either case, the value for an individual also is likely to vary from one occasion to another as a result of uncontrolled internal or external conditions. Clark (1933, Chapter VI) has discussed the nature of these individual variations in response for many different populations.

For quantal response data it is therefore necessary to consider the distribution of tolerances over the population studied. If the

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dose, or intensity of the stimulus, is measured by λ , the distribu tion of tolerances may be expressed by

$$dP = f(\lambda) d\lambda; \qquad (2.1)$$

this equation states that a proportion, dP, of the whole population consists of individuals whose tolerances lie between λ and $\lambda + d\lambda$, where $d\lambda$ represents a small interval on the dose scale, and that dP is the length of this interval multiplied by the appropriate value of the distribution function, $f(\lambda)$.

If a dose λ_0 is given to the whole population, all individuals will respond whose tolerances are less than λ_0 , and the proportion of these is P, where

$$P = \int_{0}^{\lambda_{\bullet}} f(\lambda) \, d\lambda; \qquad (2.2)$$

the measure of dose is here assumed to be a quantity which can conceivably range from zero to $+\infty$, response being certain for very high doses so that

$$\int_0^\infty f(\lambda)\,d\lambda=1.$$

The distribution of tolerances, as measured on the natural scale, may be markedly skew, but it is often possible, by a simple transformation of the scale of measurement, to obtain a distribution which is approximately normal. 'A variate is said to be normally distributed when it takes all values from $-\infty$ to $+\infty$ with frequencies given by a definite mathematical law, namely, that the logarithm of the frequency at any distance dfrom the centre of the distribution is less than the logarithm of the frequency at the centre by a quantity proportional to d^2 . The distribution is therefore symmetrical, with the greatest frequency at the centre; although the variation is unlimited, the frequency falls off to exceedingly small values at any considerable distance from the centre, since a large negative logarithm corresponds to a very small number' (Fisher, 1944, § 12). In tests of insecticidal sprays, for example, although the distribution of tolerance concentration of the toxic agent is usually far from symmetrical on

* The statement that $f(\lambda)$ is a function of λ means simply that for any given value of λ the value of $f(\lambda)$ is uniquely determined.

account of a few insects with extremely high tolerances providing an extended 'tail' to the distribution (Fig. 1), normalization can often be effected by expressing the tolerances in terms of the logarithms of the concentrations instead of the absolute values (Fig. 2); this transformation is now accepted as standard practice for expressing the results of such trials (cf. Galton, 1879). Various writers (Clark, 1933; Hemmingsen, 1933; Bliss, 1935a) have sought an explanation of the normal distribution of log tolerances in the Weber-Fechner law and in adsorption phenomena, particularly as expressed by the Langmuir adsorption law, but these explanations are beyond the scope of this book. The validity and appropriateness of the logarithmic transformation in the analysis of experimental data are not dependent on the truth or falsity of any hypotheses relating to adsorption; use of the log concentration as measuring the dosage in insecticidal trials requires no more justification than that it introduces a simplification into the analysis. There are additional advantages in having a scale on which a given proportionate increase in concentration has the same scale value at all levels of concentration, but other forms of transformation may sometimes be more suitable. Parker-Rhodes (1941, 1942a, b) has advanced reasons for expecting a normal distribution of some fractional power of the concentration of a fungicide to which suspensions of fungus spores are exposed (see § 45), though this must be only an approximation which holds over a restricted range of concentrations.

It is convenient to take x as representing the intensity of the stimulus on the scale on which the tolerances are normally distributed, and λ as the untransformed value of concentration, time of exposure, or other variate. Thus for much insecticidal work, if λ is the concentration of the toxic agent,

$$x = \log_{10}\lambda, \qquad (2.3)$$

and for some fungicides a better transformation may be

$$x = \lambda^{i}, \qquad (2.4)$$

where usually $i \leq 1$. The second normalizing transformation tends to the logarithmic as i is decreased to zero. There is no reason why

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a simple transformation should always be available; nevertheless, many classes of data have been found amenable to treatment on

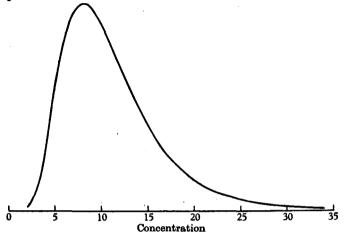
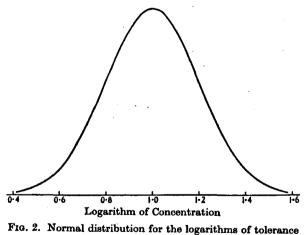


FIG. 1. Typical distribution curve for the absolute values of tolerance concentrations of an insecticide. (The area between any two ordinates represents the proportion of insects having tolerances lying between these limits.)



concentrations, derived from Fig. 1.

these lines, so that the study of the consequences of this normal distribution and of the appropriate methods of statistical analysis is of considerable practical importance. In order to distinguish

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the scales of measurement, the word *dose* will be restricted to the scale of λ , in which measurements of stimulus are in fact made, and x will be referred to as the measure of dosage, or, more briefly, just the *dosage*.

In an investigation for which tolerance can be satisfactorily defined, so that for any given dose all individuals with equal or lower tolerance values will respond, a graph of the percentage responding against the dose will give a steadily rising curve. The rate of increase in response per unit increase in dose is frequently very low in the region of zero or 100 % response, but higher in the intermediate region, so that the curve is sigmoidal (Fig. 3). When the stimulus is measured in dosage units, the curve takes the characteristic normal sigmoid form (Fig. 4). This curve does not attain the zero or 100 % response except at infinitely low or infinitely high dosage, a situation which cannot truly arise (except that, when the measure of dosage intensity is logarithmic, an infinitely low value represents zero dose). Nevertheless, the distribution may be effectively normal over the range of values which is of practical interest, the disagreement between theory and fact outside this range being of negligible importance.

6. DIRECT MEASUREMENT OF TOLERANCE

The tolerance of the test subject in respect of a given stimulus can sometimes be measured directly. Such direct measurement, for example, is involved in the 'cat' method for the assay of digitalis, in which anaesthetized cats are given a continuous slow intravenous infusion of digitalis until death occurs. If there is any appreciable time lag between the introduction of the drug and its taking effect, the lethal dose will be overestimated. Though there is no certainty that the dose required to cause death under conditions of slow application will be the same as the tolerance for more rapid application, the technique has proved suitable for assaying a preparation of unknown potency in terms of a standard.

An alternative method is to give to each subject successive doses of different intensities, allowing a suitable time interval

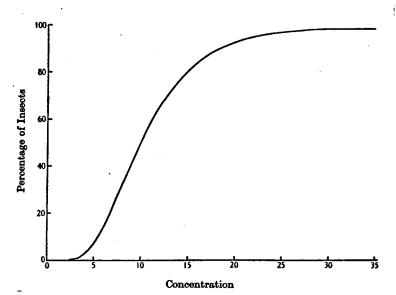


FIG. 3. Sigmoid curve derived from Fig. 1 to show percentage of insects whose tolerances are less than a specified value.

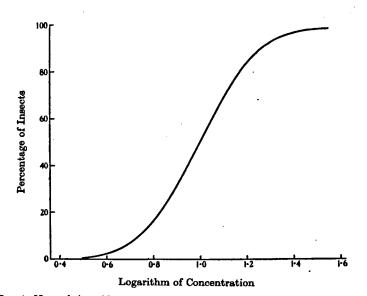


FIG. 4. Normal sigmoid curve derived from Fig. 2 to show percentage of insects whose log tolerances are less than a specified value.

after each for a return to normal and making the differences sufficiently small for a satisfactory determination of the lowest dose which causes the characteristic response. With an irreversible response, such as death, the doses would have to be given in an increasing series. For the method to be satisfactory, there must be no cumulative effect of doses already given, either as lowering or increasing the resistance of the subject, a condition which severely limits its applicability.

In either of these methods of direct measurement of tolerance, the appropriate methods of statistical analysis are the same as for other types of biological measurements. If the tolerance of each subject has been separately and independently determined, the set of values obtained may be subjected to the same analytical processes as measurements of length or weight; the estimation of means and standard errors, the comparison of distributions, and the making of tests of significance present no new features. Bliss and Hanson (1939), for example, have discussed the application of the analysis of variance and covariance to assays based on the 'cat' method.

Direct tolerance measurement is often impracticable on account of the amount of time required for either of the methods mentioned above. Even more frequently it is ruled out entirely by the nature of the problem. It is hard to conceive of any direct measurement technique for the poison tolerance of an insect, still less for that of a fungus spore. In these circumstances an entirely different approach must be adopted, and the potency of the stimulus must be assessed by means of the proportion of subjects, in random samples of the population, showing responses at different levels of dose.

7. THE BINOMIAL DISTRIBUTION

If an insect, selected at random from a population, is exposed to a dose λ_0 of a poison, the probability that it will respond is P; the probability of its failing to respond is (1-P), a quantity usually denoted by Q. The dose here may be measured by the concentration of toxic substance, the absolute quantity used, the

length of time of exposure to a fixed set of conditions, or some combination of these or other factors. If two insects are exposed to the dose, and if their reactions are completely independent, the probability that both respond is P^2 , and the probability that both fail to respond is Q^2 ; the probability that only the first responds is $P \times Q$, and the probability that only the second responds is $Q \times P$. Thus the total probabilities of 2, 1 and 0 responding are P^2 , 2PQ and Q^2 respectively, the successive terms in the expansion of $(P+Q)^2$. In a similar manner it may be seen that if a batch of n insects is exposed to the dose λ_0 , and all react independently, the probabilities of n, (n-1), (n-2), ..., 2, 1, 0 responding are the (n+1) terms in the expansion of the binomial $(P+Q)^n$. The probability of exactly r responding is therefore

$$\frac{n!}{r!(n-r)!}P^rQ^{n-r}.$$

This is known as the *Binomial Distribution* of probabilities (cf. Fisher, 1944, § 18; Mather, 1943, § 5). The average number responding in repeated batches of n from the same population is nP, and the average number failing to respond nQ. Durham *et al.* (1929) have given useful tables of sums of terms from this distribution, and Clopper and Pearson (1934) have shown similar results in the form of charts.

The reactions of separate members of a batch to the stimulus of a particular dose are not always independent; a correlation of response may result from incomplete randomness of selection of the batch, or alternatively from unsatisfactory control of experimental conditions causing the number responding to be seriously affected by some factor other than the dose. For example, if each batch consists of insects from a single brood, insects from one batch are likely to be more alike in tolerance than those from different batches, and the variation between batches in the numbers responding will be greater than that for the binomial distribution. Again, the susceptibility of insects to an insecticide might be greatly influenced by temperature; if the temperature during the tests was permitted to vary substantially from one batch to another, the variance of the numbers responding to

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a given dose of insecticide would be inflated. The extreme situation is that, in every batch tested, either all members respond or all fail to respond, so that the evidence from a batch is no more reliable than that from an individual. Whatever the cause, such heterogeneity must make the weight to be attached to the data less than is appropriate to the binomial distribution (Bliss, 1935*a*; Parker-Rhodes, 1941).

The result of testing a series of doses, each on a separate batch of insects, is to obtain for each dose a proportion, p, of insects in the batch which show the characteristic response and whose tolerances are therefore lower than that dose. Each value of pis an estimate of the corresponding P, the proportion in the population of which the batch was a sample, and it is from these quantities that the statistics* of the population may be calculated. In general both P and p will increase steadily with increasing dose (an interesting exception is discussed in \S 41), but, if the number of test subjects in a batch is small, sampling variation may interfere with the regularity of the trend in p. Trevan (1927) has shown that if two batches of five subjects are given doses which would cause 25 % and 75 % of responses respectively in the whole population, only 92 % of trials would give more responses for the higher dose. In 2 % of trials the lower dose would appear to be the more effective, in nearly 6 % the numbers of responses in the two groups would be equal, and in a very small proportion. 0.05 %, either none or all would respond to both doses. The larger the batches the greater is the assurance that there will be satisfactory discrimination between the effects of different doses, but when, as is often the case, the limiting factor to the size of the experiment is the total number of subjects to be used, it is usually preferable to have several batches of moderate size than to have two or three large ones, in order that a wide range of doses may be tested and an idea of the dose-response relationship obtained.

* The word *statistic* is here used in the sense introduced by Fisher (1944, § 11) as 'a value calculated from an observed sample with a view to characterising the population from which it is drawn'.

8. THE MEDIAN EFFECTIVE DOSE

At one time it was customary to characterize the effectiveness of a stimulus by means of the minimal effective dose, or, for a more restricted class of stimuli the minimal lethal dose. terms which fail to take account of the variation in tolerance within a population. Writing of toxicity tests, Trevan (1927) says: 'The common use of this expression in the literature of the subject would logically involve the assumptions that there is a dose, for any given poison, which is only just sufficient to kill all or most of - the animals of a given species, and that doses very little smaller would not kill any animals of that species. Any worker, however, accustomed to estimations of toxicity, knows that these assumptions do not represent the truth.' It might be thought that the minimal lethal dose of a poison could instead be defined as the dose just sufficient to kill a member of the species with the least possible tolerance, and also a maximal non-lethal dose as the dose which will just fail to kill the most resistant member. Though there will undoubtedly be doses so low that no test subject will succumb to them and doses so high as to prove fatal to all, there are considerable difficulties in the way of determining the endpoints of these ranges. Even when the tolerance of individuals can be measured directly, to say, from measurements on a sample of ten or a hundred, that the lowest tolerance found indicated the minimal lethal dose would be unwise; a larger sample might well contain a more extreme member. When only quantal responses for selected doses can be recorded the difficulty is increased, and the occurrence of exceptional individuals in the batches at different dose levels may seriously bias the final estimates. The problem is, in fact, that of determining the dose at which the sigmoid death curve for the whole population meets the zero or 100 % levels of kill and a very extensive experiment would be necessary in order to estimate these points with any accuracy.

As a characteristic of the stimulus which can be more easily determined and interpreted, Trevan has advocated the *median lethal dose*, or as a more general term to include responses other than death, the *median effective dose*. This is defined as the dose

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which will produce a response in half the population, and thus from another point of view, is the mean tolerance. If direct measurement of tolerance were possible the mean tolerance of a batch of test subjects would naturally be considered as the chief characteristic of the dose, and there is a strong case for using an estimate of the same quantity in material of the type now under discussion. The median effective dose may conveniently be referred to as the ED 50, and the more restricted concept of median lethal dose as the LD 50. Analogous symbols may be used for doses effective for other proportions of the population, ED90, for example, being the dose which causes 90 % to respond. As will become apparent in later chapters, by experiment with a fixed number of test subjects, effective doses in the neighbourhood of ED50 can usually be estimated more precisely than those for more extreme percentage levels, and this characteristic is therefore particularly favoured in expressing the effectiveness of the stimulus; its chief disadvantage is that in practice, especially in toxicological work, there is much greater interest attaching to doses producing nearly 100 % responses than to those producing only 50 %, in spite of the difficulty of estimating the former.

For any distribution of tolerances, the ED 50, Λ , satisfies the equation

$$\int_{0}^{A} f(\lambda) d\lambda = 0.5.$$
 (2.5)

When a simple normalizing transformation for the doses is available, so that x, the normalizing measure of dosage, has a normally distributed tolerance, equation $(2 \cdot 1)$ is transformable to

$$dP = \frac{1}{\sigma_{\sqrt{2\pi}}} e^{-\frac{1}{2\sigma^{2}}(x-\mu)^{2}} dx, \qquad (2.6)$$

where μ is the centre of the distribution and σ^2 its variance. Thus μ is the population value of the mean dosage tolerance, or median effective dosage, and efforts must be directed at estimating it from the observational data. This problem will be considered at length in Chapter 3. For the present the normalizing transformation will be assumed logarithmic, as defined by equation (2.3), so that μ is the log ED 50; the results obtained are in the main true

for any other transformation, at least as far as they relate to the measure of dosage, x, but modifications are required in transforming back from the x to the λ scale.

Clearly the ED 50 alone does not fully describe the effectiveness of the poison or other stimulus tested. Two poisons may require the same rates of application in order to be lethal to half the population, but, if the distribution of tolerances has a lesser 'spread' for one than for the other, any increase or decrease from this rate will produce a greater change in mortality for the first than for the second. This 'spread' is measured by the variance, σ^2 : the smaller the value of σ^2 , the greater is the effect on mortality of any change in dose. Stimuli which produce their effects by similar means (in particular, poisons whose physiological effects are similar), often have approximately equal variances of their log tolerances for any given population of test subjects, even though they differ substantially in their median lethal doses. An assessment of the relative potencies can then be made from median lethal doses alone (§ 20).

8.1. NOTATION AND TERMINOLOGY

The symbol λ was used by Gaddum (1933) for 1/b, the estimate of σ given by the reciprocal of the probit regression coefficient (§§ 9, 10). This usage has been followed by other writers, notably Bliss and Cattell (1943). It is perhaps unfortunate that, in the first edition of this book, I chose λ to represent dose; elsewhere, I have used z for dose, and should now prefer that symbol.

In biological assay, the phrase *dose metameter* (Bacharach *et al.* 1942) is generally used to describe the transformed value of dose used in statistical analysis. This is less confusing than *dosage* (\S 5), and should be adopted.

Chapter 3

THE ESTIMATION OF THE MEDIAN EFFECTIVE DOSE

9. THE PROBIT TRANSFORMATION

A TYPICAL test used in the evaluation of an insecticide is one in which successive batches of insects are exposed to different concentrations of the poison for a constant time and, after a suitable interval, scored for the numbers dead and alive. As an alternative to varying the concentration, a fixed concentration may be used throughout, but different total quantities given. Another factor which is sometimes studied at different levels is the period of exposure, the concentration and quantity of poison being kept constant. Such experimental conditions have the character of the stimuli discussed in § 5 whose effects are observed at different levels of intensity. Statistical methods for the analysis of quantal response data have been developed in recent years chiefly for use with tests of this type.

The form of analysis now used to estimate the parameters μ and σ^2 of the distribution of tolerances, equation (2.6), is generally based upon the *probit transformation* of the experimental results. The history of this transformation and of the statistical technique associated with it is outlined in § 14; Bliss (1934b) first proposed the name 'probit' for his modification of Gaddum's normal equivalent deviate, which he increased by 5 so as to simplify the arithmetical procedure by avoiding negative values. The probit of the proportion P is defined as the abscissa which corresponds to a probability P in a normal distribution with mean 5 and variance 1; in symbols, the probit of P is Y, where

$$P = \frac{1}{\sqrt{(2\pi)}} \int_{-\infty}^{Y-5} e^{-\frac{1}{2}u^2} du.$$
 (3.1)

The effect of transformation from percentages or proportions to probits is illustrated in Fig. 5. The normal sigmoid curve of Fig. 4 is reproduced here, together with the straight line obtained when its ordinates are replotted on a linear scale of probits. Along the left-hand vertical axis is a linear scale of percentages with their corresponding probit values, and on the right-hand

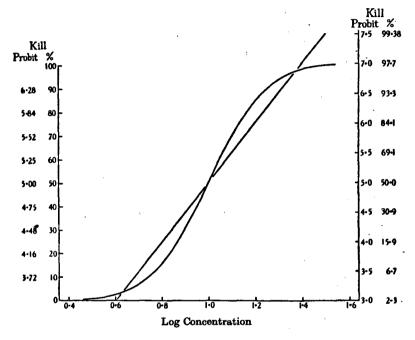


FIG. 5. Effect of the probit transformation. (The normal sigmoid curve of Fig. 4 is transformed to a straight line when the ordinates are measured on a linear scale of probits instead of percentages.)

axis is a linear scale of probits with their corresponding percentage values. The transformation may be considered as a stretching of the left-hand scale to give that on the right-hand, during which process the sigmoid curve becomes straightened.

If $(2\cdot 6)$ represents the distribution of tolerances on the x scale of dosages, the expected proportion of insects killed by a dosage x_0 is

$$P=\frac{1}{\sigma\sqrt{(2\pi)}}\int_{-\infty}^{x_0}e^{-\frac{1}{2\sigma^*}(x-\mu)^*}dx.$$

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Comparison of the two formulae for P then shows that the probit of the expected proportion killed is related to the dosage by the linear equation

$$Y = 5 + \frac{1}{\sigma}(x - \mu). \tag{3.2}$$

By means of the probit transformation, experimental results may be used to give an estimate of this equation, and the parameters of the tolerance distribution may then be estimated; in particular, the median effective dosage is estimated as that value of x which gives Y = 5.

TABLE 1. Transformation of Percentages to Probits

%	0	1	2	3	4	5	6	7	8	9
0		2.67	2.95	3.12	3 ·25	3.36	3.45	3.52	3.59	3.66
10	3.72	3.77	3.82	3.87	3.92	3.96	4.01	4.05	4 ⋅08	4.12
20	4.16	4.19	4.23	4.26	4.29	4.33	4 ·36	4.39	4.42	4.45
30	4.48	4.50	4.53	4.56	4.59	4.61	4.64	4.67	4 ·69	4.72
40	4.75	4.77	4.80	4.82	4.85	4.87	4.90	4.92	4.95	4.97
50	5.00	5.03	5.05	5.08	5.10	5.13	5.15	5.18	5 ·20	5.23
60	5.25	5.28	5.31	5.33	5.36	5.39	5.41	5.44	5.47	5.50
70	5.52	5.55	5.58	5.61	5.64	5.67	5.71	5.74	5.77	5.81
80	5-84	5.88	5.92	5.95	5.99	6.04	6.08	6.13	6.18	6.23
90	6.28	6.34	6.41	6.48	6.55	6.64	6.75	. 6-88	7.05	7.33
	0.0	0.1	0.2	0.3	0.4	0.2	0.6	0.7	0.8	0.9
99	7.33	7.37	7.41	7.46	7.51	7.58	7.65	7.75	7.88	8-09

A table giving probits for specified values of P has been prepared by Bliss (1935*a*), and this table is reproduced by Fisher and Yates (1948) as Table IX of their *Statistical Tables for Biological, Agricultural and Medical Research.* A simplified version of this table, sufficiently detailed for many purposes, is given as Table 1 above, and the full table is reproduced as Table I. The relationship between percentages and probits is shown graphically in Fig. 6.

10. THE PROBIT REGRESSION LINE

When experimental data on the relationship between dose and mortality have been obtained, either a graphical or an arithmetical process can be used to estimate the parameters. Both depend upon the probit transformation. The graphical approach is much more rapid and is sufficiently good for many purposes, but for some more complex problems, or when an accurate assessment of the precision of estimates is wanted, the more detailed arithmetical analysis is necessary. In this chapter only

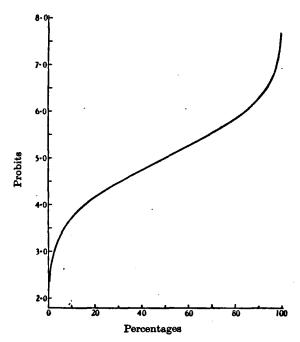


FIG. 6. Relationship between percentages and probits.

the graphical method will be discussed, though the ideas introduced will be wanted again for the discussion of the maximum likelihood estimation of parameters in Chapter 4.

In order to make either type of estimate, the percentage kill observed for each dose must first be calculated and converted to probits by means of Table 1.* The probits are then plotted against x, the logarithm of the dose (or against some other

^{*} There is seldom any advantage in using Table I for these empirical probits; even when batches of test subjects are sufficiently large to justify the use of three or more places of decimals (in itself a rare occurrence), points cannot usually be plotted with this accuracy.

normalizing function of the dose where this seems more suitable), and a straight line drawn by eye to fit the points as satisfactorily as possible. In drawing the line and judging its agreement with the data, only the vertical deviations of the points must be considered: the line must be so placed that the differences between the probit values which are plotted and the probits given by the line at the same dosages are as small as possible. Very extreme probits, say outside the range $2\cdot5-7\cdot5$, carry little weight and may almost be disregarded unless many more insects were used than in the batches giving intermediate probit values. The line is, in statistical terminology, the weighted *regression line* of the mortality probit on x.

This line may be used, as described in § 17, to initiate the arithmetical process of estimating a better fitting line. The empirical probits plotted for a carefully conducted experiment often lie so close to a straight line, however, that there is no necessity to improve on the provisional line. Only experience of the subject and of the experimental technique used can be a sound guide in this matter, but it is undoubtedly true that many experimenters who make use of probit analysis spend time unnecessarily on arithmetic when eye estimation would suffice. As will appear from § 17, the complete method for deriving the best estimate of the line is not difficult, but is laborious if adopted as a routine measure for all tests made.

If it is decided to proceed with the eye estimate alone, the log LD 50 is estimated from the line as m, the dosage at which Y = 5. The slope of the line, b, which is an estimate of $1/\sigma$, is obtained as the increase in Y for a unit increase in x. These two parameters are then substituted in equation (3.2) to give the estimated relationship between dosage and kill. To test whether the line is an adequate representation of the data, a χ^2 test (Fisher, 1944, § 20) may be used, as in Ex. 1 below. A value of χ^2 within the limits of random variation indicates satisfactory agreement between theory (the line) and observation (the data). A significantly large χ^2 may arise either because the individual test subjects do not react independently to the poison, or because the straight line does not adequately describe the relation

between dosage and probit. In the former case the scatter of the points about the line will be wider than would occur if there were no correlation between the reactions of insects in the same batch; the precision of the line will be reduced (§ 11), though its position should be free from bias providing that adequate precautions have been taken in the conduct of the experiment (§ 42). In the latter case there will generally be a systematic departure of the points from the line, indicating a curvilinear relationship; it may be possible to transform this to a linear relationship by adopting a different scale of dosage, such as that of equation $(2\cdot 4)$.

A timely warning against attaching too much importance to the probit itself, at the expense of the kill, has been given by Wadley (Campbell and Moulton, 1943), who says: 'The use of transformations carries with it a temptation to regard the transformed function as the real object of study. The original units should be mentioned in any final statement of results.' In essence the probit is no more than a convenient mathematical device for solving the otherwise intractable equations discussed in Appendix II. Though it may also be used to give a simple diagrammatic representation of the dosage-response relationship, and though familiarity enables these diagrams to be interpreted directly, any suggestion that the statistical analysis is completed by the estimation of a probit regression line must be avoided.

Many of the numerical examples in this book have been chosen to illustrate special points of analytical technique, and, since the data have been removed from their original context, their discussion may not always be carried as far as a statement of conclusions in biological or chemical terms. In practical applications of the probit method the results should finally be expressed by median effective doses, relative potencies, tolerance variances, or other suitable quantities, the units employed being dose (not log dose or dosage) and percentage kill; at this stage the word 'probit' need seldom be mentioned, though sometimes quantities such as mean probit differences (§ 24) may usefully be retained.

Ex. 1. Fitting by eye of a probit regression line to the results of an insecticidal test. Martin (1942, Table 9) has published data

showing the effect of a series of concentrations of rotenone when sprayed on *Macrosiphoniella sanborni*, the chrysanthemum aphis, in batches of about fifty. His results are reproduced in Table 2. The number affected, shown in the third column of the table, is the total of insects apparently dead, moribund, or so badly affected as to be unable to walk more than a few steps. This classification has been found convenient and has been frequently used by Tattersfield and his co-workers at Rothamsted (Tattersfield *et al.*, 1925); the total is taken as the 'kill', and normal or only slightly affected insects are considered to have survived.

Concentra- tion (mg./1.)	No. of insects (n)	No. affected (r)	% kill (p)	Log con- centration (x)	Empirical probit
10.2	50	44	88	1.01	6 ·18
7.7	49	42	86	0.89	6.08
5.1	46	24	52	0.71	5.05
3.8	48	16	33	0.58	4 ·56
2.6	50	6	12	0.41	3 ⋅82
0	49	0	0	-	

TABLE 2.	Toxicity of	Rotenone to	Macrosiphoniella	sanborni
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The rotenone was applied in a medium of 0.5% saponin, containing 5% of alcohol. Insects were examined and classified one day after spraying.

In experiments of this nature, provision should always be made for estimating the natural mortality amongst untreated insects. The last line of Table 2 records that forty-nine insects were sprayed with the alcohol-saponin medium alone, containing no rotenone, and that all of these survived. It therefore seems safe to assume that the results for the five concentrations of rotenone have not been appreciably influenced by the superimposition of a natural mortality of insects. As will be seen later (Chapter 6), adjustments to the statistical analysis are needed when there are indications of an appreciable natural mortality during the course of an experiment.

The percentage kills observed at each concentration are shown in the fourth column of Table 2. These values are estimates of the corresponding averages, P, for the whole population, and are subject to sampling errors. In order to distinguish them from the population values they will be denoted by p. (In all formulae in the text, here and elsewhere, p and P denote proportions, not percentages; it is convenient to use the same symbols in the headings of tables both for proportions and percentages, though for the latter 100p and 100P would be more correct. If this is borne in mind, no confusion should be caused by the slight ambiguity of usage.)

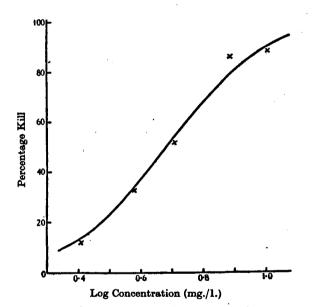


FIG. 7. Relationship between dosage of rotenone and percentage kill of *M. sanborni* (Ex. 1), showing normal sigmoid curve.

Over the range of concentrations tested, the sigmoid nature of the relationship between percentage kill and log concentration is not very apparent. The percentages are plotted against the dosage in Fig. 7, together with the normal sigmoid curve which is fitted to them by the present analysis. Between 25 % and 75 % kill this curve is practically indistinguishable from a straight line; a line drawn to fit the five points would give a dosage of about 0.68 corresponding to 50 % kill, a value in good agreement with

that obtained later for the log LD 50. The straight-line relationship for percentages must nevertheless be quite inadequate at more extreme values, even though the absence of extremes and the regularity of the data enable it to be used here. A method proposed by Kärber (see § 13 below) for the estimation of the LD 50 makes use of this linear approximation.

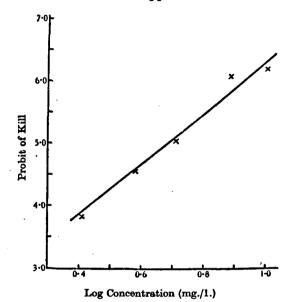


FIG. 8. Relationship between dosage of rotenone and probit of kill of *M. sanborni* (Exs. 1 and 6), showing probit regression line.

The probits of the percentage kills, read from Table 1, have been entered in the last column of Table 2. These probits are plotted against dosage in Fig. 8; they lie nearly on a straight line, and such a line has been drawn through them by eye. Using Table 1 once more, points on the straight line of Fig. 8 have been converted back from probits to percentages; these give the sigmoid curve already shown in Fig. 7, from which the mortality to be expected from any given dosage, or the dosage which will kill, on the average, a given percentage of insects, may be predicted. In practice the sigmoid seldom needs to be constructed, as all predictions can be made directly from the probit diagram. For example, in Fig. 8 a probit value of 5 is given by a dosage of m = 0.687; this therefore is the estimate of log LD 50, and the LD 50 is estimated as a concentration of 4.86 mg./l. The value may be compared with 4.85 mg./l. obtained by the arithmetical process of fitting a straight line to the same data (Ex. 6). Similarly the log LD 90 corresponds to a probit of 6.28 and is therefore 1.003; the LD 90 is thus estimated as 10.1 mg./l.

Again from Fig. 8, an increase of 0.8 in dosage is associated with an increase of 3.21 in the probit.^{*} Hence the estimated

TABLE 3. Comparison of Observed and Expected Mortality inEye Estimation for Rotenone-Macrosiphoniella sanborni Test

Log con-		No		No. a	ffected	Dis-	-
centra- tion (x)	Y	P	No. of insects (n)	Ob- served (r)	Ex- pected (nP)	$\frac{Dis}{(r-nP)}$	$\frac{(r-nP)^{\mathbf{s}}}{nP(1-P)}$
1.01	6.30	90.3	50	44	45.2	-1.2	0.33
0·89 j	5.83	79.7	49	42	39.1	2.9	1.06
0.71	5.10	54.0	46	24	24.8	-0.8	0.06
0.58	4.58	33.7	48	16	16.2	-0.2	0.00
0.41	3.90	13.6	50	6	6.8	-0.8	0-11
				•			$\chi_{[3]}^{2} = 1.56$

regression coefficient of probit on dosage, or the rate of increase of probit value per unit increase in x, is

$$b=1/s=4.01,$$

where s (= 0.25) is an estimate of σ , the standard deviation of the distribution of log tolerances. The relationship between probit and dosage may be written

$$Y = 5 + 4.01(x - 0.687), \text{ or } Y = 2.25 + 4.01x.$$
 (3.3)

Equation $(3\cdot3)$ may be used to calculate expected numbers of insects killed at each concentration. By substitution of the values

^{*} De Beer (1941) has suggested the graduation of a protractor in units of b instead of degrees, so as to read the slope of the line directly; the same instrument could be used for all diagrams having a fixed ratio between the scale units of probits and of dosage.

of x used in the experiment, the equation gives the values of 1 shown in the second column of Table 3; from Table 1 the corre sponding expected percentage kills, P, are obtained. Thus a pro bit of 6.30 corresponds to a percentage of between 90 and 91, or more accurately, $90 + \frac{2}{6} %$. If the expected proportion for an concentration is multiplied by n, the number of insects tested a that concentration, the result is the *expected number* of affected insects, or the average number which would be affected in a batcl of size n if equation (3.3) represented the true relationship be tween dosage and kill. These numbers, nP, may then be com pared with the actual numbers affected, r, in order to judge th adequacy of the equation.

Only the second concentration tested shows any appreciable discrepancy, three more insects being affected than equation (3.3 predicts, and even this may be shown to be well within the limit of random variation. There is no indication of any systemati departure from the line such as might arise if the wrong nor malizing transformation had been used and the true equation were not linear. A test of significance of the discrepancies may be obtained by squaring each, dividing the square by (1-P)and again dividing by the tabulated value nP.* The sum o these quantities is, to a sufficiently close approximation if th line in Fig. 8 has been well drawn, a χ^2 ; the degrees of freedom are two less than the number of concentrations tested, since th two parameters of equation (3.3) have been estimated from the data. The mean value of χ^2 in random sampling from a population whose tolerance distribution is defined by this equation is equa to the number of degrees of freedom; the value obtained in Table 3 is 1.56, which, being less than 3, is clearly sufficiently small to be attributed to random fluctuations about the relation ship specified in (3.3). Indeed Fisher and Yates's Table IV, a which a simplified form is reproduced as Table VI, shows that a value greater than 7.8 could occur by chance in 5% of cases hence the probit regression line in Fig. 8 appears to be a ver satisfactory representation of the results of the experiment.

* This calculation, like that of nP, may be performed on a slide rule with sufficient accuracy for the present purpose.

11. PRECISION OF THE ESTIMATES

The binomial distribution of probabilities (§ 7) shows that if a batch of *n* insects is exposed to a concentration which, on an average, will kill a proportion *P*, the probabilities of *n*, (n-1), ..., 2, 1 being killed are the successive terms in the expansion of $(P+Q)^n$, provided that the *n* insects react independently to the poison. By a well-known result of elementary staticitical theory, it follows that the standard deviation of the period affected insects in samples of size *n* is $\pm \sqrt{(nPQ)}$, and

p is standard deviation of the observed proportion p, about

n value of P, is $\pm \sqrt{(PQ/n)}$. The square of a standard

ion is known as the variance; here the variance of the proportion killed is PQ/n, which is inversely proportional to the number in the batch. The reciprocal of the variance, sometimes called the *invariance* or *quantity of information*, is proportional to n, and represents the weight to be attached to the observation on the batch in respect of the information it provides on P. In Appendix II the weight to be attached to the probit of P is shown to be nw, where

$$w = Z^2/PQ; \tag{3.4}$$

here Z is the ordinate to the normal distribution corresponding to the probability P, and may be written

$$Z = \frac{1}{\sqrt{(2\pi)}} e^{-\frac{1}{4}(Y-5)^2}.$$
 (3.5)

Bliss (1935*a*) and Fisher and Yates (1948, Table XI) have tabulated the *weighting coefficient*, *w*, for values of *Y* at intervals of 0·1; intermediate values may be obtained by interpolation if required.* A shortened version of their table is given here as Table 4; more accurate values appear in the column for C = 0of Table II. The values are symmetrical about Y = 5.0, so that, for example, Y = 3.8 and Y = 10 - 3.8 = 6.2 have the same weighting coefficient, 0.370. For many practical purposes Table 4

* Finney and Stevens (1948) have tabulated w at intervals of 0.01 in Y.

is sufficiently accurate; it is very seldom necessary to interpolate for Y, which may usually be taken correct to the nearest 0.1.

The weighting coefficients may be used in order to estimate the standard error of the log ED 50. For this purpose the value of Y corresponding to each dosage used must be read from the provisional regression line as drawn by eye, the weighting coefficient determined from Table 4 for each Y, and this coefficient multiplied by n, the number of insects tested at the dosage. The quantities nw must then be summed for all dosages; the symbol

TABLE 4. The Weighting Coefficient, $w = Z^2/PQ$

Y	0.0	0-1	0.2	0.3	0-4	0.5	0.6	0.7	0.8	
1	0.001	0.001	0.001	0.002	0.002	0.003	0.005	0.006	0.008	
2	0.015	0.019	0.025	0.031	0.040	0.050	0.062	0.076	0.092	0.110
3	0.131	0.154	0.180	0.208	0.238	0.269	0.302	0.336	0.370	0.405
4	0.439	0.471	0.503	0.532	0.558	0.581	0.601	0.616	0.627	0.634
5	0.637	0.634	0.627	0.616	0.601	0.581	0.558	0.532	0.503	0.471
6	0.439	0.405	0.370	0.336	0.302	0.269	0.238	0.208	0.180	0.154
7	0.131	0.110	0.092	0.076	0.062	0.050	0.040	0.031	0.025	0.019
8	0.012	0.011	0.008	0.006	0.002	0.003	0.002	0.002	0.001	0.001

S is used to denote the summation. If the log ED 50 is not very different from the mean value of the dosages used in the experiment, its standard error is approximately $\pm 1/b \sqrt{(Snw)}$; this expression makes no allowance for the sampling errors in the estimation of b, the slope of the line, in consequence of which it may be a serious underestimation if m, the estimated log ED 50, is far from \bar{x} , the weighted mean dosage or (Snwx)/(Snw). The variance of b is, by equation (II, 11), $1/Snw(x-\bar{x})^2$, and a more accurate value for the variance of m is therefore

$$V(m) = \frac{1}{b^2} \left\{ \frac{1}{Snw} + \frac{(m - \bar{x})^2}{Snw(x - \bar{x})^2} \right\}.$$
 (3.6)

Equation (3.6) gives the variance of the logarithm of any ED value by substitution of its estimate for m. In making a rapid analysis of data, by means of the provisional line only, inclusion of the second term in the variance of m is often unnecessary; its calculation requires part of the complete calculation for fitting a more accurate regression line, and if the computations have to

be taken so far not a great deal of extra labour is needed for the more exact result. Litchfield and Fertig (1941) have described a more rapid, but less exact, approximate method which may sometimes be useful.

This discussion of weights, standard errors and variances is based on the assumption that there is no heterogeneity of departures of the plotted points from the regression line. If the reactions of the individuals in a batch are not independent of one another, the weights, nw, though still proportional to the true weights, will be too large, and the estimated variances will therefore be too small. This will be indicated by a large value of the statistic χ^2 (§ 10), which is now seen to be a weighted sum of squares of the discrepancies between the expected and observed numbers killed. Since the expected value of χ^2 is its number of degrees of freedom, a significantly large value indicates that all weights have been overestimated by a factor $\chi^2/(k-2)$, where k is the number of dosages tested. All variances should therefore be multiplied by this heterogeneity factor as compensation for the overweighting.

When χ^2 is not greatly in excess of its expectation and it may be assumed that the weights are correct without further adjustment, the standard errors may be considered in relation to a normal distribution of errors and fiducial limits calculated with the aid of normal deviates (Ex. 2 and § 12). On the other hand, a significant χ^2 necessitates the estimation of standard errors empirically from the observed variability between batches. and these errors must therefore be used in conjunction with the t-distribution (Fisher, 1944, § 23; Mather, 1943, § 15). The fact that the standard errors themselves are not precisely known leads to a wider range of values being admissible as within the limits of experimental variation for m (Exs. 7 and 9). Fisher and Yates (1948, Table III) have tabulated the distribution of t. the deviate required for this situation, and a simplified version of their table is given as Table VII; t must be used with the same number of degrees of freedom as in the χ^2 . The values rapidly approach those of the normal distribution as the number of degrees of freedom increases.

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Ex. 2. The standard error of a median lethal dose. The foregoing remarks may be illustrated on the data of Ex. 1. In Table 5 the values of Y as predicted by equation $(3\cdot3)$ for each concentration of rotenone are tabulated to the nearest $0\cdot1$. From Table 4 the w corresponding to each Y is entered and multiplied

TABLE 5.	Calculation of	of Standard	Error of	log LD 50	in Eye
Estima	tion for Rotes	none-Macros	iphoniella	sanborni 🕽	Γest

Log con- centration (x)	No. of insects (n)	Y	w	nw	nwx
1.01	50	6.3	0.336	16.8	16-968
0.89	· 49	5.8	0.203	24.6	21.894
0.71	46	5-1	0.634	29.2	20.732
0.58	48	4.6	0.601	28.8	16.704
0.41	50	3.9	0.405	20.2	8.282
				119.6	84.580

$$\tilde{x} = 0.7072,$$
 $(Snwx)^2/Snw = 59.81418,$
 $Snwx^2 = 64.42700,$ $Snw(x-\bar{x})^2 = 4.61282.$

by the corresponding n to give the column nu; again a slide rule gives sufficient accuracy. The sum of this column is

$$Snw = 119.6$$
,

whence the standard error of m is, approximately, since χ^2 was found to be well within the limits of random variation,

$$s_m = 1/4.01 \sqrt{(119.6)}$$

= 0.0228.

The mean dosage, \bar{x} , may be found by constructing a column in Table 5 for *nwx*, summing, and dividing by *Snw*. This gives

$$\overline{x} = 84.580/119.6$$

= 0.707,

۰.

a value which differs but slightly from m = 0.687. In order to calculate the full expression for the variance of m, the entries in the column *nwx* must be multiplied again by x and added to give

$$Snwx^2 = 64.427.$$

PRECISION OF THE ESTIMATES

The sum of squares of deviations of x may then be calculated as

$$Snw(x-\bar{x})^{2} = Snwx^{2} - (Snwx)^{2}/Snw$$

= 64.427 - 59.814
= 4.613,

and therefore, by equation (3.6),

$$V(m) = \frac{1}{b^2} \left\{ \frac{1}{119 \cdot 6} + \frac{(0 \cdot 687 - 0 \cdot 707)^2}{4 \cdot 613} \right\}$$
$$= \frac{0 \cdot 008448}{(4 \cdot 01)^2} = 0 \cdot 000525.$$

The square root of this is the revised value for the standard error of m

 $s_m = 0.0229,$

which differs by a negligible amount from the approximation.

Now m is measured on a logarithmic scale (logarithms to base 10). The standard error of the LD 50, the antilogarithm of m, may be shown to be given by the formula

so that
$$S.E.(10^m) = 10^m \times \log_e 10 \times s_m,$$
 (3.7)
= $0.26.$

Thus the LD 50 is estimated as 4.86 ± 0.26 mg./l. This standard error is of little use, as the distribution of errors may be far from normal on the concentration scale, and tests of significance or fiducial limits (§ 12) should always be evaluated on the dosage scale. For example, if it is desired to test whether the LD 50 estimated for this experiment differs significantly from a true value of 4.2 mg./l., the difference should be examined in logarithmic units. The logarithm of the latter dose is 0.623, so that the estimate is greater by 0.064 ± 0.023 , a deviation which is 2.8 times its standard error. The last line of Table VII, which gives deviates for the normal distribution, shows the probability of so large a deviation to be less than 1 % (more exactly, from Table I, the probability is about 0.5 %); hence the experiment

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has given an estimate of the LD50 significantly greater than $4\cdot 2$ mg./l.

If the standard error of estimation of the log LD 90 is required, the approximate formula breaks down, since the estimate is very different from \bar{x} , and equation (3.6) must be used. The variance is

$$V(\log \text{LD} 90) = \frac{1}{b^2} \left\{ \frac{1}{119 \cdot 6} + \frac{(1 \cdot 003 - 0 \cdot 707)^2}{4 \cdot 613} \right\}$$

= 0.001701,

so that the log LD 90 is estimated as 1.003 ± 0.041 .

12. FIDUCIAL PROBABILITY

When a parameter such as the median lethal dose has been estimated from experimental data it is natural to wish to infer within what limits its true value may reasonably be expected to lie. A statement about the probability of the true value lying between certain limits cannot be made in terms of the ordinary concept of probability, by the use of which probabilities can be assigned only to statements about the occurrence of observations or of statistics calculated from the observations. In order to overcome this difficulty, the concept of *fiducial probability* has been introduced (Fisher, 1942, §§ 62, 63; 1944, § 2; Mather, 1943, § 58).

There is said to be a fiducial probability, F, that the true value of a parameter is greater than a specified value if that value is the lowest value from which the observation is not significantly different at the F level of probability. Similarly, there is said to be a fiducial probability, F, that the true value lies between specified upper and lower limits if the lower limit is the lowest value and the upper limit the highest value which would not be contradicted by a significance test at the $\frac{1}{2}F$ probability level; these are termed *fiducial limits* to the value of the parameters. Their meaning and calculation will be made clear by considering fiducial limits in the estimation of the LD 50 for the data of Ex. 1.

FIDUCIAL PROBABILITY

Ex. 3. Fiducial limits to the LD 50 in the rotenone-Macrosiphoniella sanborni test. In Ex. 2 the $\log LD 50$ for the data on the toxicity of rotenone to *M. sanborni* was estimated as

$$m = 0.687 \pm 0.023.$$

No evidence of significant heterogeneity of the points about the regression line was found, and consequently standard errors may be considered in relation to the normal distribution. From the last line of Table VII, there is a 5 % probability that the deviation from the true log LD 50 shall be at least 1.96 times the standard error, or a $2\frac{1}{2}$ % probability that the estimate shall be at least $1.96s_m$ (= 0.045) less than the true value and a $2\frac{1}{2}$ % probability that the estimaté shall be at least 0.045 greater than the true value. In other words, if the true log LD 50 were 0.732, estimates as low as or lower than 0.687 would occur in only $2\frac{1}{2}$ % of trials such as that under consideration and if the true log LD 50 were 0.642, estimates as high as or higher than 0.687 would occur in only $2\frac{1}{2}$ % of trials. There is therefore a 95% fiducial probability that the true value lies between 0.732 and 0.642, which limits correspond to concentrations of 5.40 and 4.39 mg./l. If the limits had been estimated directly from the LD50 and its standard error as $4.86 \pm 1.96 \times 0.26$ mg./l., or 5.37 and 4.35 mg./l. they would have been equidistant from the estimated LD 50, whereas the limits derived on the logarithmic scale are not symmetrically placed on the concentration scale. The difference is trivial here, but with relatively larger standard errors, or for more extreme limits, it may be considerably greater. The method just discussed is itself an approximation, sufficiently good for many data; the exact procedure for finding fiducial limits is set out later (§ 19).

13. Approximate Methods of Estimation

Before passing to the consideration of the arithmetical technique of fitting probit regression lines, other approximate methods for estimating the LD 50 which have achieved some popularity may be briefly mentioned. All these have been developed independently of the probit technique and are based on an assumed

.linearity of the relationship between the percentage kill and the dosage instead of between the mortality probit and the dosage. They may often be adequate, but they are likely to be misleading if the curvature of the dosage-response curve is appreciable over the range of dosages tested, especially if in addition the distribution of the dosages tested is markedly unsymmetrical about the log LD 50.

(a) Method of extreme lethal dosages. This method is intended for use in the limited class of experiments in which subjects are tested singly, one at each of a series of dosages; the interval, d, between successive dosages must be constant, and enough levels must be tested to cover the range from those practically certain to be ineffective to those practically certain to kill. The log LD 50 is estimated as the mean of the highest dosage which fails to kill and the lowest which kills. When d is of about the same magnitude as σ , the standard deviation in the distribution of log tolerances, this estimate, $m_{(\alpha)}$ has the approximate standard error (Gaddum, 1933)

$$s_{m(a)} = 0.75 \sqrt{(\sigma d)}.$$
 (3.8)

This standard error is of little use, however, as the easiest way of estimating σ is from a probit diagram, and if this is employed the LD 50 may as well be estimated from it.*

(b) Behrens's method is intended for experiments in which the dosages are equally spaced, and in which equal numbers of test subjects are used at each of a series of dosages covering the whole range of kills from zero to 100 %. For each dosage two quantities are calculated:

- $S_{x-}(r)$ = the total number of individuals killed at dosages less than or equal to x,
- $S_{x+}(n-r)$ = the total number of individuals surviving at dosages greater than or equal to x.

The estimate of log LD 50 is $m_{(b)}$, the value of x for which

$$S_{x-}(r) = S_{x+}(n-r),$$
 (3.9)

* Application of the probit method to experiments in which only one individual is tested at each concentration is discussed in § 43.

APPROXIMATE METHODS OF ESTIMATION

and is obtained by interpolation where necessary. The method was suggested by Behrens (1929), from whose data, and from additional results of his own, Gaddum (1933) found an approximation for the standard error of $m_{(b)}$ in the form

$$s_{m(b)} = 0.81 \sqrt{(\sigma d/n)}.$$
 (3.10)

This expression also is of little use, since knowledge of the value of σ can only be derived from a more elaborate analysis.

(c) Kärber's method (Kärber, 1931) is more adaptable than the two preceding approximations, as it does not require a constant interval between successive dosages nor even a constant number of test subjects at each dosage, though the number should not vary very widely. Once again the dosages tested must cover the whole, or practically the whole, range from zero to 100 % kill. The method assumes the whole of the increase in proportion killed between dosages x_i and x_{i+1} , $(p_{i+1} - p_i)$, to be attributable to a dosage $\frac{1}{2}(x_i + x_{i+1})$, and estimates the log LD 50 as the mean dosage. If k dosages are tested, and if $p_1 = 0$, $p_k = 1$ (that is to say the two extreme dosages kill respectively none and all the test organisms), the estimate is

$$m_{(c)} = \frac{1}{2}S(p_{i+1} - p_i)(x_{i+1} + x_i), \qquad (3.11)$$

the summation being taken over all dosages. When the interval between successive dosages has a constant value, d, this estimate may be written more simply as

$$m_{(c)} = x_k - \frac{1}{2} dS(p_i + p_{i+1}). \tag{3.12}$$

The standard error of the estimate in equation (3.12) is

$$s_{m(c)} = d \sqrt{[S(PQ/n)]},$$
 (3.13)

where P, Q are the expected proportions of killed and survivors at each dosage, for which smoothed value of p, q may be taken (Irwin and Cheeseman, 1939*a*). When the extremes of mortality are not reached within the range of dosages tested, formula (3.12) may sometimes be made applicable by the assumption that the next lower or next higher dosage at the end of the range would kill none or all respectively. The standard error for the estimate

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in equation (3.11) may be found similarly, but the more complicated formula will not be given here. Epstein and Churchman (1944) have discussed the theory of Kärber's method, but failure to distinguish between the observed and expected proportions detracts greatly from the usefulness of their work.

Gaddum (1933) has found that an average value of (3.13) may be obtained in the same form as (3.8) and (3.10); the average variance of *m* may be shown to be

$$V(m_{\omega}) = \sigma d/n \sqrt{\pi},$$

$$s_{m_{\omega}} = 0.751 \sqrt{(\sigma d/n)}.$$
(3.14)

and therefore

The variances of these three estimates of the $\log LD50$ may be compared with that of the estimate obtained by the probit method. According to Gaddum (1933), when dosages are equally spaced and cover the whole range from 0 to 100 % kill, average values of the variance are:

Method of extreme lethal dosages:	$0.57\sigma d$,
Behrens's method:	$0.66\sigma d/n$,
Kärber's method:	$0.564\sigma d/n$,
Probit method:	$0.554\sigma d/n$.

These values can be used only if σ either is known or can be assumed the same as in earlier experience of the experimental technique. The indications they give of the relative efficiencies of the four methods, and in particular their suggestion of only a trivial advantage for the probit method over that of Kärber (amounting to a 2% reduction in the variance of the estimated log LD 50), are entirely misleading; as Gaddum has pointed out, these conclusions apply only when the dosages are chosen in such a way as to permit the use of Kärber's method or of one of the other approximations. If previous experience or preliminary trials give any clue to the value of the LD 50, the procedure of distributing test subjects evenly over a wide range of dosages, in order to ensure having zero or complete kills at the extremes, is very wasteful. By concentrating on dosages nearer to the LD 50, observations of much greater weight are obtained and

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a correspondingly more precise estimate of the LD 50 is derived. A further theoretical objection to the three approximate methods is that, unless the dosages used are symmetrically situated with respect to the true log LD 50, each gives a biased estimate. Irwin (1937) found the bias to be negligible, even when the interval between successive dosages was as great as 2σ ; the bias increases with this interval, but intervals greater than 2σ are not likely to occur frequently in practice.

Kärber's method, and to a lesser extent other approximate methods distinct from probit analysis, may occasionally be of use in aiding the rapid estimation of the LD 50, particularly in preliminary or exploratory trials. But the argument usually advanced in their favour, namely that they are much quicker than probit analysis and do not require its complex calculations, is of much less force than might at first appear. The graphical estimation of log LD 50 described in § 10 is as quick as either Kärber's or Behrens's methods, and has the important advantages of being just as easy and satisfactory to use when the dosages are not equally spaced or do not cover the whole range and when the numbers of subjects tested are not equal for all dosages. If assessment of the precision of the estimate of LD 50 is also required, some form of probit analysis is almost essential in order to estimate σ , and equation (3.13) is certainly more troublesome to use than the method given in § 11. By contrast with the other approximate methods, the graphical probit method avoids considerations of whether conditions for the applicability of Kärber's or Behrens's formulae are fulfilled, permits estimation of other properties of the dose-response relationship (such as LD90) as easily as that of LD50, and may easily be converted into the more exact analysis discussed in Chapter 4 when this seems necessary. Furthermore, as will be seen in later chapters, the probit method can be applied to many more complex types of toxicity test data.

Examples of the use of Kärber's method have been given by Irwin and Cheeseman (1939a, b); Ex. 4 below discusses very briefly the application of approximate methods to the data used in Ex. 1.

Ex. 4. Estimation of the median lethal dose by Behrens's and Kärber's methods. In the experiment whose results are summarized in Table 2 the intervals between successive dosages did not differ greatly from the mean of 0.15, and the number of insects tested at each dosage was about 50. If an assumption is made that the next higher dosage would have killed all the insects and the next lower would have killed none, both Behrens's and Kärber's methods can be used for estimating the log LD 50. For the former the percentages dead and alive should be used rather than the actual numbers, in order to overcome the complication of unequal values of n. The two estimates are

$$m_{(b)} = 0.682, \quad m_{(c)} = 0.688.$$

The close agreement with the estimate made graphically in Ex. 1 is largely fortuitous; it occurs because the middle one of the five doses tested is very near to the LD50 and the other doses and percentage kills are practically symmetrical on either side of this. Thus conditions happened to be very favourable to the two approximations, but even so they have no advantage over the graphical method as they can scarcely be reached with greater ease or rapidity than the estimate made in Ex. 1.

14. HISTORY OF THE PROBIT METHOD

Though the widespread use of the probit transformation in the statistical analysis of biological data is of comparatively recent growth, the underlying principle has been known for many years. It appears to have originated with psychophysical investigators who, in the latter half of the nineteenth century, were confronted with the problem of estimating the magnitude of a stimulus from statements by test subjects that it seemed to them greater than or less than the various members of a standard series. The proportion of answers 'greater than' steadily decreases as the scale of standard stimuli is ascended, and shows a sigmoid relationship with the measure of these stimuli.

Fechner (1860) discussed the relationship of the difference between two weights with the proportion of trials in which a subject correctly judged which was the heavier. He suggested (Chapter VIII) the conversion of the proportions to deviates of a normal distribution with mean zero and precision* unity, by means of a table of the normal integral.[†] If steps are taken to eliminate biases due to the order, in time or space, in which the weights are picked up, when the weight difference is negligibly small the proportion of right answers should be one-half, and the normal deviate therefore zero. Fechner further suggested that a linear relationship would be found between the weight difference and the normal deviate. Hence, if the proportion of right answers were known for one weight difference, the factor of proportionality with the normal deviate could be estimated; estimates could then be made of the proportions corresponding to any other weight differences or vice versa. This appears to be the earliest reference to the fundamental idea of the probit method, namely, the reduction of a sigmoid response curve to a straight line by means of a transformation of the responses based on the normal integral; the credit for inventing the method should therefore be given to Fechner.

Müller (1879) recognized that the transformation from proportions to the standardized normal deviates introduced a differential weighting.[‡] He proposed to determine the parameters of the distribution of threshold values by fitting a straight line to the transformed data, weighting each point by its 'Müller weight', a quantity proportional to z^2 in the present notation. The ordinate, z, was taken as corresponding to the observed proportion, and not, as in equation (3.4), to the expected value from the fitted line. His method was called the *Constant Process* or the *Method of Right and Wrong Cases*, under which names it is still known to psychophysicists.

* The precision, $h = 1/\sigma \sqrt{2}$, was at one time commonly used instead of σ as a parameter of the normal distribution.

[†] This table is still known to psychophysicists as 'Fechner's Fundamental Table'.

* There is a danger of some verbal confusion here, since weights are used as stimuli, and, in the statistical analysis, the responses are assigned 'weights' proportional to their invariances.

Urban (1909, 1910) collected extensive experimental data, of the type considered by Fechner, on the difference threshold for lifted weights. In his experiments, a standard weight of 100 g. was lifted before each of a series of seven weights ranging from 84 to 108 g., and a judgement of lighter than, equal to, or heavier than, was given by the subject for each trial. Urban described several different methods for estimating the threshold of just perceptible weight differences, and for assessing the relationship between the weight difference and the proportion of right answers. In his account of the constant process, or, as he termed it, the $\Phi(\gamma)$ Process, he pointed out that the Müller weights failed to take account of the variability in the original proportions due to the binomial distribution of right and wrong answers. To allow for this source of variation, he introduced a factor 1/pq, and weighted his normal deviates proportionally as z^2/pq .

Thomson (1914, 1919) drew attention to certain defects in Urban's treatment of the problem, especially in his methods of estimating the standard errors of the parameters and of assessing the goodness of fit of the line to the data. His revision (Brown and Thomson, 1940, Chapter III) put the constant process into a form very similar to that of the probit method to-day, except that the weights were taken from the values of p for the observations and not from the corresponding values of P for a provisional line, and that his formulae for the standard errors of the parameters were much more complicated and laborious to compute than those now used. No provision was made for taking account of zero or 100% values of P, which were presumably simply ignored.

Independently of the work of the psychophysicists, Hazen (1914) and Whipple (1916) suggested the use of graph paper the scale of ordinates of which is graduated according to a normal probability distribution, so that the proportions are plotted as their corresponding normal deviates. A normal sigmoid curve plotted on this paper is automatically transformed to a straight line. A modified form of paper has a logarithmic scale of abscissae, so that a logarithmic transformation of dose is also made automatically at the same time.* O'Kane *et al.* (1930) made use of probability paper for plotting the results of insecticide tests.

In 1923 Shackell suggested that the normal integral might be used in interpreting the results of toxicity tests. Three years later Wright (1926), also without knowledge of the earlier work, proposed the use of an inverse function of the normal probability integral as a means of simplifying the statistical treatment of certain statistical data. His paper, however, seemed to escape the attention of biologists, who therefore had to wait a further seven years for yet another rediscovery of the method.

In 1933 Gaddum published an important memorandum on the analysis of quantal assay data in biological investigations. He proposed to transform each percentage to its normal equivalent deviation (N.E.D.), defined as the abscissa to a normal curve of zero mean and unit variance corresponding to a probability P; that is to say, P is the probability of obtaining an observation from this normal distribution whose value is less than or equal to the N.E.D. In symbols

$$P=\frac{1}{\sqrt{(2\pi)}}\int_{-\infty}^{N.E.D.}e^{-\frac{1}{2}u^2}du,$$

a transformation essentially the same as that used by Fechner. The N.E.D. of the percentage kill of various animals was found by Gaddum to give a straight line when plotted against the log dose of the drug applied. He described the regression technique for fitting the line, in a form similar to that given earlier by Urban and Thomson, but his treatment of the standard errors of the parameters and associated quantities was much simpler.

Bliss (1934*a*) suggested the division of the interval between 0.01 and 99.99 % into units of normal deviation which he called *probits*, the whole interval ranging from 0 to 10 probits and 50 % being 5 probits. When he later saw Gaddum's publication, he modified his definition of the probit and redefined it as in equation (3.1), so that it became the N.E.D. increased by 5 (Bliss, 1934*b*). In two comprehensive papers (1935*a*, *b*) he discussed the use of the probit transformation, and gave tables of probits

^{*} Arithmetic and logarithmic probability paper can be obtained from the Codex Book Company, Inc., 74 Broadway, Norwood, Mass., U.S.A., or from Wightman, Mountain and Co., London, S.W.1.

and weighting coefficients. Fisher (1935) outlined the maximum likelihood analysis for zero and 100 % responses, and showed the need for using 'working probits' at all levels of mortality, in the manner described by Bliss (1938).

Two other developments deserve notice. In 1894, Henry showed how to transform a cumulative normal curve to a straight line by use of normal equivalent deviates, and suggested graph paper with probability ruling. Haag (1926) gave examples of Henry's scheme, but, like Henry, discussed only data for which direct measurements were available and not the essentially quantal type. The work of Kapteyn (1903) and Kapteyn and Van Uven (1916) owed something to Fechner; their reaction curves are aids to the normalization of distributions, but they also were not concerned with essentially quantal data.

Meanwhile, psychologists apparently remained unaware that their method had been adopted and refined by biologists. Ferguson (1942) used the constant process for analysing data on the selection of items for mental tests; Lawley (1943, 1944), in considering the theory of this problem, came near to an independent derivation of the maximum likelihood solution. Finney (1944c) endeavoured to unite the biological and psychophysical methods by illustrating the use of the probit method on Ferguson's data.

15. ALTERNATIVE TOLERANCE DISTRIBUTIONS

If log tolerances are not normally distributed, the relationship between log dose and response rate will not be a normal sigmoid. Urban (1910) proposed the alternative

$$P = \frac{1}{2} + \frac{1}{\pi} \tan^{-1} (\alpha + \beta x), \qquad (3.15)$$

where P, x are as before and α , β are parameters. Wilson and Worcester (1943c) realized that this was one of a class of sigmoids; their theory has been extended by Finney (1947*a*, 1949*d*), using the method of Appendix II with an equivalent deviate

ALTERNATIVE RESPONSE CURVES

appropriate to the curve adopted. Of particular interest are the logistic curve,

$$P = 1/\{1 + e^{-(\alpha + \beta x)}\}, \qquad (3.16)$$

and the sine curve, which applies only to a finite range of doses,

$$P = \frac{1}{2} \{ 1 + \sin (\alpha + \beta x) \} - \frac{\pi}{2} \le \alpha + \beta x \le \frac{\pi}{2}.$$
 (3.17)

Equation (3.16) is frequently used to represent a growth curve. Its use for quantal responses was suggested by Fisher and Yates in the introduction to their *Statistical Tables*, and the estimation technique has been discussed by others (Wilson and Worcester, 1943*a*, *b*, *d*; Worcester and Wilson, 1943; Berkson, 1944, 1946). Berkson defined the *logit*, analogous to the probit (cf. Finney, 1947*b*), and showed that an ingenious approximation to the minimum χ^2 (instead of maximum likelihood) estimation of the parameters simplified the calculations. Knudsen, and Curtis (1947) commended the sine curve; its angle transformation has a constant weighting coefficient, so that the arithmetic is less laborious than for probits or logits.

The logit and angle transformations seem to have less theoretical recommendation than the probit; however inexact the assumption of a normal distribution of log tolerances may be, there is no reason to suppose that the tolerance distributions to which they correspond are any closer to the truth. Nevertheless, only very numerous data could discriminate between the three, as even the limited distribution that gives the angle transformation approximates to the normal except at extreme values. In practice, conclusions drawn from the same data by each of the three methods will often agree sufficiently for the choice to be unimportant. Consequently, for simple routine analyses, there may be a good case for using angles instead of probits. For more complex problems, the advantage for angles is less and probits are preferable. The logistic curve might be more appropriate than the normal in Ex. 33.

Chapter 4

THE MAXIMUM LIKELIHOOD SOLUTION

16. WORKING PROBITS

THOUGH the methods described in Chapter 3 are often adequate for estimating the LD 50, the variance of tolerances, or other quantities connected with the dosage-response relationship, complications in the data may prevent these being used satisfactorily and an arithmetical rather than a graphical technique for obtaining the best fitting probit regression line is then required. For example, the observations, when plotted as in Fig. 8, may be too irregular for any confidence to be placed in an eye estimation of the fitted line, or differences in the weights attached to the observations may be so marked as to make satisfactory allowance for them very difficult without objective computational methods. The need becomes still greater when several different materials or dose factors are simultaneously under test; in the more complex types of experiment discussed in Chapters 5–8 developments of the analytical technique now to be discussed are almost essential.

The mathematical basis of the method of estimating the probit regression equation $(3\cdot 2)$ by a process of successive approximations is given in Appendix II, but its details need not concern the non-mathematical reader. The process is begun by drawing a provisional probit line, just as described in § 10, and using this to determine the weights, nw, to be attached to each observation. The weighted regression equation of probit mortality on dosage is then computed. The transformation from observed proportions to probits, as defined by equation $(3\cdot 1)$, is non-linear, and, as shown in Appendix II, the regression equation should therefore be calculated for working probits rather than for the empirical

* Some writers use the term 'corrected probit' instead of working probit, but this name is an unfortunate choice; the working probit is used to give a convenient simplification of otherwise complicated calculations and is in no way a correction of the empirical value. On diagrams of probit regression lines only the empirical probits should be shown, the occurrence of zero and 100 % kills being indicated by arrows, as in Fig. 9. probits obtained directly from the proportions p. The working probit is defined in equation (II, 15) as

$$y = Y + \frac{p - P}{Z}$$
$$= Y - \frac{q - Q}{Z},$$

where Y is the *expected probit* taken from the provisional line, P and Z the corresponding probability and ordinaté respectively. Either of these formulae may be used, according to convenience. The quantity

$$y_{100} = Y + \frac{Q}{Z}$$

is known as the maximum working probit; when all the test subjects in a batch are killed, p = 1 and the corresponding empirical probit is infinite, but the maximum working probit determined from the provisional line allows this observation to play its part in determining the regression equation. Similarly

$$y_0 = Y - \frac{P}{Z}$$

is the minimum working probit, and is used when no subjects are killed. Fisher and Yates (1948, Table XI) have tabulated the maximum working probit and also the range, 1/Z, for values of Y at intervals of 0.1. Table III of this book shows both minimum and maximum working probits and the range; the second column of this table gives minimum working probits for $Y = 1\cdot1-6\cdot5$, and the fourth column gives maximum working probits for $Y = 3\cdot5-8\cdot9$, values outside these limits being of exceedingly rare occurrence. The difference between the maximum and minimum working probits for the same Y is the range, so that when only one is tabulated the other may easily be derived.

From Table III working probits for other values of p may be calculated as in Ex. 5 below. Table IV has been prepared in this way, so as to give working probits at intervals of 0.1 in Yand 1% in kill for all combinations of values whose working probits lie between 0 and 10, outside which limits working probits seldom occur.* The working probit usually does not differ from the empirical probit by as much as does the expected probit, Y.

Ex. 5. Calculation of working probits. As an example of the use of these tables, consider the working probit corresponding to a kill of 72.3% when the expected probit, as taken from a provisional line, is 6.2. From Table III the minimum working probit is 1.6429 and the range 5.1497. Hence p = 0.723 gives

$$y = 1.6429 + 0.723 \times 5.1497$$

= 5.366.

Alternatively, using the maximum working probit, 6:7926, and q = 0.277 $y = 6.7926 - 0.277 \times 5.1497$

The same result may be obtained more easily from Table IV. In this table the column for Y = 6.2 shows working probits of 5.351 and 5.402 for 72 and 73 % kill respectively: hence, by an interpolation which may be carried out mentally,

$$y = 5 \cdot 351 + \frac{3}{10} (5 \cdot 402 - 5 \cdot 351)$$

= 5 \cdot 366.

Very frequently y is required only to two places of decimals, or p is reliable only to the nearest 1 %, so that y can be read directly from Table IV with little or no interpolation.

Occasionally the expected probits may need to be taken to two decimal places. Interpolation is then necessary in order to obtain the maximum or minimum working probit and the range, after which y may be calculated as above. Thus if the value of Yin this example were 6.24 the maximum working probit and the

^{*} A useful table of working probits, at intervals of 0.1 in Y and including every value of p that can arise in a batch of twenty or less test subjects, can be obtained from the Division of Pharmacology, Food and Drug Administration, United States Department of Agriculture, Washington, D.C. Finney and Stevens (1948) have tabulated maximum and minimum working probits and ranges at intervals of 0.01 in Y.

range would be given by taking the sum of 6/10 of the values for Y = 6.2 and 4/10 of the values for Y = 6.3. Hence y may be written, in a form suitable for machine calculation with the minimum of resetting, as

$$y = 0.6 \times 6.7926 + 0.4 \times 6.8649$$

-0.277 × (0.6 × 5.1497 + 0.4 × 5.8354)
= 5.319.

The correct value, obtained from Stevens's more detailed table, is 5.323.

17. THE REGRESSION EQUATION

The linear regression equation of the working probits on the measure of dosage, x, is an improved estimate of the dosageresponse relationship. If it differs markedly from the provisional line drawn by eye, it may itself be used as a new provisional line and the process repeated. The maximum likelihood estimate is, in fact, the limit to which these estimates tend as the cycle of determining a new line with the aid of that last calculated is indefinitely repeated. With experience the first provisional line may often be drawn so accurately that only one cycle of the calculations is needed to give a satisfactory fit, though when the empirical probits are very irregular two cycles may be needed.

De Beer (1945) has developed an ingenious system of scales and nomographs for simplifying the calculations. With the aid of these, many of the expressions required as steps in the estimation of the LD 50 and its fiducial limits can be read directly from the diagram showing the provisional regression line. The results obtained are not quite the same as the maximum likelihood values, since the method does not completely distinguish between empirical and working probits, nor does it obtain fiducial limits from the exact formula (4.6) below; nevertheless it is likely to be sufficiently good for many purposes, and merits serious consideration by those who have to make many routine LD 50 estimations but who find a purely graphical method insufficiently accurate.

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Ex. 6. Arithmetical procedure in the fitting of a probit regression line. As a simple example of the computations required in the fitting of a probit regression line by the maximum likelihood method, the data of Ex. 1 will be used again. The computations are shown in Table 6. A more detailed account of computing procedure is given in Appendix I, where another set of data is used.

X	n	T	р (%)	Em- pirical probit	Y	nw	y	nwx	nwy
1.01	50	44	88	6.18	6.3	16.8	6.16	16.968	103-488
0.89	49	42	86	6.08	5.8	24.6	6.05	21.894	148.830
0.71	46	24	52	5.05	5.1	29.2	5.05	20.732	147.460
0.28	48	16	33	4.56	4 ·6	28.8	4.56	16.704	131-328
0.41	50	6	12	3.82	3.9	20-2	3.83	8 ·282	77-366
						119-6		84-580	608-472
		1/Sn	w = 0	0083612	04, z	= 0.7072	, ÿ = l	5·0876.	
	S	nwx		S	nwxy		Sn	wy¹	
	64	·42700)	44	9-5685		3177-	748	
	59	·81418	3 , '	43	0-3057		3095	637	
	4	-61282	2	1	9-2628		82	111	
							80	440	
							1.	$671 = \chi^2_{(3)}$	
			ь	= 4 ·1759	,				
			V	=5.0876	+4.17	59 /2 - 0.	7079\		

TABLE 6. Maximum Likelihood Computations for Rotenone-Macrosiphoniella sanborni Test

=2.134 + 4.176x.

The first five columns of Table 6 are repeated from Table 2. The expected probits, Y, are obtained from the provisional line of Fig. 8, and the corresponding weighting coefficients (from Table 4) are then multiplied by n and tabulated, this step being exactly as in Ex. 2. Working probits are read from Table IV, and entered in the column y. In the example the empirical probits and working probits are so nearly the same that the former could have been used instead of y without appreciably altering the conclusions, but a routine of working with y is to be preferred. Consideration of the weights, nw, indicates that two decimal places in the working probits are quite sufficient; the variances of the individual observations are 1/nw, and thus the standard errors are of the order of 0.2, so that differences of 0.01 in working probits are of little importance.

Individual values of the products nwx, nwy, are next tabulated and added; division of the totals by Snw gives the means \bar{x} , \bar{y} . The values of nwx are multiplied by x and added to give $Snwx^2$, then multiplied by y and added to give Snwxy. The products of nwy with x should be summed as a check on Snwxy, and also, for reasons which will shortly appear, the products of Snwywith y to give $Snwy^2$. The three totals are reduced respectively by $(Snwx)^2/Snw$, $(Snwx) \times (Snwy)/Snw$ and $(Snwy)^2/Snw$, to give the sums of squares and products about the mean, $Snw(x-\bar{x})^2$, $Snw(x-\bar{x})(y-\bar{y})$ and $Snw(y-\bar{y})^2$.

From equation (II, 9) the estimated regression coefficient is

$$b = Snw(x - \bar{x}) (y - \bar{y})/Snw(x - \bar{x})^{2}$$

= 19.2628/4.61282
= 4.1759.

and the fitted equation is a line with this slope passing through the point (\bar{x}, \bar{y}) :

$$Y = 5.0876 + 4.1759(x - 0.7072)$$

= 2.134 + 4.176x. (4.1)

Using this equation, Table 7 has been obtained in the same way as was Table 3 and gives an heterogeneity χ^2 of 1.62 with 3 degrees of freedom. Since

$$\chi^{2} = S \frac{n(p-P)^{2}}{PQ}$$
$$= Snw \left(\frac{p-P}{Z}\right)^{2},$$

the computation may be made more easily as the weighted sum of squares of deviations of the working probits from the provisional line, $Snw(y-Y)^2$. This quantity is $Snw(y-\bar{y})^2$ reduced by $\{Snw(x-\bar{x})(y-\bar{y})\}^2/Snw(x-\bar{x})^2$; the calculation is shown in Table 6, and gives the result

$$\chi_{[3]}^2 = 1.67.$$

Neither this nor the value 1.62 in Table 7 is strictly a χ^2 , in the recognized meaning of the term, until the maximum likelihood solution has been closely approximated by a series of cycles of computation; in the limit, the two values would be the same, and, in practice, reference of either to the χ^2 distribution at an earlier stage is usually harmless.

TABLE 7. Comparison of Observed and Expected Mortality in Maximum Likelihood Fitting for Rotenone-Macrosiphoniella sanborni Test.

Log				No. a	ffected		
concen- tration (x)	Y	P	No. of insects (n)	Ob- served (r)	Ex- pected (nP)	Dis- crepancy (r-nP)	$\frac{(r-nP)^2}{nP(1-P)}$
1.01	6.352	0.9118	50	44	45.59	- 1.59	0.63
0.89	5.851	0.8026	.49	42	39.33	2.67	0.92
0·71	5.099	0.5398	46	24	24.83	-0.83	0.06
0.58	4.556	0.3285	48	16	15.77	0.23	0.00
0.41	3·846	0-1242	50	6	•21	-0.21	0.01
	· · · - · ·	·	•				$\binom{2}{[3]} = 1.62$

The χ^2 test gives no evidence of heterogeneity of departure from the fitted probit line. The variances of the parameters are therefore, from equations (II. 10, 11),

 $V(\bar{y}) = 1/Snw = 0.00836, V(b) = 1/Snw(x-\bar{x})^2 = 0.2168,$

whence $\bar{y} = 5.088 \pm 0.091$, $b = 4.176 \pm 0.466$.

The slope has been altered from its provisional value of 4.01 by an amount equal to about one-third of its standard error,

* The χ^2 calculated as here does not necessarily decrease as the maximum likelihood solution is approached; the process of maximizing the likelihood function is not precisely equivalent to that of minimizing χ^2 , though the final χ^2 value will usually be not very different from the minimum.

and if an accurate value of b were particularly required a further cycle of computations would be desirable; the next value obtained for b is, in fact, 4.196, the alteration being only 4% of the standard error.

On the scale of Fig. 8 the probit equation $(4\cdot 1)$ is practically indistinguishable from the eye estimate (equation $(3\cdot 3)$). The estimated log LD 50 is

$$m = \overline{x} + \frac{5 - \overline{y}}{b}$$
$$= 0.7072 - 0.0210$$
$$= 0.686;$$

the variance, obtained from equation (3.6), is

 $s_{m} = 0.0220.$

$$V(m) = \frac{0.008457}{(4.176)^2} = 0.0004849,$$

and therefore

The estimate of the log LD 50 obtained in Exs. 1 and 2 (0.687 \pm 0.023) agrees remarkably well with the value 0.686 \pm 0.022 found by the maximum likelihood method. The LD 50 is now estimated to be 4.85 ± 0.25 mg./l.

18. HETEROGENEITY

The data of Ex. 6 were selected as being particularly regular and presenting no complications in their analysis. The χ^2 test for the heterogeneity of the discrepancies between observed and expected numbers is only valid when the expected numbers are not 'small'. At the more extreme dosages tested either P or Qis often nearly zero, so that, with the usual numbers of insects exposed to the poison, either the expected number killed (nP)or the expected number surviving (nQ) is too small for χ^2 calculated in the usual manner to be referred to the distribution in Table VI. The formulae used for χ^2 in Ex. 6 may be written

$$\chi^2_{[k-2]} = S_{yy} - S^2_{xy} / S_{xx}, \qquad (4.2)$$

where the abbreviations S_{xx} , S_{xy} and S_{yy} are used for the sums of squares and products of deviations about means $Snw(x-\bar{x})^2$, $Snw(x-\bar{x})(y-\bar{y})$ and $Snw(y-\bar{y})^2$, and k is the number of dosages

tested.* Small values of nP or nQ often lead to unduly large contributions to χ^2 , thus exaggerating the significance of the total.

In other applications of the χ^2 test, a value less than 5 for the expected number in any class has often been taken as a warning that the χ^2 distribution may give misleading results (Fisher, 1944, Ex. 9). There is no special virtue in the number 5; in some circumstances considerably lower expectations produce no ill effects, and in others the χ^2 distribution may be unreliable with higher expectations (Cochran, 1942). Possibly the chief danger lies in the application of the test to data in which the expectations for most classes are moderately large but one or two are very small; when all or nearly all expectations are low the disturbance of the distribution is not likely to be so serious. Further investigation into this subject is needed, and at present no more definite advice can be given than to treat as suspect any χ^2 value most of which is made up of large contributions from classes with small expectations.

No. in		Expected no.	of survivors (nP)
batch (n)	10	5	2	1
- 5			5-25	5.84
10		5.00	5.84	6-28
20	5.00	5.67	6-28	6.64
30	5.43	5.97	6.50	6-83
40	5.67	6.15	6.64	6-96
50	5.84	6.28	6.75	7.05
100	6.28	6.64	7-05	7.33
200	6.64	6-96	7.33	7.58
1000	7.33	7.58	7.88	8.09

TABLE 8. Table of Greatest Expected Probit giving atleast nP Expected Survivors in a Batch of n

The difficulty of small expectations may usually be overcome by combining the results for extreme dosages with the next highest or next lowest, so as to build up larger expectations,

* A number of writers have used A, B and C for S_{xx} , $S_{\lambda y}$ and S_{yy} , but the latter symbols have the advantage of giving clearer indication of their meaning.

though some sensitivity is thereby lost. Equation (4.2) cannot then be used for the calculation of χ^2 , and recourse must be had to the method of Ex. 1 involving the calculation of the various expectations. The degrees of freedom for χ^2 must be reduced by the number of dosage levels lost by combination (Ex. 7). Table 8 gives a rapid guide to the 'danger levels' of the expected numbers of survivors. The table states, for example, that the expected number of survivors in a batch of fifty insects will be less than 5 when Y exceeds 6.28 and will be less than 2 when Y exceeds 6.75. Similarly, the expected number killed will be less than 5 when Y is less than 3.72 (= 10 - 6.28), less than 2 when Y is less than 3.25. When the batches of insects or other test organisms are themselves small (say ten or less) it may be safe to adopt a requirement of at least 2 survivors and 2 killed in each group, but for batches of thirty and upwards a standard of about 5 seems a better working rule.

Ex. 7. Application of probit analysis to heterogeneous data. As a second example of the technique of fitting the probit regression line, this time to less regular data, results obtained by Busvine (1938) on the toxicity of ethylene oxide to the grain beetle, *Calandra granaria*, may be considered. These data have been fully reported by Bliss (1940b, Table VII); for the present purpose only the records referring to insects examined 1 hr. after exposure to the poison will be used.

The data are shown in the first three columns of Table 9, x being the logarithm to base 10 of the concentration of ethylene oxide in mg./100 ml. The empirical probits were plotted against x (Fig. 9) and a provisional line

$$Y = 3.06 + 7.95x$$

was drawn by eye, allowing for a value above the line at x = 0.391and a value below the line at x = 0.033. Using this line, by application of the maximum likelihood process, the new approximation

$$Y = 2.948 + 8.600x \tag{4.3}$$

was reached. The change in the value of the regression coefficient was sufficiently large for a second cycle of computations to seem worth while, details of which are shown in Table 9. The values of Y are obtained from equation (4.3), the working probits from Table IV; maximum and minimum working probits respectively are required at x = 0.391 and x = 0.033, and these are read either from the 100 and 0 % lines of Table IV or directly from Table III. The computations then proceed as in Ex. 6 to give the equation $V = 2.028 + 9.674\pi$ (4.4)

$$Y = 2.928 + 8.674x. \tag{4.4}$$

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 $32.421 = \chi^2_{(8)}$

As will be seen shortly, differences between $(4\cdot3)$ and $(4\cdot4)$ are negligible by comparison with the standard errors, and no further approximation is necessary.

 TABLE 9. Maximum Likelihood Computations for Ethylene

 Oxide-Calandra granaria Test (second cycle)

					,				
x	n	r	p (%)	Em- pirical probit	Y	nw	у	nwx	nwy
0.394	30	23	77	5.74	6.3	10.1	5.52	3.9794	55.752
0.391	30	30	100	00	6.3	10.1	6.86	3.9491	69-286
0.362	31	29	94	6.55	6.1	12.5	6.45	4.5250	80.625
0.322	30	22	73	5.61	5.7	15.9	5.61	5.1198	89.199
0.314	26	23	88	6.18	5.6	14.5	6.06	4.5530	87.870
0.260	27	7	26	4.36	5.2	16.9	4.38	4.3940	74.022
0.225	31	12	39	4.72	4.9	19.7	4.72	4.4325	92.984
0.199	30	17	57	5.18	4.7	18.5	5.19	3.6815	96.015
0.167	31	10	.32	4.53	4.4	17.3	4.54	2.8891	78.542
0.033	24	Ō	Ō	- 00	3.2	4.3	2.74	0.1419	11.782
						139-8		37.6653	736-077
		1/S1	w = 0	00715307	6, x =	= 0·269 4	$, \overline{y} = 5$	·2652.	
	S	nwx ²		St	wxy		Snu	ry²	
	11	·1877	8	207	.3360		3 986·	265	
	10	·1478	9	198	-3159		3 875·	60 3	
	1	.0398	9	9	0201		110.	662	

 $b = 8.6741, \quad Y = 2.928 + 8.674x.$

Application of formula (4.2) gives a χ^2 of 32.42, with 8 degrees of freedom, as a measure of heterogeneity, a value which, according to Table VI, is clearly significant. The validity of this test is suspect, however, since three of the expected probits exceed 6.0 and one is less than 4.0, thus giving dangerously low values to nP or nQ. The test may be modified as shown in Table 10 by grouping the expectations for the two highest concentrations with the next highest and that for the lowest with the next lowest. Equation (4.2) is then no longer applicable, and the separate contributions to χ^2 must be calculated by the

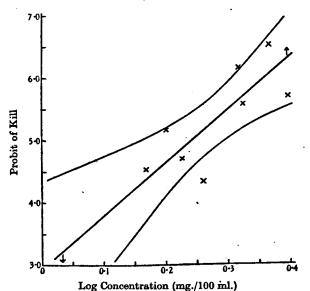


FIG. 9. Probit regression line and 5% fiducial band for toxicity of ethylene oxide to C. granaria (Exs. 7-9).

method used in Ex. 1, namely, by squaring the difference between observed and expected numbers killed, multiplying by the total number, and dividing by the product of the expected kill and expected survivors. Thus

$$\frac{91 \times (0.9)^2}{81 \cdot 1 \times 9.9} = 0.09$$

is the contribution from the first group. The total number of groups of insects has been reduced by 3 and therefore χ^2 has 5 degrees of freedom instead of 8;

$$\chi^2_{[5]} = 19.74$$

is still highly significant, and confirms the evidence for heterogeneity of the departures from the regression line. Since Fig. 9

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gives no suggestion of any systematic deviations from the line such as would arise if the regression relationship were not truly linear, the heterogeneity may be allowed for by multiplying all variances by the heterogeneity factor (§ 11)

	$\chi^2/5=3.95.$
For example	V(b) = 3.95/1.040 = 3.798,
and therefore	$b=8.674\pm1.949.$

 TABLE 10.
 Comparison of Observed and Expected Mortalities for Ethylene Oxide-Calandra granaria Test

1	
0.9	0.09
	1.
-1.0	0.19
3.7	2.75
-8.5	10.94
-2.0	0.52
6.0	5.17
	1
0.8	0-08

Again, from (4.4) the log LD 50 is

$$m = 0.239,$$

and using equation (3.6) once more

$$V(m) = \frac{3.95}{(8.674)^2} \left[0.007153 + \frac{(0.030)^2}{1.040} \right]$$

= 0.000421,

so that the standard error of m is ± 0.021 .

If 5% fiducial limits to *m* are wanted, instead of using the normal deviate, 1.96, as the multiplier of s_m , the *t*-value (Table VII) for 5 degrees of freedom, 2.57, must be used. Hence these limits are 0.239 ± 0.054 , or 0.293 and 0.185; the LD50 is estimated as 17.3 mg./l. with 19.6 and 15.3 mg./l. as the 5% fiducial limits. It will be seen later (Ex. 9) that these limits are in fact not sufficiently wide, and in this example a more exact method of estimating them is needed.

When very few degrees of freedom are available for estimating the heterogeneity factor, the corresponding t-value for any selected probability level becomes large in order to allow for the unreliability of the heterogeneity factor, and consequently the fiducial limits are widely spaced. For example, for a large number of degrees of freedom the 5% t is about 2.00, for 5 degrees of freedom it is 2.57, for 3 degrees of freedom 3.18, and for 1 degree of freedom 12.71! This last value, in particular, is practically useless in the determination of fiducial limits. The remedy is to pool the χ^2 values from comparable series of tests, whenever this is possible, and thus to obtain a single heterogeneity factor of reasonable accuracy instead of separate factors of low precision. In particular, when a single experiment consists of several series of tests which yield similar values of b, the methods of analysis described and illustrated in § 20 should always be used; unless there are strong indications that the series differ in respect of heterogeneity, a composite test of heterogeneity is then made for the whole experiment and, if necessary, a single heterogeneity factor estimated and used for all standard errors and fiducial limits.

19. FIDUCIAL LIMITS

The expected probit, Y, for any dosage, x, has been obtained in the form

$$Y=\bar{y}+b(x-\bar{x});$$

the variance of Y is

$$V(Y) = \frac{1}{Snw} + \frac{(x - \bar{x})^2}{S_{xx}},$$
 (4.5)

if no allowance has to be made for heterogeneity, but the expression must be multiplied by the heterogeneity factor when this is significantly greater than unity. Fiducial limits to Y are therefore $Y \pm s_F t$, where s_F is the square root of V(Y) and t is the normal deviate for the level of probability to be used, or,

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if there is significant heterogeneity, the t-value corresponding to this probability.

If the fiducial limits of Y are plotted for each x, they will be found to lie on two curves which are convex to the regression line and which approach the line most closely at the dosage \bar{x} . The further x is removed from \bar{x} in either direction the greater is the contribution to the variance of Y from the second term of (4.5), which represents the effect of the errors of estimation of the regression coefficient b, and in consequence the more widely spaced are the fiducial limits.

In insecticidal and fungicidal investigations the effect of a single poison is often of less interest than comparisons between the effects of two or more different poisons in the same experiment. Day to day changes in the susceptibility of the organisms may alter very considerably the kill for any selected dosage without seriously upsetting the relative toxic effects of different poisons; consequently the fiducial band for one regression line may not be very helpful for inferring future behaviour (§ 20).

Ex. 8. Fiducial bands for a probit regression line. The plotting of fiducial limits so as to give a band on either side of the probit regression line may be illustrated on the data of Ex. 7. Under the conditions of the experiment, the true value of the kill for the range of dosages may be expected to lie within this band with a degree of confidence represented by the fiducial probability level chosen.

Formula (4.5) gives for these data

$$V(Y) = 3.95 \times \left\{ 0.007153 + \frac{(x - 0.269)^2}{1.040} \right\};$$

variances and standard errors calculated from this expression are shown in Table 11. Multiplication of the standard error, s_T , by 2.57, the appropriate *t*-value for a 5% fiducial probability, gives the width of the fiducial band on either side of the regression line; the boundaries are shown in Fig. 9.

The method of determining fiducial limits which has been described in § 12 (Ex. 3) is often sufficiently good, both for the $\log LD 50$ and for the estimated dosage corresponding to any

other selected kill; strictly speaking, however, the limits are the values of x for which the boundaries of the fiducial band attain the selected value of Y. By solving an equation so as to obtain the value of x for which Y has a selected fiducial limit, exact fiducial limits to x, the dosage giving a kill whose probit is Y, are shown to be

$$x + \frac{g}{1-g}(x-\bar{x}) \pm \frac{t}{b(1-g)} \sqrt{\left(\frac{1-g}{Snw} + \frac{(x-\bar{x})^2}{S_{xx}}\right)}, \qquad (4.6)$$

 TABLE 11.
 Variance and Standard Error of Expected

 Probits in Equation (4.4)

x	V(Y)	8 Y
0.00	0.3031	0.551
0.02	0.2104	0.459
0.10	0.1367	0.370
0.12	0.0820	0.286
0.20	0.0463	0.215
0.22	0.0296	0.172
0.269	0.0283	0 168
0.30	0.0319	0.179
0.35	0.0532	0.231
0.40	0.0934	0.306

where $g = t^2/b^2 S_{xx}$ (Irwin, 1943; Fieller, 1944). Significant heterogeneity must be allowed for by increasing both g and the expression within the square root by the heterogeneity factor. When g is small compared with unity these limits are practically the same as those obtained by the method of Ex. 3, but they become more widely spaced as g approaches unity.*

* Equation (4.6) is a particular case of a very useful theorem stated by Fieller. If a and b are sample estimates of α and β subject to normally distributed random errors, and if v_{11} , v_{12} , v_{22} are joint estimates, from the same sample and based on f degrees of freedom, of the variances and covariance of a and b, then the fiducial limits of the ratio $\mu = \alpha/\beta$ are the roots of

$$\theta^2(b^2 - t^2v_{22}) - 2\theta(ab - t^2v_{12}) + (a^2 - t^2v_{11}) = 0,$$

where t is the appropriate deviate with f degrees of freedom for the chosen probability level. The limits may be written as

$$m + \frac{g}{1-g} \left(m - \frac{v_{12}}{v_{22}} \right) \pm \frac{t}{b(1-g)} \sqrt{\left[v_{11} - 2mv_{12} + m^2 v_{22} - g\left(v_{11} - \frac{v_{12}^2}{v_{22}}\right) \right]}, \quad (4.7)$$

where m = a/b and $g = t^2 v_{22}/b^3$. In the present instance $v_{12} = 0$, but elsewhere, for example in §§ 28 and 47, this is not so.

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Ex. 9. Fiducial limits of the median lethal dose. Reverting to the data analysed in Ex. 6, in which no heterogeneity was found, for the 5 % fiducial limits

and

$$t = 1.96$$

$$g = (1.96)^2 / (4.176)^2 \times 4.613$$

$$= 0.048.$$

The $\log LD 50$ in this example was 0.686, and therefore the fiducial limits are, by equation (4.6),

$$0.686 - 0.050 \times 0.021 \pm \frac{1.96}{4.176 \times 0.952} \sqrt{\left(\frac{0.952}{119.6} + \frac{(0.021)^2}{4.613}\right)}$$

or 0.729 and 0.641, as compared with 0.729 and 0.643 obtained from the approximation $m \pm 1.96s_m$. With so small a value of g, the difference between the two methods is trivial.

In Ex. 7, g is much larger; taking in the heterogeneity factor, and using the 5% value of t with 5 degrees of freedom,

$$g = 3.95 \times (2.57)^2 / 1.040 \times (8.674)^2$$

= 0.333,

a value which is certainly not negligible by comparison with unity. In this example m = 0.239 and the fiducial limits are therefore

$$0.239 - 0.499 \times 0.030 \pm \frac{2.57}{8.674 \times 0.667} \sqrt{\left[3.95 \times \left(\frac{0.667}{139.8} + \frac{0.0009}{1.040}\right)\right]}$$

or 0.224 ± 0.066 . These limits, 0.290 and 0.158, are much wider than the values of 0.293 and 0.185 obtained in Ex. 7 by the simpler method. They may be read directly from the boundaries of the fiducial band in Fig. 9. On the concentration scale the limits are 19.5 and 14.4 mg./l.

Chapter 5

THE COMPARISON OF EFFECTIVENESS

20. RELATIVE POTENCY

CHANGES in the level of tolerance of the population of test subjects frequently make it impossible to rely on assays of materials carried out singly. It is therefore customary, in many types of investigation, to assay a test material against a standard and to measure the performance in relation to that of the standard rather than as an absolute effect. In the second of his early papers on the probit method and its applications, Bliss (1935b) considered the measurement of differences between two or more comparable series of dosage mortality records. His suggestion of measuring differences in terms of relative potency has been widely adopted in the comparison of toxicity data and in biological standardization procedure. Examples have been given by Irwin (1937) and Cochran (1938).

When the dosage scale, in which tolerances of the test subjects in respect of a stimulus are normally distributed, is logarithmic the variances of the tolerances for a number of closely related stimuli are often nearly equal. For example, derris derivatives, such as rotenone, deguelin, toxicarol, and elliptone, haye been found (Tattersfield and Martin, 1938; Martin, 1942) to have similar tolerance variances of their log concentrations when used against *Aphis rumicis* or against *Macrosiphoniella sanborni* under carefully controlled conditions of spraying. This equality of variances is shown in the probit analysis by parallelism of the probit regression lines; the comparison of different series of data is then particularly simple. Parallel regression lines are much less likely to be found with non-logarithmic dosage scales.

The *relative potency* of two stimuli is defined as the ratio of equally effective doses. If two series of quantal response data yield parallel probit regressions against the logarithm of the dose, there is a constant difference between dosages producing

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the same proportion of responding subjects, and consequently a constant relative potency at all levels of response; the relative potency then provides a convenient description of the difference between the two series. The constant dosage difference is usually denoted by M, and is equal to the difference between the median effective dosages. If suffices are used to indicate the two series, the *relative dosage value* of the first series with respect to the second, or the amount by which a dosage in the first series is less than an equally effective dosage in the second, is

$$M_{12} = m_2 - m_1 = \bar{x}_2 - \bar{x}_1 - (\bar{y}_2 - \bar{y}_1)/b, \qquad (5.1)$$

b being the common slope of the two regression lines. For data having $x = \log_{10} \lambda$, the relative potency is then

$$\rho_{12} = 10^{M_{12}}.$$
 (5.2)

The variance of M_{12} is

$$V(M_{12}) = \frac{1}{b^2} \{ V(\bar{y}_1) + V(\bar{y}_2) + (\bar{x}_2 - \bar{x}_1 - M_{12})^2 V(b) \}; \quad (5.3)$$

the standard error, s_M , derived from this variance may be used for assigning approximate fiducial limits to M, but, unless b has been estimated with such precision that $g = t^2 V(b)/b^2$ is small, these limits will be too narrow. As in § 19, exact fiducial limits may be determined (Cochran, 1938) and take the form

$$M_{12} + \frac{g}{1-g} \left(M_{12} - \bar{x}_2 + \bar{x}_1 \right) \\ \pm \frac{t}{b(1-g)} \sqrt{\left[(1-g) \left\{ V(\bar{y}_1) + V(\bar{y}_2) \right\} + (\bar{x}_2 - \bar{x}_1 - M_{12})^2 V(b) \right]}; \quad (5.4)$$

t is the normal deviate for the significance level, unless heterogeneity requires it to be taken from Table VII with the number of degrees of freedom in χ^2 , in which case *g* and the variances must be increased by the heterogeneity factor. This expression, whose analogy with (4.6) is apparent, reduces to $M_{12} \pm s_M t$ when *g* is very small.

The numerical estimation of M is accomplished by straightforward extension of the methods of Chapters 3 and 4. If inspection of the probit diagram suggests that the two regression lines are parallel, the provisional lines are drawn parallel. Working probits are formed and sums of squares and products calculated for each series separately. A new approximation to the common slope, with which the computations may be repeated if necessary, is given by adding the components from each series, and taking

$$b = \frac{{}_{1}S_{xy} + {}_{2}S_{xy}}{{}_{1}S_{xx} + {}_{2}S_{xx}} = \frac{\Sigma S_{xy}}{\Sigma S_{xx}},$$
(5.5)

where Σ indicates summation over the two series. When a satisfactory estimate of b has been obtained, the heterogeneity of the data may be tested in the usual manner by comparing observed and expected numbers of test subjects; the most rapid means of deriving the value of χ^2 for this test is illustrated in Ex. 10. If there is no heterogeneity, the variance of b is

$$V(b) = 1/\Sigma S_{xx},\tag{5.6}$$

so that

$$V(M_{12}) = \frac{1}{b^2} \left\{ \frac{1}{18nw} + \frac{1}{28nw} + \frac{(\bar{x}_2 - \bar{x}_1 - M_{12})^2}{\Sigma S_{xx}} \right\}.$$
 (5.7)

These variances must be increased by the heterogeneity factor . if this is significantly greater than unity.

Though the relative potency of two poisons can be estimated from data on only two doses of each, provided that the regression lines do not markedly depart from parallelism, it is preferable to have at least three doses in each series. In the first discussion of relative potency in relation to probit analysis, Bliss (1935b) has given expressions for M when only two doses have been used for each series and also when a single dose of the test material is assayed against a series of doses of a standard. The latter is an unsatisfactory method of assaying relative potency, as there can be no certainty that the regression line will be parallel to that for the standard, yet the assumption of parallelism is implicit in the estimation and interpretation of M. The method depending on only two doses of each poison should not be used unless the regression relationship is already known to be linear, since the data themselves can provide no information on the existence of a curvature on the x-scale used (Finney, 1944b).

Ex. 10. Relative potency of rotenone, deguelin concentrate, and a mixture of the two. The data considered in Exs. 1, 4 and 6 are

one portion of the results of an experiment fully reported by Martin (1942) in which a deguelin concentrate and a 1:4 mixture of rotenone and deguelin concentrate were tested as well as rotenone alone. The complete results are shown in Table 12, and the empirical probits are plotted in Fig. 10.

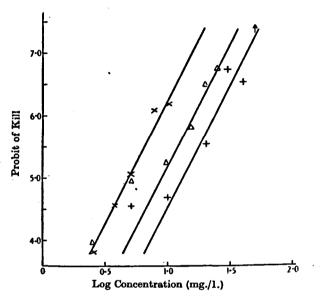


FIG. 10. Probit regression lines for estimation of relative potency of rotenone, a deguelin concentrate, and a mixture of the two (Ex. 10). \times rotenone; + deguelin concentrate; \triangle mixture.

From the figure the lowest concentration of the deguelin concentrate and the two lowest concentrations of the mixture are seen to give points markedly disagreeing with any set of parallel straight lines. The phenomenon of a 'break' in the regression line at low concentrations has been observed in a number of toxicological investigations; here it may be the fesult of the poisons not being chemically pure substances (Bliss, 1939*a*). Dosage-response relationships for mixed poisons are discussed in Chapter 8, but for the present data the difficulty may be overcome by rejecting the three aberrant points from the analysis. Though generally undesirable, this course may be defended here on the grounds that the chief interest lies in the behaviour of

TABLE 12.	Maximum]	Likelihood Con	np	utations for Estimation
of Relati	ve Potency	of Rotenone,	a	Deguelin Concentrate,
and a Mi	xture of the	Two		_

x .	n .	r	p (%)	Em- pirical probit	¥	nw	y_	nwx	nwy
					Roten	one			
1.01 0.89 0.71	50 49 46	44 42 24	88 86 52	6·18 6·08 5·05	6·3 5·8 5·1	16·8 24·6 29·2	6·16 6·05 5·05	16-968 21-894 20-732	103·488 148·830 147·460
0·58 0·41	48 50	16 6	33 12	4·56 3·82	4.6 3.9	$ \begin{array}{r} 28 \cdot 8 \\ 20 \cdot 2 \end{array} $	4.56 3.83	16·704 8·282	131-328 77-366
						119-6]	84.580	608·472
				Degu	elin co	ncentrat	θ		
1.70 1.61 1.48 1.31 1.00 0.71	48 50 49 48 48 48	48 47 47 34 18 16	100 94 96 71 38 33	∞ 6·55 6·75 5·55 4·69 4·56	7·3 6·9 6·4 5·8 4·5 3·4	3.6 7.7 14.8 24.1 27.9	7·68 6·42 6·67 5·53 4·70	6.120 12.397 21.904 31.571 27.900	27.648 49.434 98.716 133.273 131.130
			, ·			78 ·1		99-892	440.201
		·			Mixt	ure			
1.40 1.31 1.18 1.00 0.71 0.40	50 46 48 46 46 47	48 43 38 27 22 7	96 93 79 59 48 15	6.75 6.48 5.81 5.23 4.95 3.96	6·8 6·4 5·9 5·2 4·0 2·8	9·0 13·9 22·6 28·9 	6·75 6·47 5·80 5·23	12.600 18.209 26.668 28.900 	60·750 89·933 131·080 151·147
	- r	$\frac{1}{Snw} =$	• 0-0 08	361204,	\overline{x}_r =	= 0·7072,	$\overline{y}_r =$	5-0876,	
	-					= 1.2790,			
	m					= 1.1610,		= 5·8187.	
Rotenon	ð	($\frac{Snwx^{4}}{59\cdot814}$	00 4 18 4	Snwx 449·56 430·30 19·26	848 570	Snwy 3177.74 3095.63	804 691	140 = 1.671
Deguelir concent			82·039 27·764	10 1 55 1	578-40 563-028	765 391	2541·44 2481·13	933 855	
Mixture			4·274 01·862 00·282	03 8	15-378 508-683 502-600	63	60-31(2542-691 2518-965	182	29 = 4 ∙982
Total		:	1·5799 10·467		6.083 40.724				22 = 0.304 46 = 7.702

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the various toxic materials at high concentrations, in which region Fig. 10 suggests that the data may be adequately represented by three parallel lines.

Provisional lines for the three series of data have been drawn in the figure, and have equations:

$$Y_r = 2 \cdot 30 + 3 \cdot 94x, \quad Y_d = 0 \cdot 59 + 3 \cdot 94x, \quad Y_m = 1 \cdot 24 + 3 \cdot 94x.$$

From these, first approximations to the log LD 50's are

$$m_r = 0.685, \quad m_d = 1.119, \quad m_m = 0.954.$$

A provisional estimate of the relative dosage value of rotenone and deguelin concentrate is therefore

$$M_{rd} = 1.119 - 0.685 = 0.434,$$

giving a relative potency

$$\rho_{rd} = 10^{0.434} = 2.72.$$

Expected probits, Y, are read from the provisional lines and entered in Table 12. Calculations of sums of squares and products proceed for each series separately, those for rotenone being identical with those of Ex. 6, since the same set of provisional probits has been used. Corresponding values of S_{xx} , S_{xy} and S_{yy} are then summed over the three series; for example

$$\Sigma S_{xx} = 4.61282 + 4.27455 + 1.57996$$

= 10.46733.

These are used to give the new estimate of the regression coefficient

$$b = 40.72479/10.46733$$

= 3.8907.

The three regression lines are lines through the three mean points, (\bar{x}, \bar{y}) , with slope b; thus

$$Y_{r} = 5 \cdot 0876 + 3 \cdot 8907(x - 0 \cdot 7072)$$

= 2 \cdot 336 + 3 \cdot 891x,
$$Y_{d} = 0 \cdot 660 + 3 \cdot 891x,$$

$$Y_{m} = 1 \cdot 302 + 3 \cdot 891x.$$
 (5.8)

The differences from the provisional lines are small, leaving little doubt that the hypothesis of a single value of b for the three series is justified. A test of parallelism is given by comparing the sum of the χ^2 values for the three series with that obtained by a similar process from the total sums of squares and products. For these totals

$$\Sigma S_{yy} - (\Sigma S_{xy})^2 / \Sigma S_{xx} = 7.702;$$

this has 10 degrees of freedom, since there are fourteen observations and four parameters have been estimated for the equations. This quantity may be subdivided into a χ^2 with

	D.F.	Sum of squares	Mean square
Parallelism of regressions Residual heterogeneity	2 8	0·745 6·957	0·372 0·870
Total	10	7.702	

TABLE 13. Analysis of χ^2 for Ex. 10

8 degrees of freedom, which is the sum of the χ^2 values for the three series separately, and a residual χ^2 with 2 degrees of freedom dependent on the departure from parallelism of the three lines. The analysis of χ^2 in this way is shown in Table 13. The second line of the analysis shows a χ^2 of 6.957 for heterogeneity, a value which is clearly not significant; the sum of squares for parallelism may therefore be tested as a χ^2 . Since 0.745 is not significant, there is no evidence of any conflict with the hypothesis that the three lines are parallel. Had the heterogeneity χ^2 been significant, the ratio of the two mean squares (obtained by dividing the sums of squares by the degrees of freedom) would have been tested by Fisher's tables of the variance ratio (Fisher and Yates, 1948, Table V), and the mean square for heterogeneity would have been used as the heterogeneity factor in the calculation of variances.

The new estimates of the $\log LD$ 50's are, from equations (5.8),

$$m_r = 0.6847, \quad m_d = 1.1154, \quad m_m = 0.9604,$$

corresponding to LD 50 values of 4.84, 13.04 and 8.92 mg./l. respectively. The standard error of m may be calculated from equation (3.6), as in Ex. 6, remembering that b has now been estimated from the more extensive data, so that, by equation (5.6),

$$V(b) = 1/10.4673.$$

The revised value of the estimate of the relative dosage value of rotenone and the deguelin concentrate is

$$M_{rd} = 0.4307 \pm 0.0390,$$

the standard error being derived as the square root of the variance; from equation (5.7)

$$V(M) = \frac{1}{(3\cdot8907)^2} \left\{ \frac{1}{119\cdot6} + \frac{1}{78\cdot1} + \frac{(0\cdot5718 - 0\cdot4307)^2}{10\cdot4673} \right\}$$

= 0.00152.

The relative potency is therefore

$$\rho_{rd}=2.70\pm0.24,$$

the standard error of the antilogarithm of M being obtained by equation (3.7). Rotenone is thus estimated to be 2.70 times as toxic to M. sanborni as the deguelin concentrate, or, in other words, in order to give a kill as great as that produced by a given concentration of rotenone a 2.70 times greater concentration of deguelin concentrate would be required.

For the 5 % fiducial limits of M, t = 1.960 and therefore

$$g = 3.84/10.47 \times 15.14$$

= 0.024.

a value sufficiently small to be ignored; consequently fiducial limits of M may be taken as $M \pm 1.960 \times 0.390$ or 0.5071 and 0.3543. Hence the fiducial limits for ρ are 3.21 and 2.26.

In the same manner, the potency of the mixture relative to rotenone or the deguelin concentrate may be estimated. Further consideration of these three relative potencies will be deferred until Ex. 20, where the data will be used as an illustration of similar joint action of two poisons.

21. COMBINATION AND COMPABISON OF RELATIVE POTENCIES

When the relative dosage value of two poisons has been assayed separately by two or more series of tests, an improved estimate is given by the weighted mean, using for weights the reciprocals of the variances of the estimates of M (Cochran, 1938). A weighted sum of squares of deviations of M may be used as a χ^2 in a test of the heterogeneity of the estimates averaged (Miller, Bliss and Braun, 1939). When there is no evidence of heterogeneity the sum of the weights is the weight to be attached to the mean; when a significant χ^2 occurs the total weight must be divided by the heterogeneity factor.

Ex. 11. Combination of relative potencies. In separate tests Tattersfield and Martin (1938, Table I) have compared a toxicarol precursor (potash separated), prepared in three different ways, with rotenone. The dosage values relative to rotenone were -1.164 ± 0.036 , -1.186 ± 0.029 and -1.114 ± 0.042 . Differences between⁶ these barely exceed their standard errors, and it is therefore reasonable to form a combined estimate using the reciprocals of the squares of the standard errors as weights. The estimate is

$$M = -\frac{772 \times 1 \cdot 164 + 1189 \times 1 \cdot 186 + 567 \times 1 \cdot 114}{772 + 1189 + 567}$$
$$= -2940 \cdot \frac{4}{2528}$$

= -1.163.

The variance of M is 0.000396, the reciprocal of the sum of the weights, and the standard error is therefore ± 0.020 . The test of heterogeneity of the separate estimates is scarcely required here, in view of the close agreement between them; the appropriate χ^2 is

$$\chi_{[2]}^2 = 772 \times (1 \cdot 164)^2 + 1189 \times (1 \cdot 186)^2 + 567 \times (1 \cdot 114)^2 - (2940 \cdot 4)^2 / 2528 = 1 \cdot 99.$$

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With weights as nearly equal as these, little is lost by taking the unweighted mean

$$M' = -(1.164 + 1.186 + 1.114)/3$$

= -1.155,

the variance of which is

$$V(M') = \{(0.036)^2 + (0.029)^2 + (0.042)^2\}/3^2$$

= 0.000433.

This estimate therefore has a standard error of ± 0.021 and is only 9% less precise than the weighted mean.

If two poisons have been tested in different experiments, even though they show similar values of b a direct estimation of their relative dosage value as the difference in the estimated log LD 50's may be misleading: the level of susceptibility of the test subjects may have been different in the two experiments or experimental conditions may have differed in such a way that the absolute potency of any poison is different in the two experiments though relative potencies within an experiment are unaffected. An estimate of relative potency independent of any difference in susceptibility may still be made, providing that each poison can be compared with a third which has been tested in both experiments. Estimates of M_{13} and M_{23} can then be made 'within experiments'; the required M is estimated as

$$M_{12} = M_{13} - M_{23}, \qquad (5.9)$$

$$\rho_{12} = \rho_{13} / \rho_{23}.$$

The variance of M_{12} is the sum of the variances of M_{13} and M_{23} .* The method should only be used when the conditions in the two experiments are sufficiently similar for the estimate obtained to be relevant.

* Cochran (1938) suggested this procedure for testing the significance of a difference in potency of poisons used in separate experiments. The significance should be assessed from the normal distribution, not the *t*-distribution, unless the variances have been adjusted by a heterogeneity factor.

and

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Ex. 12. Comparison of relative potencies. Tattersfield and Martin (1938, Table I), experimenting with rotenone and a resin derived from a Sumatra-type derris root, found a relative dosage value

$$M_{\rm sr} = -0.800 \pm 0.025.$$

In a second experiment, using rotenone and a *Derris elliptica* resin, they found

$$M_{er} = -0.552 \pm 0.031.$$

Both experiments were spraying trials against *Aphis rumicis*, the same technique being used for both. Now

$$(0.025)^2 + (0.031)^2 = 0.001586 = (0.040)^2;$$

hence the relative dosage value of the two resins is estimated as

$$M_{se} = -0.248 \pm 0.040,$$

and the Sumatra-type resin is estimated to be 0.56 times as toxic as the *Derris elliptica*.

Ex. 13. The relative toxicities of seven derris roots. A further example is provided by earlier data from Tattersfield and Martin (1935) in which seven samples of derris root were tested for their toxicity to Aphis rumicis. Tests were made on six occasions, a different pair of roots being compared on each occasion. The proportions of badly affected, moribund, and dead at each concentration have been re-examined for this example, using as the measure of dosage the logarithm of the concentration of ether extract in mg./l. The log LD 50 and M values for these six tests were found to be (the suffices referring to the seven roots)*:

Test 1
$$m_2 = 1.539$$
, $m_5 = 1.607$, $M_{25} = 0.068$,
, 2 $m_5 = 1.695$, $m_6 = 1.452$, $M_{56} = -0.243$,
, 3 $m_5 = 1.622$, $m_7 = 1.384$, $M_{57} = -0.238$,
, 4 $m_4 = 1.631$, $m_5 = 1.682$, $M_{45} = 0.051$,
, 5 $m_1 = 1.496$, $m_3 = 1.492$, $M_{13} = -0.004$,
, 6 $m_1 = 1.440$, $m_7 = 1.421$, $M_{17} = -0.019$.

* Allowance was made for the natural death rate estimated from control batches, the methods of Chapter 6 being used.

From these estimates the relative potency of any two roots can be estimated by comparisons within experiments. For example, the relative potency of roots 1 and 4 may be estimated from the relative dosage value

$$M_{14} = M_{17} - M_{57} - M_{45} = 0.168;$$

the standard errors are not shown here, but are obtainable by the same method as before. A convenient way of summarizing the LD50's is to take a mean value for root no. 5, which was tested four times, and to place the others at the estimated distances from this; this mean might be obtained by appropriate weighting, as in Ex. 11, but the simple arithmetic mean of the four values, 1.652, is almost equally good. The log LD 50's may then be written as:

Root no. 1:	1.433	Root no.	5: 1.652
,, 2:	1.584	>> _	6: 1.409
,, 3:	1.429	,,	7: 1.414
,, 4:	1.601		

The values are very similar to those computed independently by Bliss (1939*a*). The variance of M_{14} is the sum of the variances of M_{17} , M_{57} and M_{45} , each being taken from a separate experiment, and the standard error of any relative dosage value should be obtained in this way rather than by using standard errors of each log LD 50, such as Bliss has estimated.

Had more tests of pairs of these seven roots been carried out, it would have been possible to estimate some of the relative potencies by more than one chain of pairs. In order to obtain the best possible estimates the method of least squares would then have had to be used (see Ex. 14), though data from carefully controlled trials might be sufficiently consistent for satisfactory estimates to be made, by simple averaging, without this process.

Ex. 14. Combination of relative potencies by the method of least squares. As a more complex example of the estimation of relative potencies from a series of tests, data published by Martin (1940) on tests of four different derris roots as poisons for A. rumicis

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will be considered. Of these roots, known as W. 211, W. 212, W. 213 and W. 214, two sets of three were tested on different occasions, and on a third occasion W. 211 and W. 213 were compared with rotenone. The estimated log LD 50's are shown in Table 14, together with the weights (reciprocals of the variances) to be attached to each. For each occasion the three estimates were obtained from a common regression coefficient as in Ex. 10, but in the computations modified weighting coefficients (§ 27) were used in order to take account of 2-4 % mortality amongst the control batches of insects.

TABLE 14.Estimated log LD 50's and their Weights forRotenone and Four Derris Roots on Three Occasions

Date	Rotenone	W. 211	W. 212	W. 213	W. 214	Totals
15. vi. 38		2.430	2.228		2.022	22138·82
	_	(1790)	(3610)	_	(4820)	(10220)
27. vii. 38		2.420	2.156	2.255	· _ /	21392.03
	-	(3520)	(2530)	(3290)	<u> </u>	(9340)
18. vü. 39	0.980	2.193	· ·	2.191	i —	7788-62
	(1640)	(1400)	—	(1420)	· _	(4460)
	1607-20	15938-30	13497.76	10530-17	9746-04	51319-47
	(1640)	(6710)	(6140)	(4710)	(4820)	(24020)

 $m = 2 \cdot 1365308.$

The estimated log LD 50's for any one occasion are not independent, since they are based on the same regression coefficient. Hence the variance of a relative dosage value obtained for two poisons on one occasion is not simply the sum of two variances of the form of equation (3.6), but is an expression like equation (5.3). In this example the second term of equation (3.6) was in every instance small relative to the first and thus had little effect on the variance, as may be expected when knowledge of the poisons allows the experiment to be so planned that the mean probit, \bar{y} , shall be near to 5. The complication of this nonindependence has therefore been ignored in the analysis of the data. In order to estimate the relative potencies of the five materials under comparable conditions, account must be taken of the difference in susceptibility of the insects on the three 78 THE COMPARISON OF EFFECTIVENESS

occasions. If the relative potency of any pair remained the same on all occasions, the log LD 50 of one material on one occasion is expressible as the sum of three components, the general mean, a root or column constant, and an occasion or row constant. For example, the log LD 50 for root W. 211 on 27 July 1938 should be expressible as $m+c_1+r_2$, where m is the general (weighted) mean, c_1 is one of the five column constants c_r , c_1 , c_2 , c_3 , c_4 corresponding to the root, and r_2 is one of the three row constants is to represent deviations about the general mean, the weighted totals must be zero, so that

$$\begin{array}{r}
1640c_r + 6710c_1 + 6140c_2 + 4710c_3 + 4820c_4 = 0, \\
10220r_1 + 9340r_2 + 4460r_3 = 0.
\end{array} \tag{5.10}$$

Estimates of all the constants can be obtained by minimizing the weighted sum of squares of differences between the values in Table 14 and their expectations in terms of the constants. The minimizing conditions are simply statements that the weighted total for any one of the columns or rows is equal to its expected value. These weighted totals are shown in the margins of Table 14, with the total weights in brackets. Thus, for example, the second column gives the equation

 $6710m + 1790(c_1 + r_1) + 3520(c_1 + r_2) + 1400(c_1 + r_3) = 15938 \cdot 30.$ The general mean is

 $m = 51319 \cdot 47/24020 = 2 \cdot 1365308$,

and the equations therefore reduce to the following form, in which only the coefficients of the eight unknowns are tabulated on the left-hand side:

c,	<i>c</i> 1	<i>c</i> ₂	c_3	c 4	r 1	r_{2}	r ₃		
1640	•		•		•	•	1640 = -	1896-71,	
•	6710			•	1790	3520	1400 =	1602.18,	
		6140	-	•	3610	2530	. =	379-46,	
			4710	•		3290	1420 =	467-11,	
	•	•	•	4820	4820	•	·· . = -	552.04,	(5-11)
	1790	3610		4820	10220	-	. =	303.48,	
	3520	2530	3290			9340	. =	1436-83,	
1640	1400	•	1420	•	•	· •	4460 = -	1740-31.)	

These equations are not entirely independent, and, before they can be solved, need to be supplemented by the two conditions on weighted totals already stated. Though they appear complicated, a little study of their structure should enable the analogous equations for other similar sets of data to be written down quite easily. The solution is not difficult, but is tedious; the results to any required degree of accuracy may be obtained by a process of successive approximation, and are

$$c_r = -1.0177,$$
 $r_1 = 0.0576,$
 $c_1 = 0.2507,$ $r_2 = 0.0033,$
 $c_2 = 0.0266,$ $r_3 = -0.1388.$
 $c_3 = 0.1387,$
 $c_4 = -0.1721,$

The column constants give the required balanced comparisons between the five poisons; the row constants are of no interest except for comparing the susceptibilities of the insects on the

 TABLE 15.
 Comparative Potencies of Rotenone and Four Derris Roots

			Potency relative to W. 211		
Material	log LD 50	LD 50 (mg./1.)	Present analysis	<u>Martin</u> (1940)	
Rotenone	1.119	13.2	18.50		
W. 211	2.387	244·0	1.00	1.00	
W. 212	2.163	146-0	1.68	1.76	
W. 213	2.275	188.0	1.29	1.31	
W. 214	1.964	92-0	2.65	2.68	

three occasions. By addition of the general mean, m, comparable values of log LD 50 for the five materials are obtained, and these are shown in the second column of Table 15. The last two columns of the table give the relative potencies as estimated by the above analysis and as previously given in Martin's paper,^{*} where an

• When Martin's paper was written, the modified weighting coefficients (§ 27) had not been developed, and though the percentage kills were adjusted for mortality in the controls the ordinary weighting coefficients were used. As the control mortality never exceeded 4%, the difference in method is not important.

approximate method suggested by the present writer was used; the approximation is here close to the least squares solution but might give misleading results with less regular data.

Table 16 shows the expected LD 50 values, corresponding to the experimental values in Table 14, and has been constructed by adding the appropriate pairs of row and column constants to the general mean. The goodness of the agreement of the data and the hypothesis of constant relative potencies may be assessed by means of a χ^2 test. The weighted sum of squares of deviations of the data in Table 14 is

$$1640 \times (0.980)^2 + ... + 4820 \times (2.022)^2$$

 $-(51319\cdot47)^2/24020=2779\cdot85,$

and of this the fitted constants account for an amount

$$-1896 \cdot 71c_r + 1602 \cdot 18c_1 + \dots - 1740 \cdot 31r_3 = 2765 \cdot 61.$$

 TABLE 16.
 Values of log LD 50 Fitted to Data of Table 14

 on Assumption of Constant Relative Potencies

Rotenone	W. 211	W. 212	W. 21 3	W. 214
	2·445 2·390	2·221 2·166	2·278	2·022
		<u> </u>	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $

Though apparently eight constants have been fitted, only six of these (two 'row' and four 'column') are independent, since the weighted sums were constrained to be zero. The residual χ^2 therefore has 2 degrees of freedom (9, less 1 for the mean and 6 for the constants); hence

$$\chi^2_{[2]} = 14.24.$$

The same value, apart from errors of rounding off, may be obtained as the sum of squares of the discrepancies between corresponding entries in Tables 14 and 16. The significance of χ^2 indicates that the relative potencies of the five poisons did not remain constant throughout the investigation; nevertheless, the agreement between Tables 14 and 16 is probably good enough for most practical purposes.

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The standard errors of the log LD 50's in Table 15 may be estimated by the use of routine processes in the method of least squares, but a detailed discussion is beyond the scope of the present work.

22. DESIGN OF EXPERIMENTS

When several poisons are to be compared in one experiment it may be impossible to test them all on one day or all on one homogeneous stock of insects. Precautions must then be taken to avoid any bias in the results arising from inequalities in the susceptibility of the insects on different days or in different stocks. A similar problem may be encountered when tests have to be made in several laboratories, or by several workers in the same laboratory; even though standardized methods of testing are used, variations may occur in the effectiveness of the same poison used by different workers. Illustrations have been given in Exs. 13 and 14 of methods of combining results from groups of tests where each group contains a different selection of poisons from all those under investigation and the general level of potency or effectiveness may vary from one group to another. These, however, are not ideal examples of how to deal with the general problem, since their lack of symmetry complicates the analysis and leads to final estimates of log LD 50's which differ widely in precision.

In planning an investigation of poisons which are too many for all to be tested on a single occasion, or which for some other reason have to be divided into groups for testing, it is usually an advantage to introduce an element of balance into the arrangement adopted, so that unbiased and reliable comparisons can be made between every pair of poisons. The levels of dose and numbers of insects should be chosen, in the light of existing knowledge, with the aim that all estimates of log LD 50 have about the same precision. Careful planning of an investigation before it is begun is always to be preferred to the haphazard accumulation of results, for no amount of detailed statistical analysis can extract satisfactory answers to the questions propounded if the experiments were badly designed for obtaining the answers.

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Moore and Bliss (1942) have described an experiment in which seven different organic compounds were tested for their toxicity to Aphis rumicis. Sets of three compounds were tested under comparable conditions on each of seven days; the sets were so chosen that each poison was used on three days and occurred once and once only on the same day as each of the others. Moore and Bliss have discussed analysis of variance methods appropriate to these data, ignoring the differential weighting of the observations arising in the course of the usual probit analysis. They obtained unbiased comparisons between the poisons, though still more precise estimates might possibly be made from the information contained in the data. In this experiment there was a heavy mortality amongst insects sprayed with the spreader only, and in view of the considerations advanced in Chapter 6 there should be some modifications in the analysis, but the results will not be discussed here.

This experiment is said to have a balanced incomplete block design (Yates, 1937b, 1940). If the seven insecticides are denoted by A, B, ..., G, the seven blocks or sets of three may be written

A	A	A	B	B	C	C
B	D	F	D	E	D	E
C	E	Ģ	F	G	G	F

Yates has described how the effects of differences between days may be eliminated and the seven poisons compared by means of 'within-day' comparisons only; he has also discussed the utilization of information from comparisons 'between days', an addition which may be of value in more extensive experiments though scarcely likely to be so here. There is, of course, no advantage in this arrangement in blocks unless the blocks are likely to differ appreciably in the results which they would give for any one poison. Only experience can help the experimenter to distinguish circumstances in which the control of variations in experimental conditions by means of blocks may be expected to be advantageous from those in which it is unnecessary and may even lead to a reduction in precision. Few published accounts of researches on insecticides, or, indeed, on other aspects of biological assay, refer to this problem.

The subject of experimental design has been developed primarily in connexion with agricultural field trials, but the principles are undoubtedly useful in planning biological assays. The properties of many other designs have been studied, and valuable accounts of these have been given by Fisher (1942), Fisher and Yates (1948, Tables XV-XIX) and Yates (1937*a*), to whose work the reader seeking information on randomized blocks, Latin squares, confounding, and like topics must be referred; a brief discussion here would be inadequate and a full treatment would be out of place. Further research into the adaptation of the principles of experimental design and the analysis of the results of well-planned experiments is undoubtedly needed. Another branch of this subject, factorial design, is discussed in Chapter 7.

23. PRECISION OF ASSAYS

Miller, Bliss and Braun (1939) have discussed a number of methods for increasing the precision of estimates of M. The experimenter may always attain greater precision by using larger numbers of test subjects, but this is often inconvenient or impossible. In planning assays of M, therefore, consideration must usually be given to the most economical utilization of a limited number of subjects. If some information on the value of a relative potency already exists, doses of the test material and the standard may be chosen so as to bear this ratio to one another; the mean probits in a new assay should then be nearly equal, and the third term in the expression for the variance (equation $(5\cdot3)$) will be small. In choosing the doses the experimenter should remember that very low or very high kills give low values of the weighting coefficient, w, but that, on the other hand, if the doses used are too close together the value of S_{xx} will be small and b therefore of low precision. When nothing but M is to be estimated, and some information on the LD 50's is already available, probably the best compromise is to aim at sets of expected kills lying between 4 and 6 probits and as far as possible to avoid probits lower than 3 or higher than 7. On the other hand, when little is known of the LD 50's, a sufficiently wide range of doses should

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be used to ensure that, at all costs, the kills obtained with each poison bracket the 50 % point. The slope of the regression lines, being the reciprocal of the standard deviation of the dosagetolerance distribution, is also of some interest, and for its satisfactory estimation a rather wider range of doses (and consequently of kills) may be desirable than would be ideal if M alone were considered.

The expression for the variance of M is also reduced by any procedure which increases the slope of the regression lines. The regression coefficient, b, however, is not under the control of the experimenter in the same way as are the dose levels, but is a characteristic of the poison, the test subject, and the conditions of the experiment. Though the precision of an assay of relative potency may be increased by a change in the test subject or in the experimental conditions, there may then be some doubt of whether the same quantity is being estimated. Bliss and Cattell (1943, pp. 482-5) refer to instances of drug assay with animals which have given estimates of relative potency very different from the values found for man, in some cases even the order of potency being reversed. The relative potencies of insecticides may be altered by a change in spray medium or in method of application, and may be entirely different for different species of insect.

It might be thought that, in assaying the potency of a poison or other stimulus relative to a standard, precision would be increased by comparing the response to the substance under test with that found for the standard in all previous tests, rather than with the small amount of data for the standard obtained in the course of the current assay. The danger in this approach is that, though conditions can usually be so controlled that relative potency remains reasonably constant in tests made over a period of time, absolute potency often varies very considerably from day to day. The data examined in Ex. 14, for example, indicate that the poisons on the last occasion of the tests averaged 60 % higher potency than on the first. Bliss and Packard (1941) have reported that exposure of eggs of *Drosophila melanogaster* to röntgen rays gave the same probit regression relationship between kill and intensity of irradiation on a number of occasions over a period of seven years, and Whitlock and Bliss (1943) have described a series of antihelminthic tests in which the position of the regression line remained the same, within the limits of experimental error. Nevertheless, this state of affairs appears to be exceptional, and in any new investigation the experimenter should be prepared for much greater instability in the mean log tolerance, m, than in the slope, b, until experience shows otherwise.

24. MEAN PROBIT DIFFERENCE

A second measure of the difference between two series of observations giving parallel probit regressions is the *mean probit difference* (Finney, 1943*a*). The mean probit difference, Δ , between two parallel probit lines is defined as the constant vertical difference between the lines

$$\begin{aligned} \Delta_{12} &= Y_1 - Y_2 = b M_{12} \\ &= \overline{y}_1 - \overline{y}_2 - b(\overline{x}_1 - \overline{x}_2). \end{aligned}$$
 (5.12)

The variance of this expression is

$$V(\Delta_{12}) = V(\bar{y}_1) + V(\bar{y}_2) + (\bar{x}_1 - \bar{x}_2)^2 V(b), \qquad (5.13)$$

which may be written

$$V(\Delta_{12}) = \frac{1}{{}_{1}Snw} + \frac{1}{{}_{2}Snw} + \frac{(\bar{x}_{1} - \bar{x}_{2})^{2}}{\Sigma S_{xx}}$$

when no heterogeneity factor is needed.

Though Δ is in some ways easier to use than M, particularly in respect of its fiducial limits which do not require complex expressions like (5.4), it has the serious disadvantage of giving a comparison that is much less readily interpreted in practical terms. Indeed, Δ only measures the difference in effect of equal doses—and that in a unit whose value in percentages is not constant at all parts of the scale—instead of comparing the magnitude of equally effective doses. The mean probit difference may sometimes provide a convenient statement of results, but is seldom useful without a knowledge of the regression coefficient. Values of Δ between different pairs of poisons and over a series of experiments may be combined and compared just as were values of M in Exs. 11–14, the weights again being the reciprocals of the variances.

Ex. 15. Mean probit difference of rotenone and deguelin concentrate. From equations (5.8) the mean probit difference between rotenone and the deguelin concentrate is

$$\Delta_{rd} = 1.676 \pm 0.229,$$

the standard error being obtained by equation (5.13) as

$$V(\Delta) = \frac{1}{119 \cdot 6} + \frac{1}{78 \cdot 1} + \frac{(0.5718)^2}{10.4673}$$

= 0.0524.

Fiducial limits for Δ are obtained by addition and subtraction of the appropriate multiple of the standard error. For rotenone and the deguelin concentrate the 5 % fiducial limits differ from Δ by $\pm 1.960 \times 0.229$, and are therefore 2.125 and 1.227.

25. UNEQUAL TOLERANCE VARIANCES

When the probit regression lines of two series of tests are not parallel, the interpretation of their comparative effects is more difficult. The relative dosage value of the two poisons can still be measured at a selected level of kill, but this quantity will be different at another level. The variance of such a relative dosage value is the sum of the variances of the two dosages estimated to give that kill, each being obtained from equation (3.6). Similarly, the probit difference between the poisons must be quoted for a specified dosage. Thus much of the usefulness and simplicity of these two measures is lost when the variances of the distribution of tolerance are unequal.

An apparent lack of parallelism may sometimes disappear if a more suitable x-scale is chosen, though this is unlikely unless the separate series also give indications of departing from linearity of regression on the scale first used. Up to the present no simple method of comparing the effects of two poisons whose

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regression lines are not parallel has been developed, though, as will appear in § 38, consideration has been given to the form of dose-mortality relationship for mixtures of two such poisons. Indeed, for two substances of totally different chemical constitutions or modes of action on the test subjects, there is no reason to expect an easy expression of the difference in the dose-response relationships and the term 'relative potency' may cease to have much useful meaning.

Chapter 6

ADJUSTMENTS FOR NATURAL MORTALITY

26. ABBOTT'S FORMULA

THE responses of the test subjects have so far been assumed to be entirely due to the effects of the stimuli applied, and no allowance has been made for any responses which might have occurred without these stimuli. In some instances the assumption may be justifiable, and, indeed, almost all the numerical examples used in earlier chapters were chosen as containing evidence of little or no natural mortality under the conditions of the test. This state of affairs does not always obtain. Control batches of insects, untreated or possibly treated with a spray medium having no toxic content, often show appreciable death rates in the period between the application of insecticides to the test batches and the examination of the results of this application. A similar situation arises in ovicidal tests when it is impossible to distinguish between fertilized and unfertilized eggs at the start; the observed percentage of eggs failing to hatch must be adjusted in order to allow for the percentage unfertilized in the population from which the test batches were taken. Again, in tests of fungicides by means of spore germination counts the adjustment is required in order to allow for the existence of spores which would not germinate even in the absence of any fungicide.

If in a toxicity test a proportion C of test subjects would die even without any poison, the total death rate expected from a dose sufficient to kill a proportion P of those which would otherwise survive is

$$P' = C + P(1 - C), (6.1)$$

providing that the two types of mortality operate independently. From this equation it follows that, if the total proportion dead is P', the proportion killed by the poison alone is

$$P = (P' - C)/(1 - C).$$
(6.2)

This is commonly known as 'Abbott's formula', on account of its use in a paper by W. S. Abbott (1925) on the adjustment of the results of insecticidal tests; in fact, it had been used earlier, for the same purpose, by Tattersfield and Morris (1924), and is an application of the well-known rule for the combination of independent probabilities.

27. APPROXIMATE ESTIMATION OF THE PARAMETERS

The effects of the adjustment for natural mortality on the maximum likelihood estimation of the parameters of the tolerance distribution have usually been ignored. Most important of these is that the effective number of subjects exposed to the poison is no longer n, the total number in a batch, but, on the average, n(1-C). This was realized by Bliss, who, in the discussion of an ovicidal test (1939a, p. 602), states: 'Both the number of eggs exposed and the percentage kill have been corrected for mortality in the untreated controls.' Examination of his correction shows the number of eggs tested at each concentration to have been reduced by the percentage mortality in the controls; thus the weights attached to each observation were proportionately reduced. The estimates of potency are not altered, but their precision is less than if no adjustment had been required.

Even when C is known exactly, this is not the only alteration required in the probit analysis. In the expression for the weighting coefficient (equation (3.4)), the product PQ arises as the variance of a binomial frequency distribution; when C is not zero, the relevant distribution is that defined by the total proportions of dead and surviving, P' and Q'. Finney (1944*a*) has shown that the two adjustments may be combined by taking as the weighting coefficient

$$w = \frac{Z^2}{Q\left(P + \frac{C}{1 - C}\right)} \tag{6.3}$$

instead of the usual $w = Z^2/PQ$, and multiplying by the unadjusted *n*. Values of this quantity, at intervals of 1 % in *C* from zero to 90 % and at intervals of 0.1 in the expected probit, are given in Table II.

ADJUSTMENTS FOR NATURAL MORTALITY

When C = 0, (6.3) reduces to the formula for the ordinary weighting coefficients; for any other value of C these coefficients must be multiplied by $P / \left(P + \frac{C}{1-C}\right)$. Even when C is no greater than 5 %, the reduction in the value of w may be considerable, especially if the expected probit is small. Except when C = 0, w is not symmetrical about the value Y = 5, but decreases much more rapidly for small values of Y than for large. For most practical purposes, it is sufficient to determine w from a value of C given to the nearest 1 %, so that interpolation in Table II is seldom needed. Beyond the range of Table II, w may be calculated with the aid of Table I, as illustrated in Ex. 16.

Ex. 16. Calculation of weighting coefficients. Suppose that it is required to find the value of w for an expected probit Y = 6.2 and a control mortality of 59 %. The mortality P, corresponding to Y, must first be read from Table I as

$$P = 0.8849.$$
Also
$$C/(1-C) = 1.4390,$$
and therefore
$$P/(P + \frac{C}{1-C}) = 0.3808.$$

When C = 0, the weighting factor for this Y is 0.37031 (Table II), and therefore for C = 0.59

$$w = 0.37031 \times 0.3808$$

= 0.1410.

This may be compared with the value for Y = 3.8, for which

$$P = 0.1151, P / \left(P + \frac{C}{1-C} \right) = 0.0741, \text{ and } w = 0.0274.$$

In practice C is seldom, if ever, known exactly, and must instead be estimated from a sample of the population just as are other mortality rates. If this control batch is large relative to the batches used for the different doses in the experiment, C may be estimated satisfactorily from it alone, a procedure which ignores the information on C contributed by the mortalities observed for the test doses. The observed mortality

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amongst the controls, c, is itself subject to sampling variation, and, though it is an unbiased estimate of C, a better estimate may often be obtained by the use of additional information, especially that from the lower dose levels of the poisons under test. Such a dose may, for example, give a total mortality, p', which is less than c, and thus suggest that c is an overestimate of the true natural mortality, C. Similarly, a series of values of p' at small doses which are nearly equal to one another but much in excess of c, indicates that c is an underestimate. In such circumstances an improved estimate of C may be obtained either from inspection of the data or from a freehand sketch of the sigmoid response curve relating dosage to percentage kill.

When C is not too large (say less than 20 %), a probit analysis based on the estimate c, or on an estimate modified as suggested in the last paragraph, and using equation (6.3) for the weighting coefficient but otherwise proceeding as in Chapter 4, is very often sufficiently near to the maximum likelihood treatment of the data for practical purposes. The estimate of C may be greater than certain of the p', so that Abbott's formula yields negative values for p. Though no meaning can be attached to negative mortalities, the correct procedure is still to calculate and use working probits according to the rules of Chapter 4, as only in this way can each observation exercise its right influence in the estimation of the regression line or lines. Working probits corresponding to negative values of p are not often required, and have not been included in Table IV; when needed they must be calculated from Table III. No example of the use of these modified weighting coefficients in the manner just described need be given, as the method is exactly the same as that customarily employed when C = 0.

28. THE MAXIMUM LIKELIHOOD ESTIMATES

If a series of tests provides evidence that the natural mortality rate is high, or if the control batch is so small or the test mortalities so irregular that estimation of C is difficult, the full maximum likelihood process, following the lines indicated in

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Appendix II should be carried out (Finney, 1949d). An auxiliary variate, x', is defined by

$$x'=Q/Z; (6.4)$$

numerical values of x' are given in Table II. The maximum likelihood equations are then set up:

$$bS_{xx} + \frac{\delta C}{1 - C} S_{xx'} = S_{xy}, bS_{xx'} + \frac{\delta C}{1 - C} S_{x'x'} = S_{x'y},$$
(6.5)

and

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$$a = \overline{y} - b\overline{x} - \frac{\delta C}{1 - C}\overline{x}'. \tag{6.6}$$

These equations need explanation. A first approximation to C is determined as described in §27, and used in Abbot's formula to give values of p. From these, a provisional probit line is constructed, and weights (using Table II) and working probits obtained in the ordinary manner. An additional column is added to the computing sheet to show Q/Z for each Y. Equations (6.5) give b and $\delta C/(1-C)$ as the partial regression coefficients of y on x and x'; b is a new approximation to the probit regression coefficient, $(C + \delta C)$ a new approximation to the natural mortality rate. Weighted sums of squares and products are used, so that, for example,

$$S_{xx'} = Snw(x - \bar{x}) (x' - \bar{x}')$$

= Snwxx' - (Snwx) (Snwx')/Snw.

Equation (6.6) gives the constant term in the new regression equation

$$Y = a + bx. \tag{6.7}$$

If n_c subjects are tested at zero dose, and r_c of these die, an estimate of the natural mortality rate,

$$c = r_c/n_c, \tag{6.8}$$

is derived from the control group alone; in equations (6.5),

$$\frac{n_c(1-C)}{C} \tag{6.9}$$

$$\frac{n_c(c-C)}{C} \tag{6.10}$$

and

must be added to $S_{x'x'}$ and $S_{x'y}$ respectively, as allowances for the information provided by this group.

If the revised estimate of the natural mortality rate or the revised regression equation differs appreciably from that on which the calculations have been based, a second cycle must be computed. The process of iteration must continue, in the usual manner, until differences between successive cycles become unimportant. At that stage, δC will be practically zero, so that equations (6.5) will give b as the total regression coefficient of y on x, and equation (6.6) will give the constant term in the regression equation as $(\bar{y} - b\bar{x})$. Herein lies the justification for the approximate method of analysis described in §27, for if a value of C close to the maximum likelihood estimate can be chosen by inspection of the data, only the modification of the weighting coefficients is needed in order that the method of Chapter 4 may estimate the other parameters.

When a satisfactorily close approach to the maximum likelihood estimates has been made,

$$\chi^2 = S_{yy} - bS_{xy} - \frac{\delta C}{1 - C}S_{x'y}$$
(6.11)

gives a test of heterogeneity; in assigning degrees of freedom to χ^2 , a group of subjects at zero dose counts as one dose level, and one degree of freedom must be subtracted for the additional parameter, C. In equation (6.11), S_{yy} is to be interpreted as the usual sum of squares of deviations plus a contribution $n_c(c-C)^2/C'(1-C)$ from the controls. This χ^2 is subject to the usual limitations, in that it is strictly valid only in the limit of the iteration and that it may be disturbed by small expectations.

Equations (6.5) should be solved by first deriving the inverse matrix of the coefficients, or what Fisher (1944, §29) calls the set of c-multipliers. From this, both the estimates of the parameters and their variances may be formed. An example of the calculations has been given elsewhere (Finney, 1949b); Ex. 17 illustrates an application to the fundamental biological assay problem, the comparison of two sets of data with probit regression equations constrained to be parallel.

x	n	r	p'	$p \\ (C = 17.0)$	Em- pirical probit	Y	nw	x' = Q/Z	y	nwx	nwx'	nwy	Equa- tions (6·14)
							Root W.	213					
2·17 2·00 1·68 1·08	142 127 128 126	142 126 115 58	$ \begin{array}{r} 100.0 \\ 99.2 \\ 89.8 \\ 46.0 \end{array} $	100·0 99·0 87·7 34·9	$\infty \\ 7.33 \\ 6.16 \\ 4.61$	7.6 7.2 6.3 4.6	$ \begin{array}{c c} $	$\begin{array}{c} 0.343 \\ 0.392 \\ 0.565 \\ 1.780 \end{array}$	7·94 7·31 6·15 4·61	$ \begin{array}{r} 10.199\\ 19.400\\ 58.800\\ 51.300\\ \hline 139.699 \end{array} $	$1:6121 \\ 3 \cdot 8024 \\ 19 \cdot 7750 \\ 84 \cdot 5500 \\ 109 \cdot 7395$	$ \begin{array}{r} 37 \cdot 318 \\ 70 \cdot 907 \\ 215 \cdot 250 \\ 218 \cdot 975 \\ \overline{542 \cdot 450} \\ \end{array} $	7.64 7.16 6.26 4.59
			٠.		··· 8		Root W.	214					
1.79 1.66 1.49 1.17 0.57	125 117 127 51 132	125 115 114 40 37	100-0 98-3 89-8 78-4 28-0	100-0 98-0 87-7 74-0 13-3	∞ 7·05 6·16 5·64 3·89	7·2 6·9 6·4 5·5 3·8	$ \begin{array}{r} 9.5 \\ 14.9 \\ 31.4 \\ 22.9 \\ 17.6 \\ \overline{96.3} \end{array} $	0-392 0-438 0-539 0-876 4-557	7.59 7.03 6.12 5.64 3.89	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	3.7240 6.5262 16.9246 20.0604 80.2032 127.4384	$\begin{array}{r} 72 \cdot 105 \\ 104 \cdot 747 \\ 192 \cdot 168 \\ 129 \cdot 156 \\ 68 \cdot 464 \\ \hline 566 \cdot 640 \end{array}$	7.256.896.415.523.84
	129	21	16.3	I]	I —	1 —	Control	ls I —	ı _	li — I		I	u
	. 213: . 214:	$\overline{x}_1 = 1$	·4417, ·3017,	$\overline{x}_1' = 1 \cdot 13$ $\overline{x}_2' = 1 \cdot 32$	25, 33,	1	$\bar{y}_1 = 5.5980$ $\bar{y}_2 = 5.8841$). I.		111	a.	I	
W. 213:		Snw 215-1 201-4	198	<i>Snwxx'</i> 135·6391 158·2095		Snwx' ² 163·7154 124·2803		Snwxy 820·907 782·040		Snwx'y 551-987 614-326		Sni 3147 3036	1.90
			182	- 22.5704		39.4351		38.867		- 62.339		• 111.24	
W	W. 214:		746 633	111.9036 165.8817		$396 \cdot 4995$ $168 \cdot 6453$		$779 \cdot 415$ $737 \cdot 573$		602·854 749·862		3454 3334	
		15.1	113	- 53.9781		227.8542		41.842	· .	-147.008		120).31
Co	Controls:		1.1	$n_c(1-C)/c$		$C = 629 \cdot 8235$			$n_c(c-C)/C = -$.5.312		$n_c(c-C)^2/C(1-C) = -0.04$		0.04
To	Totals:		295	- 76.5485		897	897.1128		-214.659			231.59	

TABLE 17. Toxicity of Derris Roots W. 213 and W. 214 to Oryzaephilus surinamensis

Ex. 17. Numerical example of the maximum likelihood solution.

Martin (1940, Table 5) has reported the results of tests of the toxicity of two derris roots, W. 213 and W. 214, to the grain beetle, Oryzaephilus surinamensis. Of 129 control insects, sprayed with the alcohol-sulphonated lorol medium but no derris, 21 were affected, so that c is $16\cdot3 \%$. The logarithms of the concentrations of the two roots, measured in milligrams of dry root per litre, are shown in the first column of Table 17. The columns n, r, p' contain the numbers of insects exposed, and affected, and the percentage affected respectively. The kills by the spray were substantially higher than $16\cdot3 \%$, and appear to give little additional information on C.

As a start to the computations, a provisional value of C (in fact a remarkably good guess) may be taken as 17.0 %. The percentage kills due to the poisons alone are then estimated by means of equation (6.2); for example, for the lowest concentration of root W. 213,

$$p = (0.460 - 0.170)/(1 - 0.170) = 0.349.$$

The empirical probits of p are plotted against x, and two parallel regression lines placed by eye so as to give two series of expected probits, Y. The weighting coefficients, read from Table II in the column for 17 %, are multiplied by n and the products entered in the column nw; thus, for the first line of Table 17, Y = 7.6, the weighting coefficient is 0.03298, and multiplication by 142 gives 4.7. The auxiliary variate, x', is read from Table II for the appropriate values of Y, and the working probits, y, are obtained as usual.

Product columns, nwx, nwx', nwy, are next formed, the totals of which lead to the weighted means $\bar{x}, \bar{x}', \bar{y}$, for each root. Sums of squares and products of deviations are calculated, and the contributions from the two roots added, with the further addition of (6.9)

 $129 \times 0.83/0.17 = 629.8235$

to $S_{x'x'}$, and (6.10)

 $129 \times (0.163 - 0.17)/0.17 = -5.312$

to $S_{x'y}$. Thus the combined evidence of the two roots gives equations (6.5) in the form

$$28 \cdot 8295b - 76 \cdot 5485 \left(\frac{\delta C}{1 - C}\right) = 80 \cdot 709,$$

- 76 \cdot 5485b + 897 \cdot 1128 $\left(\frac{\delta C}{1 - C}\right) = -214 \cdot 659.$ (6.12)

The first row of V, the inverse matrix of coefficients, is found by replacing the right-hand sides of equations (6.12) by 1, 0 and solving; similarly, the second row is found by solving with 0, 1 on the right. The result is

$$V = \begin{pmatrix} v_{11} & v_{12} \\ v_{12} & v_{22} \end{pmatrix} = \begin{pmatrix} 0.0448475 & 0.0038267 \\ 0.0038267 & 0.0014412 \end{pmatrix}$$
(6.13)

The solution of equations (6.12) is obtained by adding the products of their right-hand sides with each row of V in turn.

$$b = 80.709 \times 0.0448475 - 214.659 \times 0.0038267$$

= 2.7982, 1
$$\frac{\delta C}{1-C} = 80.709 \times 0.0038267 - 214.659 \times 0.0014412$$

= -0.00052.

By substitution of the provisional value of C,

$$\delta C = -0.00052 \times 0.83 \\ = -0.00043, |$$

and the revised estimate of the natural response rate is

$$C = 0{\cdot}17 - 0{\cdot}0004$$

$$= 16.96 \%$$
.

The mean values in Table 17 are then used in equation (6.6):

$$a_1 = 5 \cdot 5980 - 2 \cdot 7982 \times 1 \cdot 4417 + 0 \cdot 0005 \times 1 \cdot 1325 = 1 \cdot 564,$$

$$a_{\bullet} = 5.8841 - 2.7982 \times 1.3017 + 0.0005 \times 1.3233 = 2.242.$$

Hence the revised regression equations are

$$Y_{1} = 1.564 + 2.798x,$$

$$Y_{2} = 2.242 + 2.798x.$$
(6.14)

The revised estimate of C is almost identical with the provisional value, and expected probits calculated from equations (6.14), shown in the last column of Table 17, agree closely with those used in the first cycle; no second cycle of calculations is needed. By equation (6.11)

$$\chi^2_{161} = 5.64; \tag{6.15}$$

ten dose levels were tested, and four parameters have been estimated, leaving 6 degrees of freedom for χ^2 . Had χ^2 been large, the possibility that one or two classes with very small expectations were making unreasonably large contributions would have needed consideration. The expected probits from equations (6.14) would be converted into values of P, these used in equation (6.1) with C = 16.96 % to give P', and expectedfrequencies calculated in each group (cf. Ex. 7). In this instance, the smallness of the χ^2 in equation (6.15) would suffice to convince the experienced statistician that there is no serious deviation from parallelism. A proper test of parallelism requires the reconstruction of equations (6.12) so as to give separate regression coefficients for the two preparations; the information on C must still be pooled. The coefficients of the equations are easily read from Table 17:

$$13 \cdot 7182b_{1} - 22 \cdot 5704 \left(\frac{\delta C}{1-C}\right) = 38 \cdot 867,$$

$$15 \cdot 1113b_{2} - 53 \cdot 9781 \left(\frac{\delta C}{1-C}\right) = 41 \cdot 842,$$

$$(6 \cdot 16)$$

$$\cdot 22 \cdot 5704b_{1} - 53 \cdot 9781b_{2} + 897 \cdot 1128 \left(\frac{\delta C}{1-C}\right) = -214 \cdot 659.$$

The solutions (evaluation of an inverse matrix is unnecessary) are

$$b_1 = 2 \cdot 8302,$$

$$b_2 = 2 \cdot 7622,$$

$$\frac{\delta C}{1 - C} = -0.00187.$$

By equation (6.11), but including terms for b_1 and b_2 ,

$$\chi_{[5]}^2 = 5.61,$$

1 degree of freedom less than in equation (6.15) because an extra parameter has been estimated. The analysis of χ^2 in Table 18 shows that both the component for deviations from parallelism

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and that for residual heterogeneity are small enough to be attributed to random variation in the data.

TABLE 18. Analysis of χ^2 for Derris-Oryzaephilus surinamensis Experiment

Source of variation	D. F .	Sum of squares	Mean square
Parallelism of regressions · Residual heterogeneity	1 5	0·03 5·61	0·03 1·12
Total	6	5.64	_

The matrix V does not require multiplication by a heterogeneity factor, but gives directly the variances and covariances of b and $\delta C/(1-C)$. From the first diagonal element

V(b) = 0.04485,b = 2.798 + 0.212.

and from the second diagonal element

 $V(C) = 0.0014412 \times (0.83)^2,$ C = 16.96 + 3.15 %,

whence

The sizes of these standard errors, relative to the changes from the provisional C and expected probits to the values after the first cycle, again show a second cycle to be unnecessary. The remaining element of V, the covariance, is required in forming variances of quantities such as m or M. At the 5 % probability level $c = t^2 V(b)/b^2$

$$g = t^2 V(b)/b^2$$
$$= 0.022,$$

Variance formulae can therefore be used satisfactorily to express the precision of m or M. For the log LD 50 of either root

$$m = \overline{x} + \left(5 - \overline{y} + \frac{\delta C}{1 - C} \overline{x}'\right) / b.$$
 (6.17)

Equation (3.6) must be modified to allow for the extra term in (6.17), and becomes

$$V(m) = \frac{1}{b^2} \left\{ \frac{1}{Snw} + \overline{x}'^2 v_{22} - 2\overline{x}'(m - \overline{x}) v_{12} + (m - \overline{x})^2 v_{11} \right\}, \quad (6.18)$$

where v_{11} , v_{12} , v_{22} are as shown in equation (6.13). Hence

$$V(m_1) = \frac{1}{b^2} \left\{ \frac{1}{96 \cdot 9} + (1 \cdot 132)^2 \times 0.00144 + 2 \times 1.132 \times 0.214 \times 0.00383 + (0.214)^2 \times 0.04485 \right\}$$

= $\{0.01032 + 0.00185 + 0.00186 + 0.00205\} \div 7.830$
= 0.00205 .

Therefore $m_1 = 1.228 \pm 0.045$, and similarly $m_2 = 0.986 \pm 0.051$. The relative dosage value of the two roots is given by a modification of equation (5.1):

$$M = m_2 - m_1 = \bar{x}_2 - \bar{x}_1 - \left\{ \bar{y}_2 - \bar{y}_1 - \frac{\delta C}{1 - C} (\bar{x}_2' - \bar{x}_1') \right\} / b, \quad (6.19)$$

and for this quantity

$$V(M) = \frac{1}{b^2} \left\{ \frac{1}{{}_1Snw} + \frac{1}{{}_2Snw} + (\bar{x}'_2 - \bar{x}'_1)^2 v_{22} + 2(\bar{x}'_2 - \bar{x}'_1) \times (\bar{x}_2 - \bar{x}_1 - M) v_{12} + (\bar{x}_2 - \bar{x}_1 + M)^2 v_{11} \right\}.$$
 (6.20)

Substitution from Table 17 and equation (6.13) gives

M = -0.2423and V(M) = 0.00273:

The result, $M = -0.242 \pm 0.052$, may be compared with that given by Martin, $M = -0.248 \pm 0.042$, in his discussion of the same data. The difference in M is trivial; the present analysis shows a higher standard error because allowance for the natural mortality rate reduces the weighting coefficients. The correct technique was not known in 1940, when Martin published his paper. Fiducial limits to M are calculated by adding and subtracting 1.96 times the standard error, and thus are set at -0.344, -0.140: Root W. 213 is estimated to have a potency 57.3 % that of W. 214, and the true value is likely to lie between 45.3 and 72.4 %. Had g not been negligible, fiducial limits would have been obtained by application of formula (4.7) to equation (6.19).

Chapter 7

FACTORIAL EXPERIMENTS

29. REASONS FOR FACTORIAL DESIGN

THE introduction of the factorial principle into the planning of biological experimentation has been a revolutionary step which can now be seen as not merely useful, but essential for a full exploration of the causes underlying even the simplest biological phenomena. Fisher (1942, § 37), in an excellent chapter on the advantages of factorial experimentation, succinctly states the case for factorial design: 'In expositions of the scientific use of experimentation it is frequent to find an excessive stress laid on the importance of varying the essential conditions only one at a time.... This ideal doctrine seems to be more nearly related to expositions of elementary physical theory than to laboratory practice in any branch of research. In experiments merely designed to illustrate or demonstrate simple laws, connecting cause and effect, the relationships of which with the laws relating to other causes are already known, it provides a means by which the student may apprehend the relationship, with which he is to familiarise himself, in as simple a manner as possible. By contrast, in the state of knowledge or ignorance in which genuine research, intended to advance knowledge, has to be carried on, this simple formula is not very helpful. We are usually ignorant which, out of innumerable possible factors, may prove ultimately to be the most important, though we may have strong presuppositions that some few of them are particularly worthy of study. We have usually no knowledge that any one factor will exert its effects independently of all others that can be varied, or that its effects are particularly simply related to variation in these other factors. On the contrary, when factors are chosen for investigation, it is not because we anticipate that the laws of nature can be expressed with any particular simplicity in terms of these variables, but because they are variables which can be controlled or measured with comparative ease. If the investigator, in these

circumstances, confines his attention to any single factor we may infer either that he is the unfortunate victim of a doctrinaire theory as to how-experimentation should proceed, or that the time, material or equipment at his disposal are too limited to allow him to give attention to more than one narrow aspect of his problem.'

Factorial design has until now been most widely employed for agricultural field trials, but its value in the logical structure and interpretation of experiments is as great in the laboratory as in the field. Problems of toxicology have been investigated in the past chiefly by varying the level of a single factor in the set of conditions defining the stimulus, other factors being held as nearly constant as was practicable. Factorial design, on the other hand, entails, first, a selection of the more important factors relating to the stimulus or the subject; secondly, the adoption for the experiment of a convenient number of states or levels of each factor selected; and thirdly, the making of tests on batches of subjects under the conditions defined by various combinations of levels of these factors, non-experimental factors being held as nearly constant as possible. In this way the virtues of carefully standardized conditions are combined with the obtaining of information on the effects of variations in these conditions. The measurements of all factors constituting the stimulus may be referred to collectively as the dose.

In considering the desirability of adopting a factorial set of treatments, the different needs of an assay and an investigation into the laws determining the reaction of the subject to the stimulus should be borne in mind. The purpose of an assay is to assess the value of an arbitrary unit of the stimulus under test in terms of units of a standard stimulus; providing that the test stimulus can be fully described in these standard units, there will generally be no advantage in using several factors for the assay rather than only one. If, for example, an insecticidal spray whose only toxic constituent is rotenone is to be assayed in units of a standard rotenone preparation, the same result should be obtained (within the limits of sampling variation) from tests at different concentrations as from tests with

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different quantities of a fixed concentration, and the inclusion of different levels of other factors would have little advantage. Indeed, any indication that the result of the assay depended on other conditions (such as temperature or method of spraying) would contradict a basic assumption of the assay, and would suggest that the spray under test contained a toxic constituent whose potency could not be described simply in units of rotenone, since its equivalent in rotenone varied when experimental conditions were changed. The choice between varying the concentration and varying the quantity of spray would depend upon practical convenience and experience of which gave the more reliable results.

On the other hand, when the relationship between the reaction of the subject and the measures of the stimulus is the object of study, a factorial experiment may have many advantages. Before the action of an insecticide can be fully understood, the direct effects of various factors defining the dose and its method of application and the interactions between these must be investigated in detail. By comparison with agricultural experiments, laboratory tests of insecticides take only a short time to carry out, and, at least in the preliminary stages of a research project, a series of experiments on single factors may give better returns than one comprehensive experiment including many factors. Making use of the information gained from these simple trials, plans can be made for the more extensive factorial experiments which are essential to the elucidation of the interrelationships between the factors.

The discussion which follows is once again given in terms of insecticidal studies, but the applicability of the principles to other fields will be easily appreciated. Laboratory research on insecticidal potencies has been primarily directed at discovering the effect of variation in the concentration of the toxic substance on the mortality rate of the insects. To a lesser extent, the effect of variation in the duration of exposure to the poison has been examined, though published data from tests in which both factors have been varied are few. Other factors, such as the temperature or the quantity of poison used, have received even less attention.

30. QUALITATIVE FACTORS

In a well-planned toxicological investigation there will generally be at least one graded or quantitative factor, such as the concentration or amount of toxic substance, amongst the complex of factors measuring the dose. Methods of statistical analysis suitable for use when there are two or more quantitative factors are discussed in § 31 and subsequent sections. There may also be factors of a purely qualitative nature, such as variations in the medium in which the poison is applied (e.g. oil or water; Martin, 1943), or variations in the method of application of an insecticide (e.g. spray or film; Tattersfield and Potter, 1943). Other factors, though capable of being measured quantitatively, may simply be recorded descriptively, as in comparisons between 'warm' and 'cold' conditions during spraying or between 'old' and 'new' stocks of insecticide. A similar classification may be made of factors relating to the test subject (see §§ 51, 52).

The data from series of tests with the various combinations of conditions arising from several qualitative factors may most readily be analysed by the methods of Chapter 5 when there is only one quantitative factor, or by extensions of these when there are more. In the former case, if the probit regression lines for the quantitative factor are parallel for each combination, the effects of the qualitative factors may be measured simply by comparisons amongst their median lethal doses, and a factorial analysis of these will sort out the main effects and interaction. Yates (1937*a*) has discussed this type of factorial analysis, and, though some complication is introduced by the unequal precisions of the estimated log LD 50's, his work may be adapted to suit the present purpose. If the lines are not parallel, interpretation of the results is more difficult, but nothing need here be added to what has been said in § 25.

31. THE PROBIT PLANE

If the joint effects of two quantitative factors, such as the time of exposure to an insecticide and its concentration, are to be studied, batches of insects must be tested at various combinations of values of the two factors. The test conditions may be chosen

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as several different concentrations for each of a number of exposure times, or as several different exposure times for each of a number of concentrations. Greater symmetry may be attained by using all combinations of a set of concentrations and a set of exposure times (say 4 concentrations and 5 times used in all their 20 combinations); the more extreme combinations—low concentrations for short times, or high concentrations for long times—may be omitted if they seem unlikely to give useful results.

Experience has shown that not only is the mortality probit frequently linearly related to the logarithm of the concentration for a fixed time of exposure, but also it is frequently linearly related to the logarithm of the exposure time for a fixed concentration. As a representation of the joint effect of time and concentration, therefore, a plane may be suggested:

$$Y = \alpha + \beta_1 x_1 + \beta_2 x_2, \tag{7.1}$$

giving the probit in terms of the log concentration (x_1) and the log time (x_2) . Bliss (1940b) expressed this by saying that the logarithms of the exposure time and concentration required to give any particular kill are linearly related; he proposed to evaluate this relationship by selecting a level of mortality and estimating either the values of x_1 for a series of times or the values of x_2 for a series of concentrations, according as the experiment was performed in sets of tests with fixed exposure time and varied concentration or with fixed concentration and varied time. This method of analysis involves a great amount of tedious calculation and does not give the maximum likelihood estimates of the parameters, though the estimates obtained will usually be not very different from the maximum likelihood values.

The maximum likelihood estimation is, however, to be preferred (Finney, 1943*a*); not only is it a more symmetrical approach and a natural generalization of the one-factor analysis, but the computations, though still lengthy, are simpler and are more easily reduced to a routine process. The method applies multiple regression analysis (Fisher, 1944, § 29) to the estimation of the

coefficients β_1 and β_2 , and may be further extended to three or more factors with no difficulty other than the increase in the computations. In order to use the regression technique, a provisional plane must first be fitted to the empirical probits which have been read from Table 1 in the normal manner. If several concentrations have been tested at each of a series of times, but not necessarily the same set of concentrations for every time, lines may be drawn, by eye, relating the empirical probit to the log concentration for each time, with the restriction that all the lines shall be parallel and at distances apart proportional to the differences in the log times. If the structure of the experiment is such that a series of concentrations and a series of times are used in all, or nearly all, their combinations, a method adapted from a suggestion of Richards (1941) may be used. The probits are plotted against the sum of log concentration and log time, (x_1+x_2) ; points with constant x_2 should then lie on one set of parallel straight lines with slope b_1 , and points with constant x_1 on a second set of parallel lines with slope b_{\bullet} . The differences between pairs of lines of either set will be proportional to the differences between the corresponding values of x_2 or x_1 . In this way a plane representation of the three-dimensional figure relating probits to log concentration and log time is obtained, and expected probits can be read from two intersecting sets of parallel lines, drawn by eye, in the diagram. Whatever the means employed, it is worth taking some trouble over the drawing of provisional lines, in order that a satisfactory approximation to the maximum likelihood solution may be obtained by one cycle of the computations.

The working probit, y, and its weight, nw, may be derived from the expected probit and the percentage kill in precisely the same way as when only one factor is involved. Using the technique of multiple regression to derive a, b_1 and b_2 as estimates of the parameters, equation (7.1) is estimated just as was the one-factor probit equation in § 17; the details will be made clear by a careful study of Ex. 18. If this equation differs substantially from the provisional lines or plane, it may be used to determine a new series of provisional probits, with which the cycle of computations is then repeated. When there is an appreciable natural mortality amongst the controls the methods of adjustment discussed in Chapter 6 may be used; the approximate method, involving only a modification of the weighting coefficient, is directly applicable, and the full analysis described in § 28 may easily be extended to allow for an additional independent variate, x_2 .

The method of estimating the parameters, testing the goodness of fit of the equation, and determining standard errors will now be illustrated by a numerical example. The statistical technique was elaborated for the analysis of data obtained by Tattersfield and Potter (1943), and Ex. 18 is a revised version of an account of the computations appropriate to the results of one of their experiments; in the account as previously published (Finney, 1942b, 1943a) weighting coefficients were not modified to allow for the control mortality.

The effect of variation in concentration and deposit on Ex. 18. the toxicity of a pyrethrum preparation to Tribolium castaneum. Tattersfield and Potter (1943) have described a series of experiments on the toxicity of a solution of pyrethrum extract in heavy oil to the beetle, T. castaneum: the doses used consisted of all combinations of several concentrations of the pyrethrum extract and several weights of spray deposit on the glass disk on which the insects were placed. In the first experiment four concentrations (these have been measured in terms of pyrethrin I only) and three deposits were tested; the glass disk was covered with a loosely woven fabric, and each combination of concentration and deposit was used, on separate batches of insects, both as a direct spray and as a film on which the insects were afterwards placed. Batches of ten insects were used for each spraying, and all treatments were given in three-fold replication.

Insects were also exposed to different deposits of the base oil alone, applied both by the spray and by the film technique, but, as the mortalities gave no indication of being different from that amongst unsprayed controls, all control batches have been added. Of a total of 311 beetles, 12 were 'badly affected, moribund, or dead', giving a control rate of 3.9 %. The full data have been reported by the experimenters (loc. cit. Table 2); after adjustment for a control death rate of 4 %, the mortalities are as shown in Table 19.

An extension of the ideas of Chapter 5 suggests the fitting of two parallel probit planes to the data. Table 20 sets out full details of the lengthy, but straightforward, computations required for this, including the derivation, from the original observations, of the figures shown in Table 19. In order to avoid

TABLE 19.	Percentage Kills of Tribolium castaneum by a Pyre-
thrum Sp	ray, adjusted for 4 % Mortality amongst the Controls
(numbers	of insects shown in brackets)

Pyrethrin I concentra- tion (mg./ml.)	Deposit (mg./sq.cm.)							
		Direct spre	у	Film				
	0.29	0.57	1.08	0.29	0.57	1.08		
0.5	0	10	17	7	11	26		
	(27)	(29)	(30)	(29)	(27)	(28)		
1.0	50	64	61	31	48	59		
	(29)	(29)	(24)	(30)	(28)	(28)		
2.0	90	96	100	82	96	93		
	(30)	(27)	(31)	(29)	(28)	(28)		
4 ·0	100	100	100	100	100	100		
	(28)	(30)	(19)	(29)	(29)	(17)		

the occurrence of negative numbers, both concentrations and deposits have been multiplied by 10, so that x_1 and x_2 are each 1.00 in excess of the true log concentrations and log deposits.

These modified x_1 and x_2 values form the first two columns of Table 20, and are followed by n, the number of insects under any one treatment, r, the total killed, and p', the proportionate mortality. The control mortality is estimated with considerable accuracy from the insects untreated or treated with oil alone, and, as 4 % is low and not in conflict with the remainder of the data, the approximate method of § 27 has been used without any attempt to improve the estimate of C by means of the maximum likelihood equations. The column of adjusted mortalities, p, has therefore been obtained with C = 0.04 in Abbott's formula (equation (6.2)), and the empirical probits of p have been written down from Table 1.

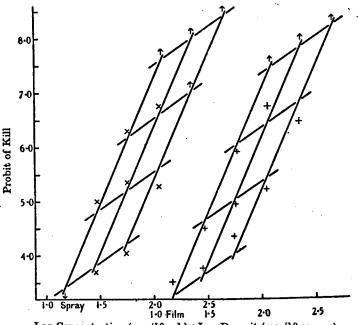
FACTORIAL EXPERIMENTS

		•										
<i>x</i> 1	<i>x</i> 3	n	•	p'	p(C=4)	Em- pirical probit	r	n 10	у	nwx ₁	nwrs	пку
	Exposure to direct spray									(
0·70 1·00 1·30 1·60	0·47 0·47 0·47 0·47	27 29 30 28	1 15 27 28	3.7 51.7 90.0 100.0	0 50 90 100	∞ 5.00 6.28 ∞	3·4 4·8 6·2 7·6	3-6 16-6 10-6 1-1	2·91 5·00 6·28 7·94	2·520 16·600 13·780 1·760		10.476 83.000 66.568 8.734
		1	ļ					31.9		34.660	14.993	168.778
0·70 1·00 1·30 1·60	0.75 0.75 0.75 0.75 0.75	29 29 27 30	4 19 26 30	13-8 65-5 96-3 100-0	10 64 96 100	3·72 5·36 6·75 20	3.8 5.2 6.6 8.0	7-9 17-0 6-1 0-4	3.72 5.36 6.73 8.30	5·530 17·000 7·930 0·640		29-388 91-120 41-053 3-320
	ľ	i						31-4		31.100	23.550	164.881
0·70 1·00 1·30 1·60	1.04. 1.04 1.04 1.04	30 24 ^31 19	6 15 31 19	20-0 62-5 100-0 100-0	17 61 100 100	4.05 5.28 co co	4·2 5·6 7·0 8·4	12·6 12·7 3·9 0·1	4-06 5-25 7-42 8-67	8·820 12·700 5·070 0·160		51·156 66·675 28·938 0·867
	•							29.3	· ·	26.750	30.472	147.636
		ļ					ļ	92.6		92.510	69-015	481-295
		•	•			Exposur	' e to fil	•	, ,	02 010		. 101 000
0.70 1.00 1.30 1.60	0·47 0·47 0·47 0·47 0·47	29 30 29 29	3 10 24 29	10-3 33-3 82-8 100-0	7 31 82 100	3.52 4.50 5.92 ∞	3·3 4·7 6·1 7·5	3·1 16·7 11·2 1·4	3·57 4·51 5·90 7·85	2·170 16·700 14·560 2·240		11.067 75-317 66-080 10-990
							•	32.4		35 670	15.228	163-454
0·70 1·00 1·30 1·60	0.75 0.75 0.75 0.75 0.75	27 28 28 29	4 14 27 29	14-8 50-0 96-4 100-0	11 48 96 100	3.77 4.95 6.75 ∞	3·6 5·0 6·4 7·8	5-4 16-5 8-1 0-7	3-80 4-95 6-67 8-12	3·780 16·500 10·530 1·120		20.520 81.675 54.027 5.684
								30.7		31.930	23.025	161-906
0·70 1·00 1·30 1·60	1.04 1.04 1.04 1.04	28 28 28 17	8 17 26 17	28.6 60.7 92.9 100.0	26 59 93 100	4·36 5·23 6·43 ∞	4·0 5·4 6·8 8·2	9·7 15·8 4·8 0·1	4·42 5·22 6·37 8·49	6·790 15·800 6·240 0·160		42.874 82.476 30.576 0.849
								30-4		28-990	31-616	156-775
								93.5		96-590	69.869	482.135
	Spray: $\bar{x}_1 = 0.9990$, $\bar{x}_2 = 0.7453$, $\bar{y} = 5.1976$. Film: $\bar{x}_1 = 1.0330$, $\bar{x}_2 = 0.7473$, $\bar{y} = 5.1565$.											
Spra y	, E	Snwx 97.019 92.420	00	Snwx ₁ x ₄ 67.43520 68.94792	56 - 40 51 - 4	0009 3704	Snwx 502.70 480.82	993 272	Snwx,y 356-5278 358-7103	2501	·423 ·565	
Film		4·598)4·279)9·782	00	-1.51272 70.86200 72.1780) 57-3		21-88 515-51 498-06	54	-2.1825 361.2989 360.2812	2566		•12 = 3•11
	-	4.496		-1.3160-		9610	17.4		1.0177		978 - 75	·65 = 4·33
Total		9.095	79	-2.82876	i 10-0	5915	39-33	288	-1.1648	192	·836 – 183	$\cdot 40 = 9 \cdot 41$

.

TABLE 20. Computations for Analysis of Results of Testing Various Concentrations of Pyrethrin on Tribolium castaneum

The empirical probits have been plotted against $(x_1 + x_2)$ in Fig. 11. Both for the direct spray and for the film technique two intersecting sets of parallel lines have been drawn, the one representing the regression of probits on x_1 for fixed x_2 and the other the regression on x_2 for fixed x_1 ; the lines were drawn by eye, remembering the existence of zero and 100 % kills at certain



Log Concentration (mg./10 ml.) + Log Deposit (mg./10 sq.cm.)

FIG. 11. Diagrammatic representation of probit planes for comparing potency of direct spray and film applications of a pyrethrum spray to *T. castaneum* (Ex. 18). \times direct spray; + film. Continuous lines show effect of change in concentration at a fixed deposit. Broken lines show effect of change in deposit at a fixed concentration.

doses, in such a way as to intersect vertically above or below the plotted points. The intersections give the expected probits, Y, appropriate to each combination of x_1 and x_2 , from which, by the methods of previous chapters, weights, nw, and working probits, y, were calculated; the weighting coefficients were taken from the 4 % column in Table II.

For either, technique of application of the insecticide, an approximation to the maximum likelihood estimate of the probit plane is derived as the weighted regression equation of y on x_1 and x_2 . In order to obtain two parallel planes, the regression coefficients must be calculated not from the sums of squares and sums of products of deviations for the techniques separately but from the totals of these (cf. § 20).

The next stage of the computations was therefore the formation of the products nwx1, nwx2 and nwy, the calculations proceeding independently for spray and for film; for nwx_2 it is sufficient to total the values of nw for each of the three deposits and multiply this total by the appropriate x_2 , but for nwx_1 and nwy individual products must be entered for each of the twelve tests and totalled in the three deposit groups. Totals of the columns Snw, $Snwx_1$, Snwx_a and Snwy were then made. By summing the products of each of the twelve values of nwx_1 with x_1 and deducting $(Snwx_1)^2/Snw$, the sum of squares of deviations of x_1 , denoted by $S_{x,x,i}$, was obtained. Similarly, multiplying each of the three subtotals of nwx_1 by the corresponding x_2 adding, and deducting $(Snwx_1)$ $(Snwx_2)/Snw$, the sum of products of deviations, $S_{x_1x_2}$, was obtained. In like manner all the sums of squares and products shown at the bottom of Table 20 for both spray and film methods have been computed, and the values for the two techniques have been added to give the last line of the table.

The regression coefficients for the two parallel planes are the solutions of the equations

$$9 \cdot 096b_1 - 2 \cdot 829b_2 = 39 \cdot 329,$$

 $- 2 \cdot 829b_1 + 10 \cdot 059b_2 = -1 \cdot 165.$

These should be solved by the inverse matrix method, as in Ex. 17, in order to obtain also the variances of parameters and other estimated quantities. Taking first values 1, 0 and then 0, 1 on the right-hand side, this matrix was obtained as

$$V = \begin{pmatrix} 0.12048 & 0.03388\\ 0.03388 & 0.10894 \end{pmatrix},$$
(7.2)

whence $b_1 = 39 \cdot 329 \times 0.12048 - 1.165 \times 0.03388$ = 4.6989, $b_2 = 1.2056.$ (7.3)

Now the fitting of the regression planes accounts for a portion

$$b_1 S_{x_1 y} + b_2 S_{x_2 y} = 183.40 \tag{7.4}$$

of the sum of squares of deviations of y, $S_{yy} = 192.84$. S_{yy} is based on two sets of 11 degrees of freedom, and the fitted parameters remove 2, leaving $\chi^2_{[20]} = 9.44$. This χ^2 has been analysed to provide a test of parallelism of the two planes. Regression coefficients were obtained for spray and film separately, using only the appropriate sections of the data of Table 20; the pairs of values are:

Spray
 Film

$$b_1 = 5.1277$$
 $b_1 = 4.2601$
 $b_2 = 1.1236$
 $b_2 = 1.2999$

TABLE 21. Test of Parallelism of Probit Planes for Ex. 18

	D.F.	Sum of squares	Mean square
Parallelism of planes Residual heterogeneity	2 18	2.00 7.44	1.00 0.41
Total	20	9.44	

The separate values of S_{uu} were then reduced by quantities calculated as in equation (7.4), namely, 109.75 and 75.65, to give residuals of 3.11 and 4.33 respectively, each being a $\chi^2_{(9)}$. Hence for a joint test of the heterogeneity of the data about two fitted planes (not now constrained to be parallel) $\chi^2_{(18)} = 7.44$; the difference between this and the previous $\chi^2_{(20)}$ is $\chi^2_{(21)} = 2.00$, which indicates no significant departure from parallelism. These results are summarized in Table 21. Had the heterogeneity χ^2 been large, the test of parallelism would have been based on the ratio of mean squares; the variances discussed below would then have been increased by the heterogeneity factor. Since the χ^2 values are all small, there is no need to give special consideration to possible excessive contributions from the tests at extreme dose levels, but in other circumstances the method of Ex. 7 could be employed for examining the separate contributions more carefully.

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The equation to either probit plane is of the form

$$Y = \bar{y} + b_1(x_1 - \bar{x}_1) + b_2(x_2 - \bar{x}_2). \tag{7.5}$$

Substituting the appropriate values of \bar{y} , \bar{x}_1 and \bar{x}_2 , the two fitted equations^{*} are:

$$\begin{split} Y_{s} &= -0.395 + 4.699 x_{1} + 1.206 x_{2}, \\ Y_{f} &= -0.598 + 4.699 x_{1} + 1.206 x_{2}. \end{split}$$

If the values of Y_g and Y_f are calculated for the twelve dosages used in the experiment they will be found to exhibit so good an agreement with the expected probits, Y, in Tuble 20 as to make it apparent that there is no necessity to repeat the cycle of computations; had a closer approximation to the solution of the maximum likelihood equations been required, Y_g and Y_f would have been used as the new expected probits.

The diagonal elements of the matrix V are the variances of b_1 and b_2 , and the remaining element is the covariance between these two parameters. Hence the standard errors of b_1 and b_2 are ± 0.347 and ± 0.330 respectively. The kill might have been expected to be independent of the separate values of concentration and deposit, within fairly wide limits, so long as the total amount of pyrethrin (the product of concentration and deposit) remained the same. This would imply that b_1 and b_2 should only differ by an amount consistent with their sampling variation, since increases in either concentration or deposit which were equal on the logarithmic scale would have equal effects on the kill. Such equality is clearly contradicted by the results, for the variance of the difference between b_1 and b_2 is

$$V(b_1 - b_2) = 0.1205 - 2 \times 0.0339 + 0.1089$$

= 0.1616,

and therefore $b_1 - b_2 = 3.493 \pm 0.402$.

In this experiment concentration has been a far more important factor than deposit in determining the kill. In fact a doubling

^{*} The equations may easily be put in terms of the logarithms of the concentrations and deposits given in Table 19 by replacing x_1 and x_2 by (x_1+1) and (x_2+1) respectively.

of the concentration was as effective as an increase of $(b_1 \log 2)/b_2$ in the log deposit, or a multiplication of the deposit by 14.9; the precision of this ratio is low, and 5 % fiducial limits determined by formula (4.7) from the variances and covariances in equation (7.2) are 5.9 to 279. The absorptive powers of the substratum are undoubtedly important in determining the relative effects of changes in concentration and in deposit: in further experiments with the same or a similar fabric covering the glass disk, Tattersfield and Potter again found concentration to have the greater effect, but when a hardened filter paper was used deposit became the more important factor.

In multifactorial experiments there is no unique median lethal dose; for example, with the direct spray any pair of values of x_1 and x_2 satisfying

$$4 \cdot 699x_1 + 1 \cdot 206x_2 = 5 \cdot 395$$

is estimated to give a 50 % kill. Similarly the relative potency of the direct spray and the film technique cannot be uniquely defined, for the inequality of b_1 and b_2 implies a difference in relative potencies in respect of concentration and of deposit. The relative dosage value, or difference between equally effective dosages, may be taken as any pair of values satisfying

$$4 \cdot 699M_1 + 1 \cdot 206M_2 = 0 \cdot 203.$$

If equal concentrations were used, log deposits 0.168 less for the spray than for the film would be expected to show equal kills, and, if equal deposits were used, equal kills would be expected when the log concentration was 0.043 less for the spray than for the film.

The mean probit difference (§ 24) was introduced (Finney, 1943*a*) in order to provide a single measure of the difference between two parallel probit planes. In the present experiment the value of this quantity is, by the obvious extension of equation $(5\cdot12)$,

$$\begin{aligned} \mathcal{A}_{sf} &= Y_s - Y_f \\ &= 0.203 \pm 0.147, \end{aligned}$$

FPA

the variance having been obtained as

$$V(\Delta) = \frac{1}{{}_{s}Snw} + \frac{1}{{}_{f}Snw} + (\bar{x}_{1s} - \bar{x}_{1f})^{2} V(b_{1}) + 2(\bar{x}_{1s} - \bar{x}_{1f}) (\bar{x}_{2s} - \bar{x}_{2f}) \operatorname{Cov} (b_{1}, b_{2}) + (\bar{x}_{2s} - \bar{x}_{2f})^{2} V(b_{2}) \quad (7.6) = 0.0108 + 0.0107 + (0.0340)^{2} \times 0.1205 + 2 \times 0.0340 \times 0.0020 \times 0.0339 + (0.0020)^{2} \times 0.1089 = 0.0216.$$

32. OSTWALD'S EQUATION

If equation (7.1) be considered as expressing a relationship between values of x_1 and x_2 which give a selected kill, it may conveniently be put in the form

$$\lambda_1^{\beta_1} \lambda_2^{\beta_2} = \text{constant},$$

 λ_1 and λ_2 being the absolute (not logarithmic) measures of dose, or

$$\lambda_1^{\beta_1/\beta_1}\lambda_2 = \text{constant.}$$

This equation has been used by Busvine (1938) and others, in the form $h_{1} = h_{2}$ (7.7)

$$\lambda^n t = k, \tag{7.7}$$

where λ is the concentration, t the time of exposure, n and k constants, as an empirical law relating the concentration and time required for a given toxic effect. Bliss (1940b) pointed out that this was a particular case of the equation

$$(\lambda - \lambda_0)^n t = k, \tag{7.8}$$

which was used by Ostwald and Dernoschek (1910) in a discussion of the relationship between adsorption and toxic effect, λ_0 being a threshold concentration below which no effect takes place.

Bliss has only discussed experiments in which the time taken to reach 100 % kill at selected concentrations was measured. This is not usually a convenient method of studying time-concentration relationships, and is liable to give results subject to wide variation, since the time measurement is determined by the most extreme member of the batch. The more satisfactory

method is to expose batches of test subjects to concentrations and for times chosen by the experimenter and to measure the mortality in each batch. A threshold concentration is unlikely to be known in advance, and would therefore have to be estimated from the data. The estimation would then become a more troublesome process than Bliss suggests, but even though the data require the obtaining of a probit equation in the form

$$Y = a + b_1 \log (\lambda - \lambda_0) + b_2 \log t,$$

rather than a direct relationship between λ and t, the principle used by him may reasonably be adopted. This entails calculating the regression equation for several values of λ_0 , but including also a quadratic term in $\{\log (\lambda - \lambda_0)\}^2$; λ_0 is finally estimated, by interpolation, as that value which makes the coefficient of the quadratic term zero. This is not the maximum likelihood estimate of λ_0 but should be a satisfactory value for practical purposes. The maximum likelihood equations have not been considered and are undoubtedly very complicated.

Fortunately, an adjustment to the concentration seldom appears to be needed. If the threshold concentration regularly differed from zero to any important extent in concentration-time tests, the same difference would presumably occur in tests carried out for a fixed time. Hence the adjustment would also be needed⁺ in one-factor experiments such as have been considered in earlier chapters, and x in equation (3.2) would have to be taken as $\log (\lambda - \lambda_0)$ instead of $\log \lambda$, λ_0 being an additional parameter to be estimated from the data. The fact that in so many experiments equation (3.2) is adequate for the description of the results seems evidence against the need of any allowance for threshold concentration. Nevertheless, some cases of data showing curvature of the relationship between probits and log concentrations might be simplified if a regression equation of the form

$$Y = \alpha + \beta \log \left(\lambda - \lambda_0\right) \tag{7.9}$$

were used instead of the more usual

$$Y = \alpha + \beta \log \lambda.$$

33. THE INTERACTION OF TWO FACTORS

The linear regression equation (7.1) implies that the effects of the two dosage factors on the mortality probit are independent and additive. If a concentration-time experiment were carried out in batches at fixed concentrations with varied times of exposure, the relationship of probit and log time might be linear for each concentration without the lines for the different concentrations necessarily being parallel. Such a situation might indicate heterogeneity of the material, or changes in the experimental conditions between the tests of different concentrations, but it might alternatively imply a true dependence of the slope of the lines upon the concentration. Bliss (1940b) has suggested that, as a first approximation, the standard deviation of the log-time tolerances (which is the reciprocal of the slope) might be expressed as a linear function of the log concentration, a method which leads to a final regression equation of Y on x_1 (log concentration) and x_2 (log time) in the form

$$Y = \frac{\alpha + \beta x_1 + x_2}{\alpha' + \beta' x_1}.$$
 (7.10)

Equation (7.10) is linear in x_2 , so that for any fixed concentration the relationship between Y and x_2 is given by a straight line, but it is not linear in x_1 . It therefore differs fundamentally from the usual findings in single-factor experiments that Y is linearly related to the log concentration. The equation

$$Y = \alpha + \beta_1 x_1 + \beta_2 x_2 + \beta_{12} x_1 x_2 \tag{7.11}$$

seems preferable to (7.10), being linear in x_1 when x_2 is held constant and vice versa.^{*} The slope of the regression line of mortality probit on either x_1 or x_2 increases (decreases if β_{12} is negative) as the other increases. The coefficient β_{12} measures the *interaction* between the two factors, or the extent to which the increase in Y for unit increase in x_1 or x_2 exceeds that predicted by the purely additive equation (7.1).

* This surface, which reduces to a plane if $\beta_{12} = 0$, is known as an hyperbolic paraboloid; sections by planes $x_1 = \text{constant}$ or $x_2 = \text{constant}$ are generators (straight lines), and sections by planes Y = constant are rectangular hyperbolas.

THE INTERACTION OF TWO FACTORS

The technique of fitting equation $(7\cdot11)$ to experimental data presents no difficulty. The first step is to form the products x_1x_2 for each dosage level. These are then treated as the measure of a third factor of the dosage, x_3 , the subsequent procedure being exactly as in Ex. 18. A multiple regression equation on x_1 , x_2 and x_1x_2 is eventually obtained in the form

$$Y = \bar{y} + b_1(x_1 - \bar{x}_1) + b_2(x_2 - \bar{x}_2) + b_{12}(x_1x_2 - \bar{x}_1x_2).$$

It should be noted that $\overline{x_1x_2}$ is the mean value of the product x_1x_2 , and is different from $\overline{x}_1\overline{x}_2$, the product of the mean values of x_1 and x_2 ; $\overline{x_1x_2} = Snwx_1x_2/Snw$. A more complex equation such as (7.11) should not be used in preference to (7.1) unless the data clearly require it as an adequate representation of the facts, or unless there is strong a priori evidence of its appropriateness.

Ex. 19. The effect of variations in concentration of hydrocyanic acid and exposure time on the mortality of Calandra granaria. Peters and Ganter (1935) tested the toxicity of hydrocyanic acid to C. granaria at seven different concentrations, using batches of ten insects and two to five different exposure times for each concentration. The results, from 270 insects in all, are given in the first three columns of Table 22 and suggest that the slope of the mortality-time regression lines decreases with increasing concentration. Bliss (1940b) showed the calculations for fitting equation (7.10), but did not examine whether in fact (7.1) might not be equally satisfactory.

When equation (7.1) was fitted to the data, the result was

$$Y = -6.15 + 6.10x_1 + 6.29x_2, \qquad (7.12)$$

with a residual $\chi^2_{(24)} = 53 \cdot 10$; here x_1 is the log concentration in g./cu.m., and x_2 is the log exposure time in hours. Thus discrepancies from this equation show heterogeneity, with a heterogeneity factor of 2.21. Since only ten insects were used in each batch, the expected numbers of dead and surviving are necessarily small, but, as pointed out in § 18, this is less likely to disturb the χ^2 distribution than the occurrence of a few isolated cases of small expectations. Detailed examination of the separate contributions

to χ^2 suggests that, though one or two are large, heterogeneity would remain after any reasonable grouping of the data. As the example is only given here in order to illustrate the technique

Log con- centration (g./cu.m.)	Log exposure time (hours)	% kill (p)	Empirical probit	Y	
1.477	0.176	10	3.72	4 ·2	
	0.301	40	4.75	5.0	
	0.477	50	5.00	6.0	
	0.544	100	∞	6·4	
1.380	0.398	60	5.25	4 ·8	
	0.477	70	5.52	5.3	
	0.544	80	5.84	5-7	
	0.602	90	6.28	6.1	
	0.653	100	00	6·4	
1.190	0.544	50	5.00	4 ·3	
	0.778	80	5.84	5-9	
	0.903	100	00	6.7	
1.061	0.699	40	4.75	4.5	
	0.778	70	5.52	5 ∙0	
	0.845	90	6-28	5.5	
	1.000	100	00	6.2	
0.929	0.903	0	00	· 5·0	
	0.954	40	4.75	5·4	
	1.000	70	5.52	5.7	
	1.079	80	5.84	6·2	
	1.176	100	so l	6.9	
0.778	1.204	60	5-25	6-2	
	1.255	100	00	6.6	
0.544	1.204	20	4.16	4 ·8	
	1.255	80	5.84	$5 \cdot 2$	
	1.301	90	6-28	5.6	
	1.398	100	00	6.3	

 TABLE 22. Results of Tests of the Toxicity of Hydrocyanic

 Acid to Calandra granaria

At every dose n = 10.

The first three columns of the table are taken from Table XII of Bliss (1940b).

for fitting the regression equation, this point will not be considered further. The variance matrix

$$V = \begin{pmatrix} 0.764 & 0.693 \\ 0.693 & 0.699 \end{pmatrix}$$
 (7.13)

must be multiplied by the heterogeneity factor to give the variances and covariance of the regression coefficients, whence

$$b_1 = 6 \cdot 10 \pm 1 \cdot 30, \quad b_2 = 6 \cdot 29 \pm 1 \cdot 24.$$

In view of the apparent decrease in the slope of separate x_3 regressions with increasing values of x_1 , equation (7.11) has also been fitted to the data. Details of the computations will not be given here as they are so very similar to Ex. 18. A first set of expected probits was taken from the column headed 'Second expected probit' in Bliss's Table XII, since these values were readily available, but usually this first set would be most easily obtained from a sketch or from a simpler equation such as (7.12). Working probits and weights were found in the usual manner, and were used to calculate a weighted regression on x_1 , x_2 and the product x_1x_2 . The resulting equation was used to give a new set of expected probits. In all, three cycles of the computations were carried out; the expected probits for the last of these are shown as Y in Table 22. The final equation is

$$Y = -9.13 + 8.31x_1 + 8.86x_2 - 1.90x_1x_2, \tag{7.14}$$

the inverse matrix leading to the variances and covariances being

$$V = \begin{pmatrix} 2 \cdot 324 & 2 \cdot 494 & -1 \cdot 305 \\ 2 \cdot 494 & 2 \cdot 782 & -1 \cdot 511 \\ -1 \cdot 305 & -1 \cdot 511 & 1 \cdot 089 \end{pmatrix}.$$
 (7.15)

The χ^2 has been calculated as in Ex. 18, by means of

$$\chi^2 = S_{yy} - b_1 S_{x_1y} - b_2 S_{x_2y} - b_{12} S_{(x_1x_2)y}$$

This gives $\chi^2_{[23]} = 51.27$, showing departures from the regression equation still to be heterogeneous, with a value of 2.23 for the heterogeneity factor. The variance of b_{12} is $2.23 \times 1.089 = 2.428$, so that

$$b_{12} = -1.90 \pm 1.56.$$

The parameter b_{12} does not significantly exceed its standard error, so that no great advantage in the representation of the data arises from using equation (7.14) instead of (7.12); in other words, the data do not seriously contradict the hypothesis that $\beta_{12} = 0$.

The values of b_1 and b_2 in equation (7.14) are not directly comparable with those in (7.12). In equation (7.14) the slope of

the regression on x_1 for fixed x_2 depends upon the value of x_2 and in that equation b_1 represents the slope when $x_2 = 0$; when x_2 has its mean value, the slope is

$$b_1 + b_{12} \bar{x}_2 = 6.79.$$

Similarly when x_1 has its mean value, the slope of the regression on x_2 is $b_2 + b_{12}\bar{x}_1 = 6.82$.

These quantities are much closer to the b_1 and b_2 of equation (7.12). The variances of b_1 and b_2 obtained from the appropriate entries in the matrix (7.15) are considerably higher than the variances of b_1 and b_2 in equation (7.12), but this is of no consequence since the parameters are very different in meaning.

Since for the parameters of equation (7.12)

$$V(b_1 - b_2) = 2 \cdot 21 \times (0 \cdot 764 - 2 \times 0 \cdot 693 + 0 \cdot 699)$$

= 0 \cdot 170,

and therefore

$$b_1 - b_2 = -0.19 \pm 0.41,$$

there is no significant difference between the two regression coefficients, and a regression equation with $(x_1 + x_2)$ as the single independent variate would represent the data almost as satisfactorily as equation (7.14). Hence, in this experiment, the toxic effect of the hydrocyanic acid may be expressed purely in terms of the number of 'grams-per-cubic-metre hours', the product of the concentration and the time of exposure. Taking the sum of x_1 and x_2 as a new variate, x, the methods of § 17 would lead to the regression equation, which may be derived with sufficient accuracy from sums of squares and products already computed as

$$Y = -6.32 + 6.27(x_1 + x_2),$$

with

$$\chi^{2}_{[25]} = 53.57$$

$$b = 6.27 \pm 1.22.$$

and

34. EXTENSIONS TO SEVERAL FACTORS

The methods outlined in this chapter may easily be applied to data relating to more than two dosage factors, though, as always in multiple regression analysis, the amount of computing involved increases rapidly with increasing number of factors. For three factors, equation $(7\cdot1)$ may be extended to the form

$$Y = \alpha + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_3, \qquad (7.16)$$

and, though expected probits are less easily obtained, the estimation of the parameters follows the same procedure as before. On the analogy of equation (7.11), a more general equation, linear in each factor separately but allowing for interactions between them, is

$$Y = \alpha + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_3 + \beta_{12} x_1 x_2 + \beta_{13} x_1 x_3 + \beta_{23} x_2 x_3 + \beta_{123} x_1 x_2 x_3, \qquad (7.17)$$

in which the regression coefficient β_{12} measures the interaction between the first two factors, and β_{123} measures the three-factor interaction. To fit such an equation to experimental results, the four products x_1x_2 , x_1x_3 , x_2x_3 and $x_1x_2x_3$ are treated as though they measured separate factors, and a seven-variate regression equation is calculated.

As in the unifactorial analysis, the cycle of computations for fitting a regression equation is repeated until satisfactory agreement with the maximum likelihood estimates is obtained, as shown by the agreement between successive sets of expected probits. Careful choice of the first set will often ensure that one or, at most, two cycles suffice. Tests of heterogeneity and of parallelism are easily made, and standard errors of parameters are derived from the inverse matrix used in solving the equations for the estimates. The notion of relative potency, so useful in tests of one factor, has no simple multifactorial analogue, but in all cases the mean probit difference can be used for comparing series of parallel results; 'parallel' is here used with the meaning that equations such as (7.16) or (7.17) can adequately describe the data, with the restriction that corresponding regression coefficients, β , shall be the same for every series and thus that the equations for the several series shall differ only in their values of α .

Chapter 8

THE TOXIC ACTION OF MIXTURES OF POISONS

35. Types of Joint Action

Any attempt to understand fully the toxic action of a group of insecticides or fungicides must ultimately involve a study of their behaviour when two or more are applied in mixture. In some cases the potency of a mixture may be greater than would be expected simply from a knowledge of the potencies of the constituents separately, a result which is clearly of practical importance in the economic utilization of the poisons; the opposite situation of reduced potency of a mixture by comparison with that of its constituents may also occur. Precise meaning can only be given to these modes of action of mixtures after the establishment or definition of a normal mode of action; the results of any series of tests may then be compared with this standard in order to judge whether the toxicity is enhanced or reduced when the poisons are applied in mixture.

The first systematic discussion of this topic in relation to probit analysis was given by Bliss (1939a) in a paper on 'The toxicity of poisons applied jointly'. He distinguished three types of joint toxic action, independent, similar and synergistic, whose properties he described as:

'(1) Independent joint action. The poisons or drugs act independently and have different modes of toxic action. The susceptibility to one component may or may not be correlated with the susceptibility to the other. The toxicity of the mixture can be predicted from the dosage-mortality curve for each constituent alone and the correlation in susceptibility to the two poisons; the observed toxicity can be computed on this basis whatever the relative proportions of the components.

(2) Similar joint action. The poisons or drugs produce similar but independent* effects, so that one component can be

* An unfortunate choice of word, since the meaning is entirely different from that in the previous paragraph.

substituted at a constant proportion for the other; variations in individual susceptibility to the two components are completely correlated or parallel. The toxicity of a mixture is predictable directly from that of the constituents if their relative proportions are known.

'(3) Synergistic action. The effectiveness of the mixture cannot be assessed from that of the individual ingredients but depends upon a knowledge of their combined toxicity when used in different proportions. One component synergizes or antagonizes the other.'

Bliss explicitly excluded from this classification cases of two constituents which react chemically to form a new compound. In one sense he may be considered to have covered all other types of joint action, independent and similar action being the simplest, and synergism including all forms of departure from the normal. *Antagonistic action*, in which the potency of a mixture is less than expected, has been described by Clark (1937, Chapter 17), who suggested various mathematical representations of it; his examples, however, are of so different a nature as scarcely to fall within the scope of Bliss's paper or the present work. Antagonism will be treated here simply as negative synergism.

The potency of a mixture whose constituents act similarly is generally greater than that of a mixture, in the same proportions, whose constituents are of the same individual potencies but act independently (§ 38). Either type of action is specified by an exact law predicting the kill produced by a mixture from the amounts of the constituents and their potencies. Hence a more exact definition of synergism is needed than Bliss's statement of its being 'characterized by a toxicity greater than that predicted from experiments with the isolated constituents'; at least it must be decided whether independent, or similar, or some other jointaction law is the norm to which any suspected case of synergism is to be referred. Bliss suggested two alternative mathematical models for synergistic action, but neither of these nor any of those used by Clark has the more familiar concepts of independent or similar action as special cases of zero synergism.

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Finney (1942a) has endeavoured to bring out the logical relationship between different types of joint action. That of chief importance in studies of insecticides and fungicides appears to be similar action, which will therefore be first considered here.

36. SIMILAR ACTION

As has been said in § 20, two poisons whose modes of action on the test organism are much alike, especially poisons of related chemical constitutions, often show parallel regression lines of mortality probits on log doses. The relative potency can then be expressed by a single figure, the ratio of equally effective doses, which is a constant at all levels of mortality. If the two regression lines are written as

$$Y_1 = a_1 + b \log \lambda, \tag{8.1}$$

$$Y_2 = a_2 + b \log \lambda, \tag{8.2}$$

where λ is the dose, the potency of the second poison relative to that of the first is given by

$$\log \rho_2 = (a_2 - a_1)/b, \tag{8.3}$$

so that (8.2) may alternatively be written

 $Y_1 = a_1 + b \log(\rho_2 \lambda).$

Multiplication by the factor ρ_2 converts doses of the second poison into equivalent doses of the first, the kill then being predictable by means of equation (8.1). A mixture containing amounts λ_1 , λ_2 of the two poisons is said to show similar action if, within the limits of sampling variation, the kill is the same as that which would be produced by a dose of the first equal to the sum of λ_1 and $\rho_2 \lambda_2$; thus similar action requires that the probit regression line for a mixture shall have the form

$$Y = a_1 + b \log (\lambda_1 + \rho_2 \lambda_2). \tag{8.4}$$

If the mixture is applied as a total dose, λ , in which the proportions of the two poisons are π_1 , π_2 , equation (8.4) may conveniently be rewritten as

$$Y = a_1 + b \log(\pi_1 + \rho_2 \pi_2) \lambda.$$

Though it is not necessary that $\pi_1 + \pi_2 = 1$, since λ may be measured in terms of some preparation containing also a proportion of inactive materials, equations (8.1) and (8.2) can be obtained by taking $\pi_1 = 1$, $\pi_2 = 0$ and vice versa. The potency of the mixture relative to that of the first poison is

$$\rho = \pi_1 + \rho_2 \pi_2, \tag{8.5}$$

and $(\pi_1 + \rho_2 \pi_2)\lambda = \lambda_1 + \rho_2 \lambda_2$ is the expression of a dose, λ , of the mixture as an equivalent, or equally effective, dose of the first poison. From these relationships it follows that if Λ_1 , Λ_2 are the ED50's for the two poisons, $\Lambda_2 = \Lambda_1/\rho_2$ and, more generally, for any mixture the ED50 is estimated as

$$\Lambda = \Lambda_1 / (\pi_1 + \rho_2 \pi_2). \tag{8.6}$$

The equations (8.4) and (8.6) are the same as (5) and (7) of Bliss's paper, except for the changed notation.

The concept of similar action and of equivalent doses is easily extended to three or more poisons, provided that all may be taken to have the same probit-dosage regression coefficient, b. If a mixture contains quantities λ_1 , λ_2 , λ_3 of three such poisons, the second and third having potencies ρ_2 , ρ_3 relative to the first, the equivalent dose is $(\lambda_1 + \rho_2 \lambda_2 + \rho_3 \lambda_3)$ and the regression line for the mixture

$$Y = a_1 + b \log (\lambda_1 + \rho_2 \lambda_2 + \rho_3 \lambda_3). \tag{8.7}$$

If the regression equations for two poisons which exhibit similar action and for a mixture of the two in proportions $\pi_1: \pi_2 (\pi_1 + \pi_2 = 1)$ are written as

$$Y_1 = a_1 + bx, \quad Y_2 = a_2 + bx, \quad Y_3 = a_3 + bx,$$
 (8.8)

the potency of the second poison relative to the first is estimated as $\rho = 10^{M}$, where

$$M = (a_2 - a_1)/b;$$

the third regression equation must then be equivalent to

$$Y'_{3} = a_{1} + b \log (\pi_{1} + \rho \pi_{2}) + bx, \qquad (8.9)$$

apart from sampling errors in the estimation of the four parameters a_1, a_2, a_3, b .

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Ex. 20. Similar joint action of rotenone and a deguelin concentrate. Data from tests of rotenone, a deguelin concentrate, and a mixture of the two in the proportion of 1:4 have been examined in Ex. 10, and have been found to be satisfactorily fitted by the three parallel probit lines shown as equations (5.8): $Y_r = 2.336 + 3.891x$, $Y_d = 0.660 + 3.891x$, $Y_m = 1.302 + 3.891x$. From the first two of these, the potency of the deguelin concentrate relative to that of rotenone is assessed, as in Ex. 10, at

$$\rho_d = 10^{-0.4307} = 0.371.$$

If the two poisons act similarly, a dose λ of the mixture will be as effective as a dose

$$(\pi_r + \rho \pi_d) \lambda = (0.2 + 0.371 \times 0.8) \lambda = 0.497 \lambda$$

of rotenone, from which the regression line for the mixture may be predicted as

$$Y'_{m} = 2 \cdot 336 + 3 \cdot 891 \log 0 \cdot 497 + 3 \cdot 891x$$

= 1 \cdot 155 + 3 \cdot 891x.

Comparison of this prediction with the equation estimated directly from the data shows the toxicity of the mixture to be a little greater than that required by the hypothesis of similar action. The logarithm of the observed potency relative to that predicted is

$$M_s = (1.302 - 1.155)/3.891$$

= 0.0378;

thus the mixture is estimated to be 9% more toxic than if the constituent poisons acted similarly. This apparent slight synergistic effect is shown in Ex. 21 to be within the limits of sampling variation, and no significance need be attached to it.

The difference between the third equation of (8.8) and equation (8.9), the probit-dosage relationship estimated directly from the data and the prediction according to the hypothesis of similar action respectively, is a mean probit difference measuring the amount of any enhancement or reduction of effectiveness, or, in general terms, a measure of synergism; it is

$$\Delta_{s} = a_{3} - a_{1} - b \log(\pi_{1} + \rho \pi_{2}). \qquad (8.10)$$

The variance of Δ_s has a complicated expression, but is not : difficult to calculate from quantities already used in estimating the various parameters:

$$\begin{split} V(\Delta_s) &= \frac{1}{(\pi_1 + \rho \pi_2)^2} \bigg[\frac{\pi_1^2}{{}_1 Snw} + \frac{\rho^2 \pi_2^2}{{}_2 Snw} + \frac{(\pi_1 + \rho \pi_2)^2}{{}_3 Snw} \\ &+ \frac{\{\pi_1 \overline{y}_1 + \rho \pi_2 \overline{y}_2 - (\pi_1 + \rho \pi_2) (\overline{y}_3 - \Delta_s)\}^2}{b^2 \Sigma S_{xx}} \bigg], \quad (8.11) \end{split}$$

suffices indicating the data for the three preparations tested, and Σ indicating summation for the three. As always, the variance must be multiplied by the heterogeneity factor when this is significantly greater than unity. Comparison of Δ_s with its standard error, the square root of the expression in (8.11), gives a test of the significance of the departure from the similar action prediction; positive values of Δ_s indicate synergism, negative antagonism.

The quantity
$$M_s = \frac{a_3 - a_1}{b} - \log(\pi_1 + \rho \pi_2)$$
 (8.12)

is the logarithm of the ratio of the observed potency of the mixture to that predicted on the hypothesis of similar action. Since

 $M_s = \Delta_s/b$,

the sign of M_s also shows whether the departure from similar action is in the direction of synergism or of antagonism, but the test of significance should be made on Δ_s rather than on M_s . Should the standard error of M_s also be required, it may be obtained from the variance

$$V(M_s) = \frac{1}{b^2 (\pi_1 + \rho \pi_2)^2} \left[\frac{\pi_1^2}{{}_1 Snw} + \frac{\rho^2 \pi_2^2}{{}_2 Snw} + \frac{(\pi_1 + \rho \pi_2)^2}{{}_3 Snw} + \frac{\{\pi_1 \overline{y}_1 + \rho \pi_2 \overline{y}_2 - (\pi_1 + \rho \pi_2) \overline{y}_3\}^2}{b^2 \Sigma S_{xx}} \right]. \quad (8.13)$$

Exact formulae for the fiducial limits of M_s and Δ_s have not been developed, but, unless g (as defined in § 19) is large, limits calculated from the standard error should be sufficiently reliable

for Δ_s and probably also for M_s . The error committed in assessing the fiducial limits of M_s as though the estimate were normally distributed will usually be less serious than in similar estimations of fiducial limits for median lethal dosages or relative dosage values; on account of the greater number of tests, the accuracy of estimation of b will be increased and g will therefore be reduced.

Ex. 21. Standard errors of discrepancies between observation and similar action predictions. The mean probit difference between the regression equation for the rotenone and deguelin concentrate mixture discussed in Ex. 20 and the equation predicted on the hypothesis of similar action is

$$\Delta_s = 1.302 - 1.155 = 0.147.$$

Also, using numerical values found in Ex. 10,

$$\frac{\pi_r \bar{y}_r + \rho \pi_d \bar{y}_d}{\pi_r + \rho \pi_d} - \bar{y}_m = -0.403,$$

and therefore, from equation (8.11), remembering that no evidence of heterogeneity was found when these data were analysed in Ex. 10,

$$V(\varDelta_{s}) = \frac{(0 \cdot 2)^{2}}{119 \cdot 6 \times (0 \cdot 497)^{2}} + \frac{(0 \cdot 297)^{2}}{78 \cdot 1 \times (0 \cdot 497)^{2}} + \frac{1}{74 \cdot 4} + \frac{(0 \cdot 403 - 0 \cdot 147)^{2}}{15 \cdot 138 \times 10 \cdot 467}$$
$$= \frac{0 \cdot 0014638}{0 \cdot 24701} + 0 \cdot 01344 + \frac{0 \cdot 065536}{158 \cdot 45}$$
$$= 0 \cdot 01978.$$

Hence

$$\Delta_s = 0.147 \pm 0.141,$$

a value which is not significantly different from zero and which therefore shows that the indication of synergism is within the limits of sampling variation.

The calculations for $V(\Lambda_s)$ have been shown in some detail, as by arranging them in this way some steps can be used again in calculating the variance of the log ratio of potencies, M_s .

From equation (8.13)

$$V(M_s) = \left[\frac{0.0014638}{0.24701} + 0.01344 + \frac{(0.403)^2}{158.45}\right] / 15.138$$

= 0.001347.

The value of M_s has already been found in Ex. 20, whence

$$M_s = 0.0378 \pm 0.0367.$$

Fiducial limits to M_s are approximately, for 5% probability, $0.0378 \pm 1.96 \times 0.0367$ or -0.0341 and 0.1097, so that the true potency of the mixture may be expected to lie between 92.4and 128.7% of the similar action prediction.

The test of significance of Δ_s may be carried out as a χ^2 test, taking $(2 - 4^2/V/4)$ (9.14)

$$\chi_{[1]}^2 = \Delta_s^2 / V(\Delta_s),$$
 (8.14)

this test being precisely equivalent to the test based on the normal distribution and the standard error of Δ_s . If tests have been carried out on several mixtures of two poisons of different proportionate constitution, each giving an estimated Δ_s , a composite test of agreement with the similar action hypothesis may be obtained by adding the χ^2 values calculated as in (8.14) and testing as a χ^2 with the total number of degrees of freedom. If allowance has had to be made for heterogeneity, the total of the χ^2 values must be divided by the degrees of freedom and the result used in a variance ratio test of significance. The various Δ_s and $V(\Delta_s)$ are, strictly speaking, not independent, since all are dependent upon the common value of b and upon information from tests on the constituent poisons used separately. Nevertheless, the composite test derived from equation (8.14) should give an approximate test of synergism for use when the more complicated procedure described in § 37 seems unnecessary.

In any experiment designed to throw light on the potency of mixtures of poisons, the constituent poisons should be tested alone as well as in mixture. Even when a considerable amount of information on their toxic effects is already available, they should not be omitted from further tests. The insects or other test subjects used in work of this nature often show great variability in response, both amongst themselves and as a result of uncontrolled experimental conditions; hence, unless it is certain that conditions have not changed appreciably in any important aspect, tests of mixtures cannot safely be compared with tests of the separate constituents made on a different occasion.

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The internal consistency of results for three or more mixtures of two poisons with the hypothesis of similar action may, however, be judged, even without tests on the poisons separately, by expressing intermediate members of the series as though they were obtained by mixing, in the appropriate proportions, the two mixtures of most extreme constitution; the illustration given in Ex. 22 will suffice to show how this is done.

Ex. 22. Expression of one mixture in terms of two others. Suppose tests have been made on three mixtures, containing respectively proportions 4: 1, 7: 3, 2: 3 of two poisons. The first and the last are the most extreme in composition, and the second may easily be expressed in terms of them. The proportions of the two constituent poisons in the mixtures are (0.8, 0.2), (0.7, 0.3)and (0.4, 0.6); the second mixture may therefore be considered as composed of proportions π_1 , π_2 of the first and third where

$$0.8\pi_1 + 0.4\pi_2 = 0.7, \quad 0.2\pi_1 + 0.6\pi_2 = 0.3,$$

whence $\pi_1 = 0.75$, $\pi_2 = 0.25$. It is easily verified that if the 4:1 and 2:3 mixtures are themselves mixed in the ratio 3:1 a mixture of the original poisons in the ratio 7:3 is obtained. The agreement of the results of the experiment with predictions made from the hypothesis of similar action may then be examined by means of the methods and formulae just discussed.

37. A GENERAL TEST FOR SIMILAR ACTION

When several mixtures with different proportionate constitutions have been tested, another method (Finney, 1942a) may be used for examining the agreement of the median effective doses with the values predicted by similar action. This method explicitly uses only the median effective doses and their standard errors; parallelism of the regressions is implied, however, since otherwise no meaning can be given to the concept of similar action. There are theoretical objections to the method on account of certain assumptions of normality and independence, but providing that the regression coefficient is estimated with reasonable precision the resulting tests of significance should be reliable. From equation (8.6), if λ_1 is the LD 50 of a poison, ρ the relative potency of a second poison, and π_1 , π_2 the proportions of the two poisons in a mixture, the LD 50 of the mixture is λ , where

$$\frac{1}{\lambda} = \left(\frac{1}{\lambda_1}\right) \pi_1 + \left(\frac{\rho}{\lambda_1}\right) \pi_2. \tag{8.15}$$

Consequently, if values of λ have been estimated for mixtures of two poisons in a series of different proportions, $1/\lambda_1$ and ρ/λ_1 may be estimated as the regression coefficients of $1/\lambda$ on π_1 and π_2 , each value of $1/\lambda$ being weighted inversely as its variance. If λ is the LD 50 corresponding to a log dose m ($m = \log_{10} \lambda$)

$$V(1/\lambda) = \frac{V(m)}{\lambda^2} (\log_e 10)^2 = \frac{5 \cdot 302 V(m)}{\lambda^2},$$

and therefore the weight to be attached to $1/\lambda$ is

$$W = 0.1886\lambda^2 / V(m).$$
 (8.16)

The regression equation (8.15) contains no 'constant term', but is constrained to give λ infinite when the content of both constituents is zero; hence total sums of squares and products must be used in calculating the regression coefficients, not sums adjusted so as to refer to deviations about means. Comparison of the values of λ calculated for each mixture from equation (8.15) with the values from which the equation was estimated shows how well the observations are fitted by the similar action hypothesis and permits a test of significance of the agreement.

Tattersfield and Martin (1935; see also Ex. 13 above) have published results of toxicity tests with ether extracts of seven different derris roots to *Aphis rumicis*, and have given the LD 50 for each in terms of rotenone and a dehydro mixture. The latter is known (Martin, private communication) to have varied considerably from root to root in its relative proportions of different dehydro compounds. Though he recognized that any comparison of toxicities based on the two constituents alone must be of doubtful value, Bliss used these data as an illustration of synergistic action between two poisons and found them to agree satisfactorily with a generalization of one of his formulae for

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synergism, provided that one root (no. 2), which behaved anomalously, was omitted. On account of the non-independence of the LD 50's and their standard errors, the analysis given is not strictly correct, but the disturbance so caused can only be slight. A more serious error of tactics is that the data were not examined for their possible agreement with the simpler hypothesis of similar action. Finney (1942*a*) has shown the median lethal doses to be satisfactorily fitted by equation (8.15), using the regression procedure outlined above. Details of the calculations will not be shown here, since the process is sufficiently illustrated by Ex. 23, in which the method is used to test similar action between three components.

Ex. 23. Similar action between the toxic constituents of derris root. Results of toxicity tests on four derris roots, carried out by Martin (1940), have been examined in Ex. 14, and comparative LD 50 values have been estimated, allowing for different levels of susceptibility of the test insects on the three days of testing; these values are given in Table 15. The variances of the median lethal doses were not discussed in Ex. 14, but for present purposes the sum of the weights in the appropriate column of Table 14 will be taken as the weight to be attached to any log LD 50; a little consideration shows this figure to be too high, though it should be of the right order of magnitude. Allowance must also be made for the heterogeneity of the relative potencies which was demonstrated by a χ^2 test at the end of Ex. 14, but this can most conveniently be done at a later stage of the analysis.

Martin subdivided the toxic constituents of each of the four roots into rotenone, a toxicarol fraction (possibly including sumatrol, malaccol, and other materials in addition to toxicarol) and a deguelin concentrate fraction (including elliptone, if present, as well as deguelin), and determined the proportions of these three for each root. The average probit regression coefficients for the three days of testing were 5.77 ± 0.39 , 5.07 ± 0.35 and 4.66 ± 0.32 . Though differences between these are not significant, there is some indication of a decrease from the beginning to the end. There is no evidence of consistent differences between roots in their log-tolerance variances. An examination of the adequacy of similar action to explain the observations is therefore of interest and provides a useful illustration of the method. If the three components act similarly, equation (8.7) should be adequate to describe the observed mortalities, and the LD 50 for any root should be given by

$$\frac{1}{\lambda} = \left(\frac{1}{\lambda_r}\right) \pi_r + \left(\frac{\rho_t}{\lambda_r}\right) \pi_t + \left(\frac{\rho_d}{\lambda_r}\right) \pi_d, \qquad (8.17)$$

 π_r , π_t , π_d being the proportions of the three components in the root, λ_r the LD 50 of rotenone, and ρ_t , ρ_d the potencies of the toxicarol and deguelin fractions relative to rotenone.

 TABLE 23. Computations for Test of Similar Action between the Constituents of Derris Root

Material	1/V(m)	λ	1	W		π	π_t	_1	π_d
Rotenone	1,640	13.2	0.0	054 × 10 ⁶	1	0000	0.00	0	0.000
W. 211	6,710	244	75.	3 × 10 ⁶	0	0146	0.15	2	0-086
W. 212	6.140	146	24.	7 × 10 ⁶	0	0414	0.04	3	0.124
W. 213	4.710	188	31.4	4 × 10 ⁶	Ŏ	0346	0.02	4	0.082
W. 214	4,820	92.0	7.0	69 × 10 ⁶		0794	0.02		0.122
Material	1/λ	W	π _r	Wπ _t		W	π _d		W/A
Rotenone	0.07576	. 54	,000	,	0		0	4	I,091·0 4
rocentino		1 1 000	200	11.445.6	00	6.475	800	308	3.730
W. 211	0.00410	1,099	,000	11,440,0					
	0.00410	1,099		1.062.1		3.062			.195
W. 211			,580		00		,800	169	

Table 23 shows the first stage in the fitting of this equation to the data for rotenone alone and for the four roots. The column 1/V(m) contains the column totals from Table 14, representing approximately the weight to be attached to each log LD 50, and the column λ contains the values of the LD 50 from Table 15; from these and equation (8·16), W has been calculated. The proportions of rotenone, toxicarol and deguelin $(\pi_r, \pi_i \text{ and } \pi_d)$ are reproduced from Martin's Table 12 (1940), $1/\lambda$ is tabulated, and the products of W with each of the four following columns are then entered in full.

Sums of products of the entries in each of the last four columns in Table 23 with π_r , π_t , π_d and $1/\lambda$ are next formed. Sums of products such as $SW\pi_r\pi_t$ are obtained in two ways, the agreement

being a check on the arithmetic; sums of squares such as $SW\pi^2$, must be recalculated as a check. Systematic arrangement of the results gives the following equations for the three regression coefficients in (8.17):

$$198,457(1/\lambda_{\tau}) + 253,026(\rho_{l}/\lambda_{\tau}) + 384,926(\rho_{d}/\lambda_{\tau}) = 28,020\cdot1,$$

$$253,026(1/\lambda_{\tau}) + 1,808,686(\rho_{l}/\lambda_{\tau}) + 1,202,210(\rho_{d}/\lambda_{\tau}) = 60,384\cdot8,$$

$$384,926(1/\lambda_{\tau}) + 1,202,210(\rho_{l}/\lambda_{\tau}) + 1,262,298(\rho_{d}/\lambda_{\tau}) = 71,426\cdot9.$$

These equations must be solved to a greater number of decimal places than is needed for the comparison of potencies, in order to have sufficient accuracy in the test of significance of the departure from similar action. The solutions are:

$$1/\lambda_r = 0.0800010, \ \rho_t/\lambda_r = 0.0021760, \ \rho_d/\lambda_r = 0.0301168.$$

Also the weighted sum of squares of $1/\lambda$ is

$$SW(1/\lambda)^2 = 4532.04,$$

and the portion of this accounted for by the linear regression on π_r , π_l , π_d is

$$(1/\lambda_r) SW\pi_r/\lambda + (\rho_l/\lambda_r) SW\pi_l/\lambda + (\rho_d/\lambda_r) SW\pi_d/\lambda = 4524.18.$$

Now $SW(1/\lambda)^2$, being a sum of squares *not* adjusted for a mean, has 5 degrees of freedom, and the residual after the fitting of three constants therefore has 2 degrees of freedom. If the weights, W, had been truly the reciprocals of the variances of $1/\lambda$, the residual would have been a χ^2 , namely,

$$\chi^2_{[2]} = 7.86.$$

Quite apart from the considerations of non-independence mentioned earlier, the weights require to be reduced on account of the heterogeneity of relative potencies found in Ex. 14

$$\chi^2_{(2)} = 14 \cdot 24.$$

Hence a test of significance for departures from the similar action law is obtained by comparing mean squares from these two χ^2 values (Fisher and Yates, 1948, Table V). Since the mean square for departures from similar action is less than the heterogeneity mean square, the data do not contradict similar action, but the test is not very precise because of the few degrees of freedom available for the heterogeneity χ^2 and mean square.

The LD 50 for rotenone, λ_r , is estimated as 12.50 mg./l., and the potencies of the toxicarol and deguelin fractions relative to rotenone, ρ_i and ρ_d , are estimated as 0.02720 and 0.37645 or $\frac{1}{37}$ and a little over $\frac{1}{3}$, as compared with the values of $\frac{1}{15}$ and $\frac{1}{5}$ used by Martin in computing rotenone equivalents for the four roots. The rotenone equivalent is $(\pi_r + \rho_i \pi_i + \rho_d \pi_d)$, and the estimated LD 50 for any root is obtained by dividing the LD 50 for

TABLE 24. Comparison of Median Lethal Doses of Derris Roots and Values Predicted according to Similar Action (doses in mg./l.)

Sample	LD 50 in	Predicted all da		Predicted, omitting rotenone		
Gampie	Table 15	% rotenone equivalent	LD 50	% rotenone equivalent	LD 50	
Rotenone W. 211	13.2	100.000	12.5	100.000	(9-09)	
W. 211 W. 212	244 146	5·111 8·925	245 140	3·728 6·240	244 146	
W. 213 W. 214	188 . 92·0	6·612 12·603	189 99-2	4·819 9·898	189 91·9	

rotenone by the rotenone equivalent. Table 24 shows the results and makes it clear that the expression of the toxic contents of the four roots in terms of their rotenone equivalents gives a very reasonable evaluation of their potencies.

The rotenone content of the roots is so small, and in consequence the potencies of the roots are so very different from that of rotenone, that it may seem more appropriate to examine the consistency with the similar action law of the data for the four roots alone, ignoring the rotenone tests entirely. The only changes which the omission of the first line in Table 23 produces in the equations for the three parameters are that $SW\pi_r^3$ becomes 144,457 instead of 198,457 and $SW\pi_r/\lambda$ becomes 23,929·1 instead of 28,020·1. The solutions now lead to

 $\lambda_r = 9.093, \quad \rho_l = 0.06642, \quad \rho_d = 0.14634.$

That the four roots show excellent agreement with the similar action hypothesis is evident from the rotenone equivalents and estimated LD 50's, also shown in Table 24, though naturally the discrepancy for rotenone itself becomes much greater than in the earlier analysis.

From his examination of the same data, Martin concluded that there was some evidence of synergistic action between the three toxic constituents of the roots. The present more complete analysis shows that the indications of synergism disappear when the best possible values for the relative potencies of the constituents are estimated from the data themselves instead of from an earlier investigation. Nevertheless, synergism may be present in a form that leaves the data internally consistent with similarity; the matter can scarcely be settled satisfactorily without comprehensive trials including tests both on the roots and on the constituents.

38. INDEPENDENT ACTION

The distinction between similar and independent action must be kept clear. In mixtures whose constituents act similarly any quantity of one constituent can be replaced by a proportionate amount of any other without disturbing the potency, but for mixtures whose constituents act independently the mortalities, not the doses, are additive. This type of action may occur with a mixture whose constituents produce their toxic effect in entirely different ways, as, for example, a mixture of two insecticides of which one is a stomach poison and the other a contact poison.

[•]Suppose that the doses of two poisons given in a mixture are capable of producing mortalities P_1 , P_2 when used separately. If the two act independently, a proportion P_2 of the test subjects which would survive the first poison is expected to succumb to the second, thus giving an expected total mortality (cf. § 26)

$$P = P_1 + P_2(1 - P_1).$$

This may be written

 $P = 1 - (1 - P_1)(1 - P_2) \tag{8.18}$

in order to display the symmetry in P_1 and P_2 . Equation (8.18) is the basic expression of independent action between two

poisons; for three constituents, the corresponding relationship is

$$P = 1 - (1 - P_1) (1 - P_2) (1 - P_3), \qquad (8.19)$$

whence the method of extension to a greater number of constituents is apparent (see $\S 53$).

Bliss (1939*a*) has suggested a modification of equation (8.18) to allow for a correlation in the susceptibility to the two components. If $P_1 > P_2$, so that the first component is the more toxic, his equation is $P_1 = P_1 + P(1 - P_2)(1 - r_1)$ (8.20)

$$P = P_1 + P_2(1 - P_1)(1 - r), \qquad (8.20)$$

r representing the degree of correlation between the susceptibilities to the two poisons.* When r = 0, this last equation reduces to (8.18); when r = 1 it becomes simply

 $P = P_1$,

so that the combined mortality is the same as that for the more toxic ingredient applied alone. Negative correlation of susceptibilities does not fall within the scope of equation (8.20), since negative values of r would make P > 1 for certain values of P_1 and P_{e} . As will be seen from Ex. 26, a critical experiment to distinguish between equations (8.18) and (8.20), even for values of r close to 1, would usually require many more test subjects than are normally available; there appears to be no evidence, either theoretical or experimental, that equation (8.20) represents any real biological situation, and therefore no detailed study of it will be attempted here. Indeed, so far as is known to the writer, no clear experimental demonstration of the occurrence of the simplest form of independent action has yet been made. and equation (8.18) may well be too crude an approximation to any interaction of effects of mixed poisons to be of much practical value. A discussion of the types of relationship to which the equation can lead is of interest, however, as giving some indication of what may be expected in a future approach to more complex problems.

Simple though the concept of independent action is, the statistical treatment of data relating to poisons acting in this

^{*} The symbol r is used here with a meaning entirely different from that in other sections of this book.

manner is much more difficult than that of data relating to similar action. Even though the two constituents of a mixture give mortality probits which are linearly related to the log dose and the two probit regression lines are parallel, the regression relationship for the mixture will not be a straight line; still less can this be so when the regression lines for the constituents are not parallel. No exact methods of statistical analysis are available, but a few illustrations of curves obtained according to the independent action law, for mixtures whose constituents have the usual normal distribution of log tolerances, will show the types of probit-dosage relationship which may be encountered.

Ex. 24. Independent action between constituents with parallel probit regression lines. Consider two poisons having the probit regression lines

$$Y_1 = 6 + 2x, \quad Y_2 = 4 + 2x;$$

the first has ten times the potency of the second at all levels of mortality. In a 1:1 mixture of the two, the concentration of the first poison in a total concentration λ will be $\frac{1}{2}\lambda$, and the log concentration of the first will therefore be $(x - \log 2)$. For low values of x, the kill produced by the second poison will be negligible relative to that produced by the first, and the total effect will be almost identical with that caused by the content of the first poison alone; hence for low values of x the mixture gives mortality probits determined by

$$Y = 6 + 2(x - \log 2) = 5 \cdot 40 + 2x.$$

With increasing total concentration, the amount of the second poison present in the mixture begins to produce an appreciable effect, so that the total kill is greater than that for the first alone. The curve relating the probit of the total mortality to the dosage is shown in Fig. 12.*

If the standard deviation of the log-tolerance distributions for the two poisons is altered, without change of the mean probit

* Points on this curve are most easily calculated by first tabulating Y_1 and Y_2 for a series of values of x; for example, when x = 0.20 for the mixture, the log concentration of each constituent present in the mixture is x = -0.10, so that $Y_1 = 5.8$, $Y_2 = 3.8$. Values of Q_1 , Q_2 .

difference, the only change in Fig. 12 is a change in the scale of x. For example, for two poisons giving the separate lines

$$Y_1 = 6 + 10x, \quad Y_2 = 4 + 10x,$$

a 1:1 mixture shows the same response curve as Fig. 12, except that one unit of x in that figure must now be read as only $\frac{1}{2}$ unit.

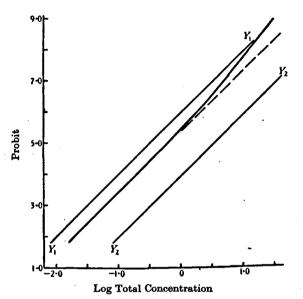


FIG. 12. Independent action in a 1:1 mixture of poisons with probit regression lines $Y_1=6+2x$, $Y_2=4+2x$. Curve shows dosage-response relationship for mixture (Ex. 24). Broken line shows dosage-response relationship for correlated independent action with r=1 (Ex. 26).

If the standard deviation remains unaltered, but the mean log tolerances are changed, the curve is again unaltered as long as the relative potency is the same. For example, if

$$Y_1 = 3 + 2x, \quad Y_2 = 1 + 2x,$$

$$Q = Q_1 Q_2,$$

the proportions of test subjects separately surviving these dosages, are then obtained from Table 1; here $Q_1 = 0.2119$ and $Q_2 = 0.8849$. Now equation (8.18) can alternatively be written

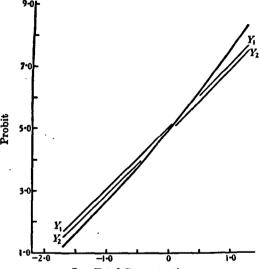
so that tabulation of products gives the proportion surviving the mixed dose, or in this instance Q = 0.1875; the value of Y = 5.89 is then read directly from Table 1.

the two lines and the curve are shifted horizontally by 1.5 units; the change may be accomplished by writing (x+1.5) for x in Fig. 12.

If the relative potency of the two poisons is changed, the curve for the mixture is changed though still very similar in general form. For the lines

$$Y_1 = 5 \cdot 1 + 2x, \quad Y_2 = 4 \cdot 9 + 2x,$$

a 1:1 mixture gives the curve in Fig. 13.



Log Total Concentration

FIG. 13. Independent action in a 1:1 mixture of poisons with probit regression lines $Y_1 = 5 \cdot 1 + 2x$, $Y_2 = 4 \cdot 9 + 2x$. Curve shows dosage-response relationship for mixture (Ex. 24).

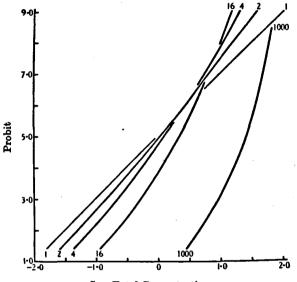
The contrast between similar action and independent action is emphasized by consideration of the interesting limiting case of a mixture of two poisons whose separate probit lines are identical. Under similar action, any such mixture, irrespective of the proportions, would give the same line. Under independent action a curve is obtained, of a form very like those just discussed, which at low concentrations lies below the lines for the two constituents, but at high concentrations lies above. If

$$Y_1 = Y_2 = 5 + 2x$$
,

the curve for a 1:1 mixture is obtained by calculating, for various values of x, $Q = Q^2$.

$$Q = Q_1^2;$$

this curve is shown in Fig. 14, together with further curves for mixtures of 4, 16 and 1000 components in equal proportions, all having the same individual probit lines.



Log Total Concentration

Fig. 14. Independent action in mixtures of several poisons each with probit regression line Y=5+2x. Curves are drawn for 2, 4, 16 and 1000 components (Ex. 24).

From consideration of Figs. 12-14 the general nature of the curve for a mixture in equal proportions, at least when the two constituents give parallel probit lines, may be inferred. At low concentrations the curve is indistinguishable from the line representing the effect of the mixture's content of the more potent constituent applied alone; if the relative potency of the more potent constituent is less than two (Fig. 13) the kill at these concentrations is even less than for the same total concentration of the less potent constituent alone. At higher concentrations, the kill is augmented by an appreciable effect of the weaker

constituent, so that the curve turns upward and is no longer parallel to the two original lines. If the two constituents do not differ greatly in potency (Figs. 13, 14), as the concentration is increased the mixture soon becomes more effective than the same total concentration of the stronger constituent, but if the difference is large (Fig. 12) this does not occur until very high concentrations and kills are reached.

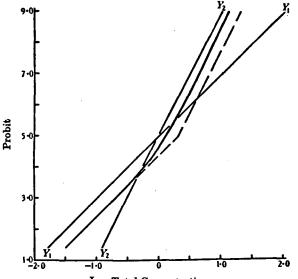
The discussion just given may be extended to cover mixtures in proportions other than 1:1 by a simple transformation. A 2:3 mixture, for example, may be considered as a mixture in equal proportion of the first constituent with a new second constituent of 3/2 the potency of the original. Hence on the probit diagram the line for the second constituent may be shifted a distance $b \log (3/2)$ to the left and the behaviour of the mixture then deduced as that appropriate to a 1:1 mixture. For numerical calculation nothing is gained by this process, as the ordinates of the curve for the mixture may just as easily be calculated directly.

Bliss (1939a) implied that mixtures of two poisons showing similar action always had greater potency than mixtures of two poisons with potencies equal to those of the first pair but showing independent action. This is true at moderate dose rates if the relative potency of the constituents is large, but Figs. 13 and 14 indicate that if the relative potency approaches unity independence gives a lower potency at low doses and a higher potency at high doses than does similarity.

Ex. 25. Independent action between constituents with intersecting probit lines. The curve relating mortality probit to dosage for a 1:1 mixture of independently acting poisons whose separate regression equations are

$$Y_1 = 5 + 2x, \quad Y_2 = 5 + 4x,$$

is shown in Fig. 15; in general characteristics it is typical of the curve resulting from mixing poisons whose regression lines intersect. At concentrations sufficiently low for the constituent with the greater log-tolerance variance (i.e. lesser value of b) to be the more potent, a 1:1 mixture produces almost the same effect as would its content of this constituent applied alone. At higher concentrations, the curve for the mixture eventually rises rather more steeply than the line for the constituent with the greater value of b; as the curve increases in steepness with increasing concentration it may be presumed eventually to intersect the line for this constituent alone, but unless the original lines are nearly parallel this intersection seems not to occur until very high kills have been reached.



Log Total Concentration

F1G. 15. Independent action in a 1:1 mixture of poisons with probit regression lines $Y_1 = 5 + 2x$, $Y_2 = 5 + 4x$. Curve shows dosage-response relationship for mixture (Ex. 25). Broken line shows dosage-response relationship for correlated independent action with r = 1 (Ex. 26).

Curves calculated for 1:1 mixtures whose constituents have the regression lines

$$Y_1 = 4 + 2x, \quad Y_2 = 4 + 4x, \\ Y_1 = 2 + 2x, \quad Y_2 = 2 + 4x,$$

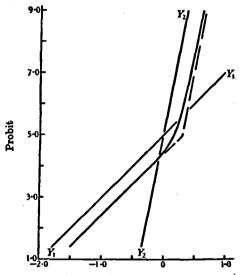
or

have been found to be very like those of Fig. 15, except for the displacement caused by the changed point of intersection of the lines, but they are not quite identical in shape. Two poisons whose probit lines are

$$Y_1 = 5 + 2x, \quad Y_2 = 5 + 10x,$$

differ more markedly in tolerance variance than those just considered, but the curve for a 1:1 mixture has the same general appearance (Fig. 16). Mixtures in proportions other than 1:1may be discussed as 1:1 mixtures by means of the transformation suggested at the end of Ex. 24.

Ex. 26. Correlated independent action. The curve representing the toxic effect of a mixture of two poisons showing correlated independent action, as defined by equation (8.20), is always



Log Total Concentration

F16. 16. Independent action in a 1:1 mixture of poisons with probit regression lines $Y_1=5+2x$, $Y_2=5+10x$. Curve shows dosage-response relationship for mixture (Ex. 25). Broken line shows dosage-response relationship for correlated independent action with r=1 (Ex. 26).

intermediate in position between that for completely independent action and the line or sections of lines for the more potent constituent alone. Thus for complete correlation (r = 1) in Ex. 24, since one constituent is always more potent than the other at all dosages, the mixture would follow the line for the amount of this constituent present in the mixture; this line is shown in Fig. 12 and is a continuation of the initial rectilinear portion of the curve for r = 0. For less complete correlation, represented by 0 < r < 1, an intermediate curve would be obtained. In Ex. 25 the more potent constituent is not the same at all dosages. Sections of lines corresponding to r = 1 are also shown in Figs. 15 and 16; again lesser values of r would give curves in intermediate positions. The two extremes (r = 0 and r = 1) are often not very different, and only very extensive and precise data could permit satisfactory discrimination between different values of r. With experimental results for poisons whose effects are like those shown in Fig. 16, for example, it would be very difficult to assess r with any precision. Indeed, in practice, data which are appreciably less satisfactorily fitted by r = 0 than by some other value of rwill rarely be encountered.

As stated above, no exact statistical treatment of data relating to independent action has yet been developed. If probit lines for the constituents are estimated in the same series of tests as those on the mixture, these lines may be used to predict the form of the curve for the mixture, and the observed mortalities may then be compared with the expected, by a χ^2 test in the manner of that used in Ex. 1. Such a test ignores the errors of estimations of the lines for the constituents, and is therefore liable to exaggerate the significance of discrepancies, but no alternative can at present be suggested. No examples from experimental data will be given here, as none suitable have been found in the published literature.

Mortality probits corresponding to low concentrations of a poison have often been found to be higher than predicted by the line fitted to the whole data. Bliss (1939*a*) has suggested that this phenomenon of a 'break' in the line may indicate that the poison is in fact a mixture of two or more toxic components; the possibility merits further consideration, but detailed experimentation at many concentrations is needed if it is to be examined adequately. Such breaks are often drawn as sudden changes in slope, a situation which can only occur with independent action if r = 1. The data are seldom sufficiently precise, however, for any certainty that the change in slope is not more gradual and of the type found for lesser values of r, perhaps even for r = 0. Frequently only the upper portion of the curve is of interest,

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since this usually extends over the important range of kills, and difficulties of analysis may be avoided by ignoring the results for the lower dosages, a policy which is theoretically objectionable but practically justifiable.

Murray (1938) published the results of tests on the toxicity of a pyrethrin spray to the house-fly which give some evidence of independent action of pyrethrins I and II; a distinct 'break' occurs for the female flies, but the more susceptible males show only one phase of action over the whole range of concentrations tested. The proportions of the two pyrethrins present in the spray were not stated, nor were tests made on either component alone, so that no critical judgement can be formed. The data have been discussed by Bliss.

39. SYNEBGISTIC ACTION

Many writers have used the terms synergism and antagonism to describe the joint action of certain mixtures of poisons without giving any unambiguous definition of their meaning. The first attempt to formulate algebraic relationships which would represent this type of action was due to Bliss (1939*a*); he proposed two alternative equations for the simple case of similar synergistic action, in which the probit-dosage regression lines for mixtures and for the constituent poisons are all parallel.

Bliss endeavoured to find the relationship between the quantities of two poisons present in equipotent doses of mixtures behaving synergistically. If a total dose λ contains quantities λ_1 and λ_2 of two constituents, the first being the 'more active ingredient', an equation will connect the values of λ_1 and λ_2 for which the total kill is constant.* The first equation suggested by Bliss may be written in the form-

$$\lambda_1^{\epsilon}(\lambda_1 + \lambda_2) = k, \qquad (8.21)$$

where ϵ depends only upon the two poisons and k is a function only of the level of kill selected. Bliss does not state by what criterion it is to be decided which ingredient is the more active, though some standard independent of dose seems to be implied.

* For similar action, the equation is $\lambda_1 + \rho \lambda_2 = \text{constant}$.

The equation cannot hold for very small doses of the first poison, since it implies that the addition of a trace of this to a moderate dose of the second would reduce rather than increase the potency.

The second equation, which Bliss considered more satisfactory, is $\lambda_{s}^{\varepsilon}(1+h\lambda_{1}) = k$, (8.22)

where ϵ again depends only on the two poisons and h, k are functions of the level of kill selected for comparing the data. On the analogy of Clark's findings for drug antagonism (1937), Bliss suggests that ϵ is frequently very close to unity; he also states that in one example the product hk appeared to be almost independent of the kill and proposes that its value might therefore be used as a measure of the intensity of synergism. Equation (8.22) must break down at very small doses of the second poison since it implies that a preparation containing only a trace of the second mixed with the first would be less toxic than the same amount of the first applied alone.

Both these equations must be judged unsatisfactory representations of similar synergistic action. Apart from the discontinuity for small doses and the difficulty of defining the more active ingredient, there is the disadvantage that neither includes similarity or independence, the two simpler forms of joint action already discussed, as cases of zero synergism. Though the final appeal must be to experiment rather than to abstract argument, equations (8.21) and (8.22) appear unlikely to be very helpful in describing synergistic action, particularly as each requires at least two parameters, one of which depends upon the level of mortality.

An alternative equation, which may satisfactorily fit some data whose probit regression equations, both for mixtures and for their constituents, are rectilinear and parallel, has been suggested by Finney (1942*a*). It may be written, using the same notation as in § 36,

$$Y = a + b \log (\pi_1 + \rho \pi_2 + \kappa \sqrt{\rho \pi_1 \pi_2}) + bx, \qquad (8.23)$$

and is an extension of equation (8.9). A dose λ of the mixture produces the same effect as a dose $(\pi_1 + \rho \pi_2 + \kappa \sqrt{\rho \pi_1 \pi_2})\lambda$ of the first constituent alone. If $\kappa = 0$, equation (8.23) represents similar

10-2

action. If κ is positive, the potency is greater than that predicted by similar action, and the poisons act synergistically; if κ is negative, they act antagonistically. Equation (8.23) may prove at least a satisfactory empirical formula for mixtures whose probit lines are parallel to those for their constituents; the constant κ will be called the *coefficient of synergism*. The equation will easily generalize so as to include mixtures of several toxic constituents.

In order that the agreement between experiment and a proposed law of synergistic action may be examined, toxicity tests must be carried out with a range of doses of mixtures in at least two different proportions as well as on the constituents separately. From data collected in such an experiment the coefficient of synergism, as defined above, can be estimated by a generalization of the method described in § 37, using now a regression equation in the form

$$\frac{1}{\lambda} = \left(\frac{1}{\lambda_1}\right) \pi_1 + \left(\frac{\rho}{\lambda_1}\right) \pi_2 + \left(\frac{\kappa \rho^{\mathbf{i}}}{\lambda_1}\right) (\pi_1 \pi_2)^{\mathbf{i}}.$$
 (8.24)

The only data found in published papers which give definite evidence of synergism and which are shown in a form suitable for testing the adequacy of equation (8.23) are discussed in Ex. 27. Before the many problems of synergism can be elucidated, a great amount of further experimentation, testing mixtures in several different proportions, must be carefully planned and executed. Without much more experimental evidence than is at present available any complete discussion even of similar synergistic action is impossible; no attempt will be made here to unravel the complexities that may arise when the separate regression lines are not parallel.

Ex.27. The toxicity of rotenone-pyrethrins mixtures to the housefly. Le Pelley and Sullivan (1936) have reported the results of two series of trials in which adult house-flies were sprayed with alcoholic solutions of rotenone, pyrethrins, and a mixture of the two; in the first series the mixture contained rotenone and pyrethrins in the proportion of 1:5 (by weight), and in the second series the proportion was 1:15. About 1000 flies were tested at five levels of each toxic preparation, and the kills obtained were as shown in Table 25, these figures having been abstracted from the diagrams in the original paper. The authors interpreted the results as indicating no striking antagonistic or synergistic effect in the mixtures. A footnote by H. H. Richardson asserted that in the first series there was pronounced synergism and in the second an effect in the same direction but of less

	First series		Second series				
Concentra- tion (mg./c.c.)	No. of flies	% kill	Concentra- tion (mg./c.c.)	No. of flies	% kill		
	Rotenone		Rotenone				
0.10	1000	24	0.10	000	28		
0.15	1000	44	0.15	900	51		
0.20	1000	63	0.20	900	72		
0.25	1000	81	0.25	900	82		
0.32	1000	90	0.35	900	89		
	Pyrethrins			Pyrethrins			
0.20	1000	20	0.50	1 900 í	23		
0.75	1000	35	0.75	900	44		
1.00	1000	53	1.00	900	55		
1.50	1000	80	1.20	900	72		
2 ·00	1000	88	2.00	900	90		
M	lixture (1 : 5)		М	ixture (1 : 15)			
0.30	1000	27	0.40	900	23		
0.42	1000	53	0.60	900	48		
0.60	1000	64	0.80	900	61		
0.875	1000	82	1.20	900	76		
1.175	1000	93	1.60	900	93		

TABLE 25. Toxicity of Rotenone and Pyrethrins to House-flies

apparent significance; Richardson here used a prediction for the mixture equivalent to the similar action law. Bliss (1939a) confirmed Richardson's conclusion, and Finney (1942b), after a new analysis of the data, also agreed that there was evidence of synergism.

In this last analysis, for each series, parallel probit lines were fitted to the data for the two poisons and their mixture. For the first series, the LD 50's were 0.156, 0.918 and 0.455 mg./c.c. for rotenone, pyrethrins and the 1:5 mixture respectively. The pyrethrins were slightly more than one-sixth as toxic as the rotenone (more precisely, $\rho = 0.170$), whence the similar action

law predicts a relative potency of 0-308 for the mixture, or an LD 50 of 0-506 mg./c.c. The mixture was thus 11 % more potent than would be expected if its constituents acted similarly; a test of the significance of this synergistic effect may be obtained from

$$\Delta_{s} = 0.168 \pm 0.050,$$

leaving little doubt that the enhanced toxicity is greater than can be attributed to random sampling variation. Analysis of the second series gave 0-142, 0-889 and 0-651 mg./c.c. as the LD50's for rotenone, pyrethrins and the 1:15 mixture. The values for the two constituents of the mixture were thus very close to those obtained in the first series, and gave $\rho = 0.160$. Similar action then predicts a relative potency of 0-212 for the mixture, and therefore an LD50 of 0-670 mg./c.c. This mixture was 3% more potent than predicted, but the difference here is not significant since

$$\Delta_{e} = 0.039 \pm 0.067.$$

Both series of tests give some indication of synergism, though in only one is the departure from similarity significant. It is therefore of interest to inquire whether the equation for similar synergistic action (equation (8-23)) will fit the results. The data are insufficient for any precise estimation of the coefficient of synergism, but a value of about 0-15 may easily be seen to be satisfactory. Using $\kappa = 0.15$, the expression $(\pi_1 + \rho \pi_2 + \kappa \sqrt{[\rho \pi_1 \pi_2]})$ gives the expected potency of a mixture relative to that of rotenone; for the two mixtures under test the values are 0-331 and 0-227 respectively. The corresponding LD 50's, 0-471 and 0-626 mg./c.c., agree well with the experimental determinations of 0-455 and 0-651 mg./c.c.; even without exact statistical tests the discrepancies are seen to be not significant, so that at least the data do not contradict the hypothesis expressed by equation (8-23).

40. PLANNED TESTS OF SYNERGISM AND SIMILABITY When experiments on the joint action of two poisons have to be planned in the absence of any information about the existence of synergism or antagonism between them, a working hypothesis that the action will be similar may reasonably be adopted. Unless there are *a priori* considerations governing the proportionate constitutions of mixtures to be tested, it is preferable that the probit lines for mixtures should be fairly evenly spaced between those for the constituents used separately. If the second constituent is ρ times as potent as the first, its probit line will be a distance $\log \rho$ to the left of the first. On the hypothesis of similar action, if a mixture in the proportion $\pi:(1-\pi)$ yields a probit regression line at a distance $\theta \log \rho$ to the left of that for the first poison, where θ is some fraction between 0 and 1, then

$$\log\left\{\pi+\rho(1-\pi)\right\}=\theta\log\rho,$$

whence

$$\pi = \frac{\rho - \rho^{\theta}}{\rho - 1}.$$
 (8.25)

In Table 26 π is tabulated, as a percentage, for a series of values of ρ and θ ; providing that an approximation to ρ is available from earlier experiments, the table may conveniently be used in planning toxicity tests intended for the investigation of similar and similar synergistic action.

Ex. 28. The use of Table 26 in planning toxicity tests. Suppose that toxicity tests are to be planned for two poisons and a mixture whose constitution shall be chosen so that, if similar action is operating, the probit regression line for the mixture will be midway between the lines for the constituents, all dosages being measured as the logarithms of total poison content. Suppose further, that, from previous experience, the second poison is believed to be about four times as toxic as the first. Entering Table 26 with $\rho = 4$ in the column for $\theta = 0.5$, the required proportion of the first poison is found to be 67 %, and therefore a 2:1 mixture of the two poisons should be used.

Again, suppose that three different mixtures of two similarly acting poisons are required such that the mixtures give probit lines equally spaced between those for the separate poisons, and assume that the second constituent is known to be about twelve times as potent as the first. Interpolation in Table 26 for $\rho = 12$ and $\theta = 0.25$, 0.5 and 0.75 gives figures of about 91, 77 and 50 %

for the amounts of the first poison in the mixtures. Suitable mixtures would therefore be made in the proportions of 10:1, 7:2 and 1:1.

In order to obtain reliable evidence of the nature of the joint action of the two constituents of a mixture, a minimum of four concentrations of each toxic preparation should be tested. If

ρ	0.1	0.5	0.3	0-4	0.5	0.6	0.7	0.8	0.9
1.1	90.4	81	71	61	51	41	31	21	10
1.2	91.7	83	74	65	55	.45	34	23	12
2	92.8	85	77	68	59	48	38	26	13
3 4	94.2	88	80	72	63	53	42	30	16
4	95-0	89	83	75.	67	57	45	32	17
5	95.6	90-5	84	77	69	59	48	34	19
6	96-1	. 91-4	86	79	71	61	50	36	20
7 8 9	96.4	92-1	87	80	73	63	52	38	21
8	96.7	92.6	88	81	74	65	53	39	21
9	96.9	93-1	88	82	75	66	54	40	22
10	97.1	93.5	89	83	76	67	55	41	23
15	97.8	94.9	91.0	86	79	71	60	45	25
20	98.2	95.7	92· 3	88	82	74	62	47	27
25	98-4	96-2	93·2	89	83	75	65	49	29
30	98 ∙6	96 ∙6	93·9	90	85	77	66	51	30
40	98·9	97-2	94.8	91.3	-86	79	69	54	32
50	99.0	97.6	95·4	92.3	88	81	70	55	33
60	99·1	97.9	95·9	93.0	89	82	72	57	34
80	99.3	98·2	96 ∙6	94·0	90	84	74	59	36
100	99·4	98 .5	97·0	94.6	90.9	85	76	61	37

TABLE 26.	The Function $\pi = 100(\rho - \rho^{\theta})/(\rho - 1)$ used
in Planı	ing Tests of Mixtures of Two Poisons

there is no prior knowledge of synergism, the test doses of each should roughly be inversely proportional to their relative potencies. For example, consider the trials on two poisons of relative potency twelve and their three mixtures in the proportions determined in the last paragraph. The relative potencies of the five preparations are, by equation (8.5), approximately 2:4:7:13:24, and doses inversely proportional to these numbers should be used. Hence if the LD50 for the stronger poison were thought to be about 0.2 unit and experience had shown the suitability of a two-fold increase in dose between

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successive levels, the following sets of five doses of each preparation might be chosen:

		Mixture in proportions of								
	1:0	10:1	7:2	1:1	0:1					
	(0.004	0.008	0.012	0.025	0.02					
	0.008	0.016	0.03	0.02	0.1					
Doses -	0.016	0.032	0.06	0.1	0.2					
	0.032	0.064	0.12	0.2	0·4					
	0.064	0.13	0.24	0.4	0.8					

41. COMPOUND RESPONSE CURVES

An unusual dosage-response curve has been reported (Dimond et al. 1941) for tests of the toxic effect of tetramethylthiuram disulphide to spores of Macrosporium sarcinaeforme. Dr Dimond has kindly made available more information about these tests than was originally published. Five concentrations, ranging from 0.2% to 0.00002%, were sprayed on to glass slides, and several different spray times between 50 and 5 sec. were tested for each concentration. In this way, deposits ranging from 35 to $0.00105 \ \mu g./sq.cm.$ were obtained, some of these being duplicated by being made up from two different combinations of concentration and time. Spore suspensions were then added to the dried residues of the spray so that the concentration of the toxicant in the drop of suspension was proportional to the density of the dried deposit on the slide. The percentage inhibition of spore germination was measured; Fig. 17, which has been copied from that published by Dimond et al., shows the relationship between the inhibition probits and log deposits. The very close agreement between percentage inhibitions from pairs of results with equal deposits, but different combinations of duration of spraying and concentration, suggests that the total deposit was the chief factor determining the response and that the method of obtaining this deposit was comparatively unimportant. The curve indicates that tetramethylthiuram disulphide had a maximum potency at about 0.06 μ g./sq.cm., above which level any increase in dose decreased the inhibition of germination until a minimum was

reached at about $0.3 \mu g./sq.cm$. At still higher doses, the potency again increased and rose beyond the previous maximum; inhibition was complete for the six highest levels tested.

The authors make the following comments:

'A possible explanation of this type of behaviour is that dissociation or association of the toxicant occurs. It seems likely that weakly-dissociating materials may dissociate (or associate),

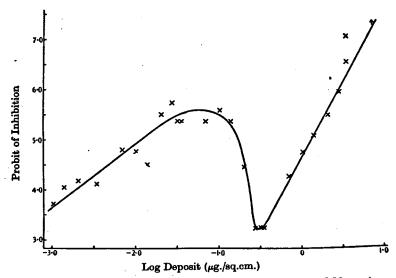


FIG. 17. Toxicity of tetramethylthiuram disulphide to spores of M. sarcinaeforme. Each point is based on a count of 100 spores. Complete inhibition was recorded for log deposits of 0.85, 1.02, 1.15, 1.32, 1.45 and 1.54.

forming a complex which has toxicity markedly different from the original molecule. In the case of tetramethylthiuram disulfide, the toxicity of the dissociation complex would be greater than that of the undissociated molecule.

'In the first phase of toxic action," where decrease in toxicity is proportional to dilution on the logarithmic-probability scale, the proportion of dissociated molecules as compared with the dissolved, undissociated molecules of toxicant might be very

* The authors explain that the argument in this paragraph has been put in terms of decreasing concentration instead of increasing, in order to simplify the discussion of the effect of molecular dissociation. small, and toxic action would be determined largely by the undissociated molecules. As dilution continues, however, the proportion of dissociated to undissociated molecules increases, and, if the dissociated molecule is very much more toxic than the undissociated molecule, the inhibition of spores will rise with further dilution. Finally, when the original toxicant is completely dissolved and dilution has progressed to the stage at which there are no undissociated molecules left, the second peak of toxicity has been reached. Dilution beyond this point can only cause a decrease in concentration of dissociated molecules and in toxicity.'

Thus they suggest that tetramethylthiuram disulphide has two separate types of toxic action, that of the undissociated molecule occurring when high concentrations are used, and that of the dissociated molecule occurring when low concentrations are used. At intermediate concentrations presumably the toxicant must behave as a mixture of the two toxic materials; the proportionate constitution of the mixture will depend upon the total concentration of the poison, and as the concentration decreases the proportion of the dissociated molecule will change smoothly from 0 to 1. Montgomery and Shaw (1943) have confirmed the occurrence of this form of response curves for several thiuram sulphides with spores of *Venturia inaequalis*.

Any study of the form of dosage-response curve that may be expected in these circumstances requires knowledge not only of the potency of a mixture of the two toxic constituents in any given proportions but also of the law determining the proportions that occur at any concentration. The two extreme linear sections of the curve in Fig. 17 are clearly not parallel, and therefore similar action cannot be operative. But, even with parallel lines to represent the extreme phases of toxic action, an increase in toxicity with decrease in concentration at intermediate levels could not occur as a result of similar action unless the total concentration of dissociated molecules, not merely the proportion of dissociated to undissociated, were then increasing; the dissociation would therefore have to be of a different type from that normally encountered, since this could not happen according

to accepted dissociation laws. On the other hand, if there were a pronounced antagonism between the toxic action of the dissociated and undissociated molecules, a curve similar to Fig. 17 might arise. Thus a possible explanation of Dimond's results is that the dissociated and undissociated molecules differ widely in their potency and in the tolerance variance shown by the spores (as evidenced by the difference in slope of the two linear sections), and that some type of antagonism is displayed when the two are in mixture. Further understanding of this complex problem must await the accumulation of a greater amount of experimental evidence.

As an attempt to set up a mathematical model of what might occur when two toxic materials which show similar action in mixture are mixed, under conditions that make the proportionate constitution of the mixture dependent upon the total concentration, a study has been made of the behaviour of a mixture in proportions $\pi_1 = 1 - \pi_2 = 10^{-k\lambda}.$

Here
$$\lambda$$
 is the total concentration of poison, k a positive constant;
the law is not intended as even an approximation to any physico-
chemical relationship, but has simply been chosen so that the
proportion of the first constituent, π_1 , is zero at high concentra-
tions and changes continuously to 1 at low concentrations. If

$$Y = a + b \log \lambda$$

gives the probit of the kill for this constituent, and the relative potency of the second is ρ , equation (8.4) gives the probit of the kill for any concentration as

$$Y = a + b \log\{\rho + (1 - \rho) \, 10^{-k\lambda}\} + b \log \lambda. \tag{8.26}$$

If ρ is less than $1/(e^2 + 1)$, or about 0.119, equation (8.26) has both a maximum and a minimum, and represents a curve of the same general form as that in Fig. 17, except that the two extreme sections are now parallel.

Ex. 29. Response curves given by equation (8.26). Fig. 18 shows the curve derived from equation (8.26) when the probit line for the first constituent is

$$Y=5+2\log\lambda,$$

the relative potency of the second constituent is $\frac{1}{100}$, and $k = \frac{1}{20}$. These values have been chosen as roughly corresponding to the curve obtained by Dimond *et al.*; the similarity with Fig. 17 is noticeable, especially in respect of the wide peak and narrow trough, though the trough in Fig. 17 is deeper. A modification of equation (8.26) so as to base it on similar synergistic action would alter the proportions of the curve in accordance with the

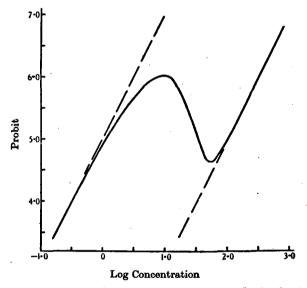


FIG. 18. Response curve from equation (8.26) for $\rho = \frac{1}{100}$, $k = \frac{1}{10}$ (Ex. 29).

amount of synergism or antagonism, but unless the departur from similarity were very great the main features would remain

When ρ is greater than 0.119 but less than 1, the maximum an minimum disappear, and increasing concentration always in creases the kill. Fig. 19 has been drawn for $\rho = 10^{-\frac{1}{2}}$, $k = \frac{1}{2^{\frac{1}{2}}}$ the probit line for the first constituent being the same as before. Typical of the situation in which the second constituent is the more toxic is Fig. 20; here $\rho = 100$, $k = \frac{1}{20}$, and the equation for the first constituent is again the same. This curve has been drawn for probits far beyond the range of practical importanc in order to show its eventual agreement with the upper line.

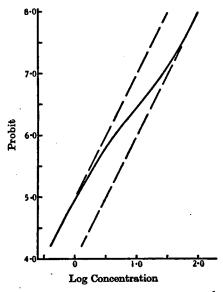


FIG. 19. Response curve from equation (8.26) for $\rho = 10^{-1}$, $k = \frac{1}{10}$ (Ex. 29).

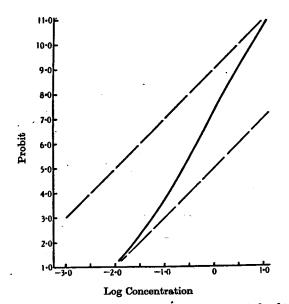


FIG. 20. Response curve from equation (8.26) for $\rho = 100$, $k = \frac{1}{30}$ (Ex. 29).

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Further discussion of the possible effects on the dose-response curve of molecular dissociation of poisons or of other types of compound response curves would be unprofitable at this stage. The only conclusion to be drawn at present is that the data of Dimond *et al.* are in general agreement with the behaviour that might occur if undissociated and dissociated molecules have very different toxicities and behave antagonistically. The account that has been given in this section is intended only as a suggestion of a mode of action that may sometimes be found, and is in no way an exact treatment of this complex problem; more detailed consideration is impossible without a much greater amount of experimental evidence as well as a fuller understanding of the underlying theory.

Chapter 9

MISCELLANEOUS PROBLEMS

42. VABIATION BETWEEN BATCHES

In the analyses discussed in previous chapters it has been tacitly assumed that all the subjects at one dose were members of a single batch and were tested at the same time. Many assays of insecticides and fungicides are in fact conducted by exposing several distinct batches to each dose of the poison. For example, in some forms of apparatus for the testing of insecticidal sprays not more than about twenty insects can be used at one time, and therefore, if conclusions based on approximately sixty insects at each dose are required, three batches must be tested. Examination of the variability between mortalities in batches given the same dose then provides a measure of the heterogeneity of the behaviour of the batches, including both biological differences between batches and variation from batch to batch in experimental technique. Comparison of this variability with the residual variation between doses (after the removal of the probit regression component) enables a test to be made of whether the latter can be explained as due only to the natural batch variation, and thus of whether the regression line is an adequate description of the relationship between dosage and response.

The usual procedure when several batches are used for each dose is merely to add the values of n (number of test subjects) and r (number killed) and to treat the results as if they referred to a single batch. Very often this course is satisfactory. If all batches contain the same number of subjects, the maximum likelihood estimates of the parameters are the same whether all subjects for one dose are treated as a single batch or the batches are kept distinct throughout, and, providing that the same provisional line is used, the estimates are the same at the end of each cycle of computations. This remains true for batches of unequal size as long as the total variance of the percentage mortality at a given dose is inversely proportional to n. Even when there is a component of batch variance independent of n, the batches will often be of nearly the same size, and the less onerous computations needed when batches are combined make that the generally preferred method.

When the residual variation between doses after the fitting of the linear probit regression equation gives a significant χ^2 , this may indicate either a real departure from linearity or a non⁴ independence of the responses of individuals of the same batch. If the latter explanation is adopted, all variances have to be multiplied by a heterogeneity factor, and, since the degrees of freedom available for the estimation of this factor are often very few, its estimate may be of low precision; fiducial limits will then vary irregularly in repeated determinations and will tend to be widely spaced on account of the increased value of t for few degrees of freedom. By using the data for each batch separately, not only may a test of linearity be derived but, supposing that there is heterogeneity between batches though no significant departure from linearity, a heterogeneity factor based on a greater number of degrees of freedom may be estimated.

At this point a brief digression on the order in which a series of tests should be carried out seems appropriate. Strictly speaking, the different batches of test subjects should be assigned to the doses entirely at random and the doses should be tested in random order. or in some restricted randomization in accordance with modern principles of experimental design (Fisher, 1942; Yates, 1937a). In practice, order of testing is usually held to be unimportant in a well-controlled experiment, provided that the whole is completed within a reasonably short time (with insecticides, preferably within one day); the theoretical requirements of randomization are therefore frequently sacrificed to the practical convenience of testing all batches at one dose consecutively and making all tests on one poison before those on a second are begun. The possibility of bias from this source must not be neglected, and every precaution to eliminate it must be taken lest any difference in susceptibility of insects tested at different

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times in the day be interpreted as a difference in potency of two poisons (McLeod, 1944). If the doses of a single poison are tested in ascending or in descending order, any steady trend in susceptibility of the subjects or any increase in the dose actually received (through incomplete cleaning of the apparatus after the preceding dose) may manifest itself by an increase or decrease in the estimated slope of the regression line or even by a departure from linearity; if the doses are given in random order, though these influences may increase the heterogeneity between batches they will not bias any estimates of potency. In insecticidal work the random selection of insects for the batches may be important; any method of selection which allows the more active insects to be used first, or which takes insects from a culture as they reach a certain stage of development, may have undesirable consequences through the batches being heterogeneous in respect of sex ratio or some other important factor correlated with susceptibility to the poisons under test (Murray, 1937; Bliss, 1939b). In other branches of biological assay analogous considerations arise, and, unless a strict randomization has been employed throughout, the experimenter should always be on his guard against any bias in his data resulting from non-random selection and order of testing.

When there is no evidence of heterogeneity of any type—and, with experience, this may often be judged from inspection of the data before any statistical analysis is made—there is nothing to be gained by maintaining the identities of different batches tested at each dose, and the less laborious analysis of combined batches should be adopted. When there is heterogeneity, such as is indicated, in the usual manner, by a significant residual χ^2 , a complete analysis may sometimes be made with advantage, in order to gain degrees of freedom for the heterogeneity factor, even though the estimates of potency will not be affected. If more than one cycle of computations is likely to be needed the detailed work should only be done in the last cycle, since the only use of the earlier ones is to improve the provisional line. The tests to be made can best be described by means of a numerical example. Ex. 30. The toxicity of ammonia to Tribolium confusum. Strand (1930, Table I) has given the results of tests of ammonia as a fumigant for T. confusum. Two batches of insects were tested at each of eight concentrations of the fumigant; from the form in which Strand states the results it seems likely that the tests were carried out as two distinct experiments, one batch in each, but for present purposes this point will be ignored and the pairs of batches will be assumed to be replicates of the type just discussed.

The computations necessary for fitting a probit regression line, when the batches are all kept separate, are shown in Table 27.

TABLE 27. Co	mputations	for	Ammonia	•Tril	bolium	confusum	Tests
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x	n	•	р	Em- pirical probit	Ŷ	nw	y	nwx	n	wy .
0.72	29 29	2 1	7 3	3.52 3.12	3.3	6·0 6·0	3·57 3·14	4·320 4·320	21·420 18·840	40.260
0.80	30 31	7 12	23 39	4·26 4·72	4·1	14·1 14·6	4·27 4·87	11·280 11·680	60·207 71·102	1 31·3 09
0.87	31 32	12 4	39 12	4·72 3·82	4 ·8	19·5 20·1	4·72 4·03	16·965 . 17·487	92·040 81·003	173.043
0.93	28 31	19 18	68 58	5·47 5·20	5.5	16∙3 18∙0	5·47 5·18	15·159 16·740	89·161 93·240	182.401
0·98	26 31	24 25	92 81	6·41 5·88	6 ∙0	11·4 13·6	6·32 5·87	11.172 13.328	72·048 79·832	151-880
1.02	27 28	27 27	100 96	∞ 6·75	6·4	8·2 8·5	6∙94 6∙67	8·364 8·670	56·908 56·695	113-603
1.07	26 31	26 29	100 94	∞ 6·55	6.9	4∙0 4∙8	7·34 6·42	4·280 5·136	29·360 30·816	60-176
1.10	30 31	30 30	100 97	∞ 6·88	7.2	2·8 2·8	7·59 6·75	3.080 3.080	21·252 18·900	40.152
						170.7		155-061	892.824	

 $1/Snw = 0.00585823, \quad \bar{x} = 0.9084, \quad \bar{y} = 5.2304.$

Snwx ²	Snwxy	Snwy [*]
142-46391	827.4877	4869-816
140-85480	811.0263	4669.799
1.60911	16-4614	200.017 168.402
		31.615

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They follow the usual plan (cf. Table 6), but, for use at a later stage, subtotals for each dosage are shown in the column *nwy*. In the table x is the logarithm of the ammonia concentration (in mg./l.) and the other symbols have their usual meanings. The residual sum of squares, 31.62, instead of being tested simply as a $\chi^2_{(14)}$ can be subdivided into two portions. The subdivision is accomplished by first calculating a sum of squares for y between dose levels, the expression for which is

$$(40 \cdot 260)^2 / 12 \cdot 0 + (131 \cdot 309)^2 / 28 \cdot 7 + ... + (40 \cdot 152)^2 / 5 \cdot 6 - (892 \cdot 824)^2 / 170 \cdot 7 = 4856 \cdot 852 - 4669 \cdot 799 = 187 \cdot 053;$$

the numerators of the fractions are the squares of the subtotals in the nwy column and the denominators the subtotals of nw for the doses. The analysis of variance of the mortality probits can now be completed as in Table 28. The first line is the sum of squares removed by the regression, the third line is the total sum of squares between the eight doses, the fifth line is S_{uu} , and the other lines are obtained by subtractions. The sums of squares in the second and fourth lines add to give the previously calculated residual, 31.62. The sum of squares in the fourth line is a $\chi^2_{[8]}$ which gives a test of the homogeneity of the results for different batches; its non-significance indicates that any heterogeneity is not sufficiently great to be disclosed by inter-batch variations. The sum of squares which measures departures from linearity, 18.65, if tested as a $\chi^2_{(6)}$, is judged significant, but inspection of the probit diagram (not shown here) discloses no systematic deviation from linearity. Since the two mean squares are not significantly different (Fisher and Yates, 1948, Table V) but both are greater than unity, their expectation for homogeneous data, the most reasonable conclusion to draw seems to be that heterogeneity between batches has increased both. This heterogeneity may be measured by combining the sums of squares to give a factor of $31 \cdot 62/14 = 2 \cdot 26$, with 14 degrees of freedom.

If the analysis had been made only on the totals for the eight doses, without using data from separate batches, only the first three lines of Table 28 would have appeared; in fact, 18.65 would then have arisen in the ordinary way as the residual χ^2 , and, since the probit diagram indicated no systematic non-linearity, its significance would have been interpreted as due to heterogeneity between batches. The heterogeneity factor would then have been taken as 3.11 with only 6 degrees of freedom. In either analysis, the regression equation is obtained as

$$Y = -4.063 + 10.230x,$$

and other parameters are estimated as required.

	D.F.	Sum of squares	Mean square
Regression	1	168-40	
Deviations from linearity	6	18.65	3.11
	I —		
Between doses	7	187.05	
Between batches at a dose	8	12.97	1.62
Total	15	200.02	

 TABLE 28. Analysis of Variance of Mortality Probits for Ammonia-Tribolium confusum Tests

Even when the number of batches is not the same for every dose the calculations follow exactly the same plan. If three or more batches are tested at each dose the number of degrees of freedom for differences between batches will be substantially greater than for deviations from linearity, thus enabling a more sensitive test of heterogeneity to be made and leading to a more precise estimate of the heterogeneity factor. In addition, a test for departures from linearity independent of that for heterogeneity can always be made, but, unless the probit diagram indicates systematic non-linearity, any significance in the linearity test should be interpreted as additional evidence of batch heterogeneity.

43. INDIVIDUAL MORTALITY RECORDS

In testing the effect of a drug or poison, it is sometimes impossible to feed the test subjects with predetermined doses; though the doses can only be roughly controlled, however, exact measurement of the amount taken may be possible afterwards. For

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example, some techniques for the testing of stomach insecticides. such as Campbell's poison-sandwich technique (Campbell and Filmer, 1929; Campbell, 1930), involve the feeding of separate amounts of poison to each insect, though the dose can only be measured after ingestion. Consequently the experimental results consist of a series of doses with, for each, a record of whether or not a single insect was killed. Such techniques tend to be more troublesome to carry out than those commonly employed for testing batches of insects at selected doses, and only comparatively short series are usually obtained. Tests on less than fifty insects in all cannot be expected to yield results of a precision comparable with that obtained when batches of this size are tested at each of several dosages. Nevertheless, the median lethal dose and other parameters can still be estimated by the method of probit analysis. Little modification of the instructions given in preceding chapters is required, but, as the analysis presents some unusual features, a brief account of it will be given. Bliss (1938) has given a similar but fuller discussion: his conclusions in respect of precision, however, must be treated with some reserve.

The first novelty lies in obtaining the provisional regression line necessary for the initiation of the computations. The observations show either zero or 100 % kill for each dose, and cannot therefore be plotted directly as probits. Instead, the results for a set of consecutive doses must be grouped so as to give percentage kills based on ten or more individuals, and the probits of these must be plotted against the mean log dose or other measure of dosage. In some experiments the doses themselves may indicate a convenient grouping, the experimenter having deliberately aimed at certain values. In others the doses may be spread fairly uniformly over the whole range so that the grouping has to be entirely arbitrary; non-independent overlapping groups of about ten might then be used in order to obtain more points, taking, say, the first to the tenth dosages as one group, the sixth to the fifteenth as a second, the eleventh to the twentieth as a third, and so on. A diagram showing the empirical mortality probit of each group plotted against the mean dosage allows a provisional

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line to be drawn in the usual manner; the grouped data tend to underestimate the slope of the line, which should therefore be drawn so as apparently to err slightly on the side of steepness.

From this stage, the improvement of the line may proceed exactly as described in § 17, each experimental dosage being used individually and each giving a maximum or a minimum working probit according as the corresponding insect was killed or survived. As usual, if the calculated line differs markedly from the provisional it should be used as a new 'provisional' for a second cycle of computations. When thirty or more doses have been tested, the process of working with 'batches of one' is tedious and seldom worth all the trouble involved. Grouping into small independent (non-overlapping) groups of four or five consecutive dosages greatly reduces the labour of finding sums of squares and products, and, though the regression coefficient will be slightly underestimated, the estimate of the log LD 50 will not be greatly affected provided that the experimental dosages are well distributed on either side of it. Difficulties of estimating a provisional line satisfactorily from meagre data frequently make it necessary to carry out two or more cycles of the computations, and, even though it may be intended to base the final line on the analysis of individuals, grouping may assist the rapid completion of the first cycles.

Unfortunately the troubles encountered with the routine χ^2 test for homogeneity (§ 18) arise in their most acute form in an analysis of individual mortality records, and are also severe when very small groups are used. A single instance of survival at a high dose or death at a low may inflate the value of χ^2 to an undue extent, as may easily be seen by considering, for example, the contribution to this χ^2 given by an observation of 1 in a class whose expectation is only 0.05. The sampling distribution of such a χ^2 is very different from that tabulated in Table VI, values usually being much lower than for a true χ^2 but there being also an excess of high values. If valid conclusions are to be reached by means of an ordinary χ^2 test, the method suggested in § 18 must be followed; after the calculation of expectations for each

dosage these must be further grouped so as to give reasonably large total expectations, both of dead and alive, in every group, and χ^2 must then be recalculated from these groups. If the total number of insects in the experiment is under fifty, the number of groups that can be formed will be small and the resulting test of homogeneity will not be very sensitive. On the other hand, no easily applied test of greater sensitivity is available. Indeed, even if the individual tolerances of these few individuals were measured directly, a significance test on the departure from normality of the distribution of their logarithms could not be. very sensitive; when the results are only obtainable as quantal responses a sensitive test is even less to be expected.

Bliss (1938) proposed to modify the usual test of heterogeneity by referring χ^2 , calculated according to equation (4.2), to a distribution with less than (k-2) degrees of freedom (where k is the number of groups). His rule involves classing as one group enough of the terminal groups at each end of the range of doses to make the expected number of survivors at the upper end and deaths at the lower end at least 10 % of the number of subjects in the standard group, even when the latter consists of only one: 'Thus if there were three in a standard group, the end groups at the upper end would be combined until 0.3 or more live animals was expected.' The number of degrees of freedom is then taken as two less than the number of groups after this combination. The test might seriously exaggerate the apparent significance of χ^2 , since that statistic is not recalculated from the combined groups. Only a full amalgamation of groups and recalculation of χ^2 can give an unbiased test. In a short series of tests the data will generally be insufficient to show any significant departure from the fitted line; if χ^2 calculated according to equation (4.2) is well below the significance level for (k-2) degrees of freedom (Table VI), no further test need be made and variances of estimates may be used without any heterogeneity factor. When a large value of χ^2 occurs, it may be necessary to recalculate on grouped data, or at least to investigate the possibility that an exaggerated effect of one or two anomalous observations is responsible.

No example of the computations for the fitting of the regression line is given here, as the initial determination of a provisional line is the only stage likely to trouble those who are familiar with the standard technique. A good illustration has been given by Bliss (1938) in the analysis of data on the toxicity of sodium fluoride to grasshoppers; the modified χ^2 test which he advocates has been criticized in the preceding paragraph, but inspection of these data shows there to be no significant heterogeneity. The approximate fiducial limits assigned to the LD 50 in this example are too narrow on account of the high variance of b (in fact g = 0.44); the exact formula (4.6) should have been used, and this gives limits of 0.062 and 0.135 mg./g. instead of 0.070 and 0.125 mg./g. Bliss has analysed these data both as individual records and by grouping in various ways; the estimated values of b differ quite widely, though they always lie within the range of sampling variation, but the estimates for the LD50 are remarkably consistent. Finney (1947b) gives another example.

A common practice in tests of this nature is to express the doses as amounts per unit weight of the test subject, in order to make some allowance for the varying size of these and their consequent probable variation in resistance. Thus the doses of sodium fluoride in the example used by Bliss were used as mg./g. of body weight. As an approximate method of adjustment this is not unreasonable, but a more exact approach would be to use the logarithm of the actual rather than the proportional dose as the dosage measure, and to make use of body weight as a concomitant variate. The assumption that resistance to the poison is directly proportional to body weight is thereby avoided, and instead the influence of body weight is estimated from the data. The computations may be carried out as in § 31, so that a probit plane is determined which relates mortality to log dose and log body weight; if desired, the effect of body weight may then be averaged out (§ 44).

In the light of the above discussion, little difficulty should be encountered in applying the methods of earlier chapters to individual mortality records. The various statistical techniques and formulae appropriate to comparisons of poisons, the action

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of mixtures, and multifactorial data can all be adopted even when the number of test subjects per batch is reduced to one. Obtaining a set of provisional probits for starting the computations is more troublesome when two or more dosage factors have to be considered in the same analysis, but instances of this are not likely to be encountered except by the more experienced workers to whom the estimation of some reasonable values will not be an insuperable obstacle; useful as it is to make a good first approximation, a poor one only delays the obtaining of satisfactory estimates, by requiring additional cycles of computations, and does not invalidate the final analysis.

44. THE AVERAGE KILL

Though not usually of great interest in insecticidal and fungicidal studies, the expected kill when a selected average dose is given, but the amounts received by individual test subjects vary, is sometimes required in other applications of the probit method. If the population has a true median lethal dose whose logarithm is μ , and the tolerance values of the log dose are normally distributed about this with variance σ^2 , the true probit regression equation is

$$Y=5+(x-\mu)/\sigma,$$

more commonly written as

$$Y=\alpha+\beta x,$$

where $\beta = 1/\sigma$. Suppose now that the logarithms of the doses received by the subjects are normally distributed about a mean ξ with variance γ^2 . Then the proportion killed will be the proportion receiving doses greater than their tolerance values. It might at first be thought that the result would be obtained by substituting ξ for x in the regression equation, but further consideration shows that the proportion also depends upon γ , being nearer to 50 % when γ is large compared with σ . By integration, the proportion is found to be that whose probit is \overline{Y} , where

$$\overline{Y} = 5 + (\alpha - 5 + \beta \xi) / (1 + \beta^2 \gamma^2)^{\frac{1}{2}}.$$
 (9.1)

In practice α and β have to be replaced by estimates a and b derived from experimental data. For example, if routine probit analysis has given a line

$$Y=3+2x,$$

corresponding to a log LD 50 of 1 and a variance of $\frac{1}{4}$ for the distribution of the logarithms of individual tolerances, equation (9.1) gives

$$\overline{Y} = 5 + (2\xi - 2)/(1 + 4\gamma^2)^{\frac{1}{2}}$$

Hence, if individuals are given doses whose logarithms are normally distributed about a mean of 1, the expected proportion killed is 50 % ($\overline{Y} = 5$), whatever the variance of the dosage distribution may be. If the mean log dose given is 2, the probit of the expected kill is nearly 7 (97.7 %) when γ , the standard deviation of the distribution of log doses, is very small, decreases to 6.41 (92.1 %) when γ is $\frac{1}{2}$, and decreases still further to a limiting value of 5 as γ increases indefinitely.

A slightly more complex situation arises when the mortality probit has been expressed, as in § 31, in terms of two dosage factors, say by the equation

$$Y = a + b_1 x_1 + b_2 x_2.$$

If the dosages to which the population is exposed are such that x_1 can be controlled but x_2 is normally distributed about ξ_2 with variance γ_2^2 , the mortality probit is still linearly related to x_1 , but the regression coefficient is reduced to

$$b_1' = b_1/(1+b_2^2\gamma_2^2)^{\frac{1}{2}}.$$

The one-factor regression equation is then

$$\overline{Y} = 5 + (a - 5 + b_2 \xi_2) / (1 + b_2^2 \gamma_2^2)^{\frac{1}{2}} + b_1' x_1.$$
(9.2)

This last equation is useful when x_2 , though formally a 'dose factor', is in fact a measurable characteristic of the individuals tested. For example, the potency of a poison may depend not only on the concentration but also on the weight of the subject. By subdivision of the data into weight classes, or by using individual records as described in § 43, the weight (or perhaps the logarithm of the weight) can be introduced as an x_2 into the

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probit regression equation. In order to predict the effect of any chosen concentration on a random selection of subjects, the contribution of weight to the equation must be averaged by means of equation (9.2). For this purpose ξ_2 and γ_2 may be estimated from the animals used for the toxicity test, provided that these were randomly selected from the whole population, or alternatively the parameters of the weight distribution may be estimated from measurements of a subsidiary random sample on which no toxicity tests have been made.

45. THE PARKER-RHODES EQUATION

A. F. Parker-Rhodes has made an extensive series of tests of the toxicity of metallic salts and other related compounds to spores of Macrosporium sarcinaeforme and Botrytis allii, and also to Bacillus agri. As a result of these tests, he has formulated a 'Theory of Variability', intended to explain, at least in part, the comparative toxic effects of different compounds (especially salts of the same metal) in terms of their chemical constitutions. Details of his ingenious theory, with experimental results, have been presented by the author in a series of papers (1941, 1942a, b, c, 1943a, b) to which the reader must be referred for the chemical and biological arguments. Discussion of the validity of these arguments is outside the scope of this book, but a brief outline of the statistical implications may be helpful to those concerned with fungicidal investigations of a like nature, the more particularly as Parker-Rhodes's treatment of this aspect seems scarcely adequate. For this purpose his notation will be brought into line with that of earlier chapters.

It has so far usually been assumed that the logarithm of λ , the tolerance of an individual test organism, is normally distributed, though mention has been made of the case of λ itself being normally distributed. O'Kane *et al.* (1930, 1934) published data from insecticidal studies which suggested that the logarithm of the mortality probit, rather that the probit itself, was linearly related to the log concentration. This theory may be expressed by the statement that the probit is proportional to a power of the dose. Parker-Rhodes's theory introduces the generalization

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that λ^i , a fractional power of the individual tolerances, shall be normally distributed. The relationship between mortality probit and dose thus becomes

$$Y = \alpha + \beta \lambda^i, \tag{9.3}$$

of which O'Kane's equation is a particular case (Bliss, 1935c) Parker-Rhodes defines i (for which he uses the symbol α) as the *index of variation*; i = 0 is a limiting case corresponding to the normal distribution of log tolerances, and i = 1 gives a normal distribution of tolerances. In general, the normalizing transformation for the dosage is

$$x=\lambda^i$$
,

the tolerances on the x-scale being normally distributed about a mean value μ with variance $\sigma^2 = 1/\beta^2$. This normal distribution can only be an approximation and must break down for small doses (assuming *i* to be positive); λ^i cannot be less than zero, and equation (9.3) therefore suggests that the kill approaches a minimal value represented by $Y = \alpha$ as λ tends to zero.

He defines the variability^{*} of the spores relative to the toxic substance under investigation as

$$W_i(\lambda) = \sigma^2/i^2\mu^2, \qquad (9.4)$$

which for the limiting case of i = 0 takes the form

$$W_0(\lambda) = \sigma^2. \tag{9.5}$$

Using estimates of σ and the median lethal dose obtained from the observations, an estimate of the variability is

$$U_i(\lambda) = 1/i^2 b^2 m^2, \qquad (9.6)$$

or, in the limiting case,

$$U_0(\lambda) = 1/b^2. \tag{9.7}$$

The properties of the variability and index of variation in relation to the chemical and fungicidal behaviour of the

* This definition of variability is likely to be confused with other aspects of the variation in the behaviour of the spores, and the name is an unfortunate choice for a precisely defined quantity. In order to follow Parker-Rhodes, the word will be used in the strict sense for the remainder of this section.

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compounds under investigation have been developed in the second of the series of papers (Parker-Rhodes, 1942*a*). The two chief results are:

'It is shown that the variability of a given population of spores to a compound which can penetrate the spore wall is less than to any other compound of the same element which cannot do so unless it undergoes one or more reactions on the surface of the spore with substances secreted by it, and that the greater the number of such successive reactions that are required to bring it to a permeable form, the greater will the variability be.

'It is shown that the variability of the spores to any compound is proportional to the square of the number of atoms of the effective element in a molecule of that compound, and that the index of variation is inversely proportional to that number, provided only one compound is permeative.'

Any study of a series of compounds in relation to the Theory of Variability must therefore pay special attention to the indices of variation and to the variabilities. In the most general case, not only have the parameters α and β to be estimated from the data, but also the index of variation. Experience indicates, however, that the index takes either simple fractional values or values so near these as to be indistinguishable from them. Indeed, Parker-Rhodes states (1943a): 'On a priori grounds, however, it appears that the index of variation must always be a rational number, and is more likely to be a simple fraction than a complicated one.' For example, in addition to the common values of 0 and 1, he finds (1942b) values very close to $\frac{1}{4}$ and $\frac{1}{4}$ (but see Ex. 32 below). Negative values, $i = -\frac{1}{3}$ and $i = -\frac{2}{3}$, have also been found (1943*a*), in association with negative values of β . A negative index of variation implies that, as the concentration is increased, the kill will approach, asymptotically, a maximum value corresponding to $Y = \alpha$; if this level is less than 50 %, formal solution of the equations will give an imaginary value for the median lethal concentration, the value of m (measured on the $x = \lambda^i$ scale) being negative. There may in truth sometimes be such a limiting mortality, but any expression of the situation in terms of 'imaginary LD 50's' is useless. In practice it seems

likely that, as in Parker-Rhodes's examples, at the higher concentrations some other phase of toxic action will supervene. Parker-Rhodes's statement that 'An imaginary value may be taken to imply absence of fungicidal potency' is misleading, even if the word 'potency' is considered to refer only to the one phase of toxic action, for though this phase may fail to attain an LD 50 it may well possess, say, an LD 45.

If the view that the index of variation is a simple rational fraction is accepted, the necessity of forming a statistical estimate from the data may be avoided; the subsequent calculations are then much simplified and the precision of estimation of α and β is increased. If the value of *i* can be decided either from previous experience or from a preliminary inspection of the data, the technique for estimating α and β is exactly as described in § 17, using $x = \lambda^i$.

Parker-Rhodes has apparently based his formula (1942b, c) for testing the significance of differences between two estimates of variability on the belief that, when there is no heterogeneity, for i = 0

$$V(U^{-\frac{1}{2}}) = 1/S_{xx},\tag{9.8}$$

and for other values of i

 $V(U^{-1}) = i^2 m^2 / S_{xx},$

m being, as usual, defined by

$$m = \bar{x} + (5 - \bar{y})/b.$$

The first of these equations is correct, but the second is easily seen to be entirely wrong. From equation (9.6)

$$U^{-1} = ibm$$

= $i(5 - \overline{y} + b\overline{x}),$
$$V(U^{-1}) = i^2 \left(\frac{1}{Snw} + \frac{\overline{x}^2}{S_{xx}}\right).$$
 (9.9)

whence

The criterion of significance at the 5% level given by Parker-Rhodes (1942b, p. 141) should therefore be amended to read

$$\frac{U_1 + U_2 - 2(U_1U_2)^{\frac{1}{2}}}{U_1U_2\{V(U_1^{-\frac{1}{2}}) + V(U_2^{-\frac{1}{2}})\}} \ge 3.84,$$

where the two variances are calculated according to equations $(9\cdot8)$ or $(9\cdot9)$, whichever is appropriate; the test may alternatively be considered as a test of the difference between U_1^{-1} and U_2^{-1} by means of their standard errors. When the heterogeneity χ^2 is significant, all variances should be multiplied by the heterogeneity factor, and the tests based on normal distribution of errors should be changed to the corresponding *t*-tests. In the early papers of the series, Parker-Rhodes used a conventional value of 50 or 100 for the heterogeneity factor, but in a note to the fourth (1942c), he recognized this to be wrong and suggested an amendment.*

Ex. 31. Variability of Macrosporium sarcinaeforme to hydrogen sulphide, sodium dithionite, and sodium tetrathionate. As part of the data from an investigation into the toxicity of various sulphur compounds to M. sarcinaeforme, Parker-Rhodes has published results for series of concentrations of hydrogen sulphide, sodium dithionite, and sodium tetrathionate (1942b, Tables 2, 5 and 6). The concentrations there given are expressed in arbitrary units, since the only use made of the data is for the estimation of variability, a quantity independent of the unit of concentration. Parker-Rhodes uses x instead of λ to represent concentration, q instead of p to represent proportionate mortality, and his n'is the number of sets of 50 spores counted. In his Table 8, Parker-Rhodes gives 0.95, 0.48 and 0.24 for the estimated indices of variation for the three compounds; these values are estimated from the data, by a method which is discussed and criticized below, but, in view of what has been said about likely values for the index of variation, they may reasonably be taken as 1, $\frac{1}{2}$ and 1. Table 29 gives details of the calculations for estimating the variability for sodium tetrathionate, on the assumption that $i = \frac{1}{4}$. The first stages are exactly as in the ordinary probit

* In the next paper (1943*a*) Parker-Rhodes introduced a form of the test equivalent to the use of the heterogeneity factor, though still involving the incorrect form for the variance of U^{-1} . The reference to the present writer at this point unintentionally suggests that he advocated the adoption of an 'admittedly false assumption'; in fact, his advice was not so revolutionary but only recommended bringing the test into line with other uses of the heterogeneity factor.

computations, except that x is taken as λ^{\ddagger} instead of $\log \lambda$.* Apparently Parker-Rhodes adjusted the values of p, the proportion of spores failing to germinate, so as to take account of a control mortality, but he gives no details, and for this example the modifications introduced in Chapter 6 have been ignored.

$x = \lambda^{\frac{1}{2}}$	n	р	Em- pirical probit	Y	nw	y	nwx	nwy
1.00	400	4.8	3.34	3.4	95	3.34	95.00	317.30
1.50	500	11.5	3.80	4 ·0	219	3.82	328·50	836-58
2.00	500	43.6	4.84	4.7	308	4.84	616-00	1490.72
2.51	500	64·1	5.36	5.3	308	5.36	773.08	1650-88
2.99	500	80·4	5.86	5.9	236	5.86	705·64	1382-96
3 ·50	500	91·5	6.37	6∙5	135	6∙36	472.50	858-60
					1301		2990.72	6537.04

 TABLE 29. Estimation of Variability of Macrosporium sarcinaeforme relative to Sodium Tetrathionate

 $1/Snw = 0.0007686395, \quad \bar{x} = 2.29879, \quad \bar{y} = 5.02463.$

Snwx ²	Snwxy	Snwy ²
752 3 ·79	15837.47	33884·16
6875·02	15027·25	32846.19
648.77	810-22	1037-97
		1011.85
		$26.12 = \chi^{2}_{[4]}$
	b = 1.24886,	••
	Y = 2.1538 + 1	1.2489x,
	$m=2\cdot 279,$	
	$\frac{1}{Snw} + \frac{\overline{x}^*}{S_{rr}} = 0.000769$	+0.008145
	= 0.008914	•

The computations present no new features until the regression coefficient, b, and the median lethal x-value, m, have been calculated. The fitted equation is

$$Y = 2 \cdot 1538 + 1 \cdot 2489 \lambda^{0.25}.$$
 (9.10)
$$U^{-4} = ibm = 0 \cdot 25 \times 1 \cdot 2489 \times 2 \cdot 279$$

It follows that $U^{-1} =$

$$= 10m = 0.25$$

= 0.712.

whence $U_{t}(\lambda) = 1.97$

is the estimated variability for sodium tetrathionate. Parker-Rhodes's estimate is 1.92; the difference is presumably due to

* x is easily obtained as the antilogarithm of $\frac{1}{2}\log \lambda$.

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different degrees of approximation in the maximum likelihood estimation of the regression line and to the slightly different indices of variation used, 0.25 and 0.24.

For the other two compounds, U^{-1} and U have been similarly calculated, and their values are shown in Table 30. In each case there is evidence of heterogeneity, the values of χ^2 being, for hydrogen sulphide

$$\chi^2_{[3]} = 8.60,$$

TABLE 30.Variability of Macrosporium sarcinaeforme relativetoHydrogen Sulphide, Sodium Dithionite, and SodiumTetrathionate

				Variabi	lit y (U)
Compound tested	Index of variation	V (U−i)	Ū−i	Present calcula- tions	Parker- Rhodes
Hydrogen sulphide Sodium dithionite Sodium tetrathionate		0-0548 0-0122 0-00352	$3.114 \pm 0.234 \\1.895 \pm 0.110 \\0.712 \pm 0.059$	0·103 0·278 1·97	0·104 0·272 1·92

for sodium dithionite (one concentration of the twelve tested was so high as to give zero weight)

$$\chi^{2}_{[9]} = 66.44,$$

and for sodium tetrathionate

$$\chi^2_{(4)} = 26.12.$$

Mean squares derived from these do not differ significantly, and the heterogeneity factor, obtained from

$$\chi^2_{(16)} = 101.16,$$

has the value 6.32. Now from Table 29

$$\frac{1}{Snw} + \frac{\overline{x}^2}{S_{xx}} = 0.008914$$

for sodium tetrathionate, whence, by equations (9.9),

$$V(U^{-\frac{1}{2}}) = 6.32 \times (0.25)^2 \times 0.008914$$

= 0.00352.

The standard errors of U^{-1} make it clear that variability differences between the three compounds are highly significant. The variance of the difference between any pair of values of U^{-1} is the sum of the corresponding variances tabulated in the third column; if an exact test were needed, a *t*-test with 16 degrees of freedom (the number of degrees of freedom on which the heterogeneity factor is based) would show whether or not the difference was significantly greater than zero.

When the estimation of the index of variation from the data seems necessary, a more complex procedure must be adopted. Parker-Rhodes (1942a) has discussed this problem and has suggested a method of obtaining the maximum likelihood estimate. Comparison with the discussion of the general maximum likelihood equations in Appendix II shows his method to be at fault in several respects. The estimates obtained by it will frequently not differ greatly from the true maximum likelihood values, but since the correct method is no more difficult or laborious to apply there seems every reason for preferring it.

Equations (II, 3) may be adapted to the estimation of the parameters of equation $(9\cdot3)$ in much the same way as they were adapted in § 28 to give equations (6.4). The most convenient method of computation is first to tabulate an auxiliary variate^{*}

$$x' = \lambda^i \log_e \lambda. \tag{9.11}$$

If a provisional value is taken for *i*, and a provisional regression line of probits against $x = \lambda^i$ drawn, weights and working probits can be formed as usual. Calculation of the regression of y on x and x' then gives revised estimates:

$$\frac{bS_{xx} + (b_0 \delta i)S_{xx'} = S_{xy}}{bS_{xx'} + (b_0 \delta i)S_{x'x'} = S_{x'y}}$$
(9.12)

$$a = \overline{y} - b\overline{x} - (b_0 \delta i) \overline{x}', \qquad (9.13)$$

* The safest procedure is to use natural logarithms in determining each value of x': if no tables of these are available, multiply logarithms to base 10 by 2.303. Logarithms to base 10 may be used, with appropriate adjustments at the end of the calculations, but the danger of mistakes is greater.

and

where b_0 is the slope of the provisional regression line, a, b, and $(i + \delta i)$ the parameters of a revised equation

$$Y = a + bx^i. \tag{9.14}$$

The process, which is analogous to that of §28, may be iterated, using equation (9.14) as a new provisional line, until successive values of the parameters are not appreciably different.

Estimation of the variances of the index of variation and of the variability will not be discussed in detail here. A variance matrix, V, for the parameters is obtained by the methods of Appendix II (cf. equation (6·13)). Unfortunately, discrimination between the effects of small alterations in b and small alterations in i is difficult; the sampling variations of the parameters are usually closely correlated, and several cycles of computation may be needed in order to locate the maximum likelihood estimates satisfactorily. The variance of b will be much greater than it would have been if i had been known a priori, and, without data vastly more extensive than are usually available, precise estimation of the index of variation will seldom be possible. The variability, however, is more easily determinable, and is not very sensitive to small changes in i, since sampling errors in b and i are often compensatory.

Ex. 32. Maximum likelihood estimation of index of variation of Macrosporium sarcinaeforme relative to sodium tetrathionate. Further examination of the sodium tetrathionate data analysed in Ex. 31 indicates that the value of $\frac{1}{4}$ taken for the index of variation is substantially higher than the maximum likelihood estimate. After several cycles, the last starting from b = 4.264 and i = 0.112, the equation

$$Y = -0.9312 + 4.1053\lambda^{0.1153} \tag{9.15}$$

was obtained. The measure of heterogeneity, now with 3 degrees of freedom because an extra parameter has been estimated, is

$$\chi^{2} = S_{yy} - bS_{xy} - (b_{0}\delta i)S_{x'y}$$

= 15.25, (9.16)

which gives 5.08 as the heterogeneity factor. The difference between the χ^2 values in Exs. 31 and 32 is the improvement due

to estimating *i* (1 degree of freedom); as a test of the significance of the difference between the estimated index of variation and its previously assumed value, this should be compared with the residual mean square, using a variance ratio test since the second χ^2 is clearly significant (Table 31). Reference to Fisher and Yates (1948, Table V) shows the ratio of mean squares to be nonsignificant, but, with so few degrees of freedom, the test is insensitive. In Table 32, the observed percentage mortalities are compared with those estimated by equations (9·10) and (9·15).

TABLE 31. Test of Significance for the Difference between the Estimated Index of Variation and its Previously Assumed Value

Source of variation	D.F.	Sum of squares	Mean square
Removed by i	1	10.87	10.87
After estimating a, b, i	3	15-25	5.08
After estimating a, b only	4	26.12	

 TABLE 32. Comparison of Two Probit Equations fitted to

 Sodium Tetrathionate-Macrosporium sarcinaeforme Data

(i) $Y = 2.1538 + 1.2489\lambda^{0.25}$ (ii) $Y = -0.9312 + 4.1053\lambda^{0.1158}$

,	_	P	ercentage morta	lity
~	n	Observed	Equation (i)	Equation (ii)
1	400	4.8	5.5	3.4
5	500	11.5	16.4	16-1
16	500	43.6	36.4	39.0
40	500	64-1	61.6	63.7
80	500	80.4	81.3	80.9
150	· 500	91.5	93.6	91.7

The matrix of variances and covariance for b and $b_0 \delta i$ is the inverse matrix formed in the solution of equations (9.12), namely,

$$V = \begin{pmatrix} v_{11} & v_{12} \\ v_{12} & v_{22} \end{pmatrix} = \begin{pmatrix} 4 \cdot 8600 & -0 \cdot 40551 \\ -0 \cdot 40551 & 0 \cdot 033965 \end{pmatrix},$$

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multiplied by the heterogeneity factor. Hence the standard error of b, with only 3 degrees of freedom, is 4.97, much greater than when i was assumed known (Ex. 31). Also

$$V(i) = 5.08v_{22}/b_0^2$$

= 0.0095;

though i = 0.1153 is very different from the value 0.25 assumed in Ex. 31, the standard error of the estimate, 0.97, is so great as not to discriminate clearly between the two, and, as shown by Table 31, the data do not definitely contradict the hypothesis that the true index of variation is 0.25.

From equations (9.6) and (9.15), $U^{-\frac{1}{2}}$ may be estimated; its variance is given by the lengthy formula

$$V(U^{-1}) = i^2 \left(\frac{1}{Snw} + \bar{x}^2 v_{11} + 2\bar{x}\bar{x}' v_{12} + \bar{x}'^2 v_{22} \right) + 2ibm(\bar{x}v_{12} + \bar{x}' v_{22})/b_0 + (bm)^2 v_{22}/b_0^2, \quad (9.17)$$

which must be multiplied by the heterogeneity factor when there is one. The result of these calculations is

$$U^{-\frac{1}{2}} = 0.684 \pm 0.054,$$

in good agreement with Table 30, thus illustrating the stability of the variability (now estimated as $2 \cdot 14$ instead of $1 \cdot 97$). The reduction in the standard error of U is an effect of the smaller heterogeneity factor (5.08 instead of 6.32). A composite heterogeneity factor from the three compounds would have more degrees of freedom and would provide a more reliable standard error.

Chapter 10

GRADED RESPONSES

46. THE LINEAR DOSAGE-RESPONSE CURVE

THE discussion in earlier chapters has been concerned only with quantal characteristic responses. In many biological investigations it is possible not merely to state that a subject has responded to the treatment applied but also to measure the magnitude of that response. Vitamin preparations, for example, are often assaved by comparing the weight increases of rats fed for a specified period on suitable doses with the weight increases of other rats fed for the same time on a similar range of doses of a standard preparation with known vitamin content. Again, insulin may be assayed in terms of the percentage fall in blood sugar of injected rabbits. All such data could be reduced to a quantal form by a simple dichotomy of the measurements into those greater than and those not greater than an arbitrarily selected value; the relationship between the dose and the percentage of subjects whose responses exceed this value could then be studied by the methods of probit analysis already described. This procedure, however, would be very wasteful, as it discards entirely the information provided by the distribution of the magnitudes of responses within the 'greater than' and 'not greater than' classes.

When quantitative responses to a stimulus are measured over a sufficiently wide range of doses, some curvature of the relationship between dose and mean response will almost certainly become evident. Indeed, the response curve will frequently be sigmoid in type. Nevertheless, especially if the responses are plotted against a logarithmic dosage scale, the curve often does not differ appreciably from a straight line over a considerable section of the dosage range. The central section is usually that of chief interest to the experimenter, and he may reject the results for the more extreme levels of dose which give responses outside the linear range in order to have data which can be

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analysed by the ordinary linear regression technique (Fisher, 1944, §§ 25, 26). The accurate estimation of the regression slope requires that the whole of the linear section should be included in the tests, but pre-existing knowledge may enable economies in experimental material to be effected by omitting very extreme doses which are almost certain to lie outside this range.

Extreme responses will thus be avoided, and it may then be reasonable to assume that the variance of the response of each subject is constant, irrespective of the dose. A linear regression equation of mean response on dosage may therefore be calculated, weighting the mean response at each dosage in proportion to the number of subjects. Since these weights are independent of the expected responses, the complications of probit analysis do not arise and no method of successive approximations is required in order to obtain the maximum likelihood solution. Burn (1950) and Coward (1938) have used regression analysis extensively in a variety of problems of biological assay from quantitative responses. Bliss and Marks (1939a, b) have given an excellent description of the statistical analysis of an insulin assay, showing details of the computations and discussing many important points, including the use of covariance analysis for improving the precision of the estimate of relative potency by making adjustments for preliminary variation in relevant measurable characteristics of the experimental rabbits. The same considerations in respect of fiducial limits as are outlined for quantal response data in § 19 arise for quantitative responses also, and the general formula (4.7) may be applied (Irwin, 1943; Fieller, 1944).

Details of the application of standard linear regression technique to assay data will not be given here, for, though the underlying theory is comparatively simple, the methods could not be adequately illustrated without many examples, and a full account falls outside the scope of this book. In addition to references already given, Bliss (1940*a*), Bliss and Rose (1940), Fieller (1940), Finney (1945*a*), and Irwin (1937) may be consulted for useful examples of experimental planning and the statistical reduction of the results. Before leaving this topic, however, one small point of interest may be noted in connexion with what has come to be known as the four-point assay, the very simple type of assay in which a test material and a standard are each used at two rates only; the difference between the higher and lower rates is the same for both materials on the dosage scale used, thus implying that, if a logarithmic dosage scale has been adopted, the two pairs of doses are in the same ratio. Gridgeman (1943) and Wood (1944a) have shown that for this type of assay the relative potency may be estimated even when there is an appreciable departure of the dosage-response relationship from linearity over the range of doses tested. Provided that the relationship can be adequately represented by a second-degree equation, the formula for relative potency derived on an assumption of linearity still gives an unbiased estimate. The four-point assay may nevertheless give misleading results because of the very little evidence it supplies on the identity of shape of the response curves for the two materials tested; the conditions of its usefulness and validity have been discussed by Finney (1944b). Gridgeman (1944) and Wood (1944b).

47. QUANTITATIVE RESPONSES AND THE PROBIT TRANSFORMATION

When quantitative responses are studied over an unrestricted range of doses, they will frequently be found to show a sigmoid relationship with dose, and by a suitable choice of dosage scale (again often logarithmic) the relationship may usually be made to approximate to the normal sigmoid form. Though the probit transformation was devised as an aid to the analysis of quantal response data, its property of converting a normal sigmoid curve into a straight line is not dependent upon the source or nature of the data, and the same end will therefore be achieved if the sigmoid represents the relationship between dosage and a quantitative response. There is, however, an important difference between the statistical procedures for the two types of data. For quantal responses the proportion of subjects responding to any dose is assessed as the ratio of the number showing the characteristic response to the total number receiving the dose; for quantitative responses the mean response to each dose can be calculated from the data, but in general no maximum possible response is known and consequently no proportional response can be directly calculated. Sometimes, as in Ex. 33 below, one subject or set of subjects can be given a dose such that each will show its individual maximum, but this can only be an estimate of the mean for all subjects, since it is itself subject to the natural variability of the population. The difficulty is similar to that of correcting quantal data for the proportion of responses amongst the controls (Chapter 6), but has to be faced relatively more frequently. Indeed, the rarity of investigations in which quantitative responses can be reckoned as proportions of a known maximum probably accounts for the failure to realize the possibility of applying the probit transformation to quantitative data (Bliss, 1941; Finney, 1943b).

The variance of the response of all test subjects receiving the same dose can usually be estimated empirically from the separate observed responses to that dose, and thus independently of the expected response to that dose; the variance of the mean response then takes the form v/n, where v is the variance of a single response and n is the number of responses contributing to the mean. At low dosages the mean response tends to zero, and, unless negative responses of individual test subjects are possible, v must also become very small; at high dosages also, when the mean response approaches its maximum, v may again become small. Nevertheless, unless very extreme doses are used, v may often be taken as substantially constant and may then be assessed by pooling estimates for different dose levels.*

If the mean response to n subjects tested at dosage x (on the normalizing scale) is u, the expected value of u may be written

$$U = HP; (10.1)$$

* An approximate test of the heterogeneity of a set of variance estimates $v_1, v_2, ..., v_k$, with degrees of freedom $f_1, f_2, ..., f_k$ is given by treating $2 \cdot 3026(f \log v - Sf_i \log v_i)$

as a χ^2 with (k-1) degrees of freedom. The summation is to be taken over the k values, and $f = Sf_i$, $fv = Sf_iv_i$; v is the pooled estimate of variance to be used if the test discloses no heterogeneity (Bartlett, 1937).

here H is the mean limiting response for high dosages, and P, the proportionate response at this dosage, is as defined by equation (3.1). When H is known, and therefore does not have to be estimated from the data,

$$p = u/H \tag{10.2}$$

is an estimate of P. The probits of the several values of p may then be fitted by a linear regression on x by the method of § 17, except that the weight to be attached to each point is $\operatorname{not} nZ^2/PQ$ but nH^2Z^2/v . The standard errors of parameters estimated from the analysis are proportional to \sqrt{v} and are based on the same number of degrees of freedom as v instead of pertaining to a normal distribution. Instead of a χ^2 test of the significance of the departures of the observations from the fitted probit regression line, a variance ratio test must be used, as illustrated in Ex. 33.

In the present chapter, a weighting coefficient, w, defined by

$$w=Z^2, \qquad (10.3)$$

will be used instead of that defined by equation (3.4). Values of both Z and Z² are given in Table V for values of Y at intervals of 0.1 from 1.0 to 9.0; both functions are symmetrical about Y = 5.0. For the simple problem of fitting the probit regression line when H is known, the only differences from the procedure of § 17 are the changed weighting coefficient and the introduction of a factor H^2/v into all weights, or v/H^2 into all variances, at the end of the analysis. The working probit is not affected by the change in weighting.

When H is unknown, three parameters must be estimated from the data. An auxiliary variate

$$x' = P/Z \tag{10.4}$$

is introduced, its values being obtainable by entering the Q/Z column of Table II with (10 - Y) instead of Y. If H is a provisional value for the maximum response, and h is the observed mean response to n_h subjects tested under the 'control' conditions for which P = 1, an adjustment, δH , to H and revised parameters for a regression line are given by

$$bS_{xx} + \frac{\delta H}{H}S_{xx'} = S_{xy},$$

$$bS_{xx'} + \frac{\delta H}{H}S_{x'x'} = S_{x'y}$$
(10.5)

and

 $a = \bar{y} - b\bar{x} - \frac{\delta H}{H}\bar{x}'.$ (10.6)

Contributions from the controls, n_h and $n_h(h-H)/H$, must be added to $S_{x'x'}$ and $S_{x'y}$ respectively (cf. §28). The estimates are obtained by a formal calculation of the multiple regression of yon x, x', the values of x', w, y being based on a provisional line. In the limit of successive cycles, the inverse matrix of the coefficients on the left-hand side of equations (10.5) gives the variances and covariance of b and $\delta H/H$, except for the omitted factor v/H^2 (and a heterogeneity factor, if required).

Ex. 33. The repellent effect of lime sulphur on the honeybee. Butler et al. (1943) have discussed a series of experiments on the attractiveness or repellency to the honeybee of various constituents of orchard sprays. In one of these experiments individual cells of dry brood comb were filled with measured volumes of emulsions of lime sulphur in M/1 sucrose solution, and were placed in an experimental chamber. Seven different concentrations of lime sulphur were used, ranging from 1/100 to 1/1,562,500 by successive factors of one-fifth. Eight cells of each of these, together with eight cells of M/1 sucrose solution alone, were arranged in eight rows of eight to form an 8×8 Latin square, thus eliminating as far as possible any positional effects within the chamber. Additional non-experimental cells of sucrose solution were placed between the rows and around the square so that there should be no shortage of the unadulterated solution. About 100 honeybees were then released into the chamber for two hours, after which the volume of liquid within each cell was again measured. The difference between the two volumes, corrected by a very small amount representing the evaporation during the experiment, is the quantity taken up by the bees. Table 33 shows the arrangement of the experimental cells and the corrected uptake figures for each; totals of rows and columns of eight cells are also shown. The effectiveness of lime sulphur in repelling the honeybees is measured by comparing the total uptakes from the eight cells of each treatment, shown in Table 34.

TABLE 33. Lay-out and Results of Experiment on Uptake of Lime Sulphur by the Honeybee

(Uptake recorded in mg. per cell)

									row
									totals
	D	С	F	H	E	Α	B .	G	
	57	84	87	130	43	12	8	80	501
	\mathbf{E}	в	н	Α	D	С	G	F	
	95	6	72	4	28	29	72	114	420
	В	н	А	\mathbf{E}	G	F	C	D	
	8	127	5	114	60	44	13	39	410
	н	D	\mathbf{E}	С	Α	G	F	в	
	69	36	39	9	5	77	57	14	306
	G	E	D	F	С	В	А	н	
	92	51	22	20	17	4	4	-86	296
	F	A	С	G	в	D	\mathbf{H}	Е	
	90	2	16	24	7	27 [·]	81	55	302
	С	F	G	в	н	\mathbf{E}	D	Α	
	15	69	72	10	81	47	20	3 [.]	317
	A	G	в	D	F	н	Е	С	
	2	71	4	51	71	76	61	19	355
Column	_								
totals	428	446	317	362	312	316	316	410	2907

The meaning of the symbols for the eight treatments is given in Table 34.

TABLE 34.	Uptakes of Various Concentrations
of I	ime Sulphur by Honeybees

		Treatment		Total of 8 cells (mg.)	Mean per cell (mg.)
A: <i>M</i>	/l sucrose	+ lime sulpl	nur (1 %)	37	4.6
в:	**	,,	(0-2 %)	61	7-6
C:	,,	**	(0.04 %)	202	25-2
D:			(0.008 %)	280	35.0
E:	,,	**	(0.0016 %)	505	63-1
F:	**		(0·00032´%)	552	69-0
G:	••		(0.000064 %)	548	68.5
	/l sucrose		()	722	90.2
Stand	ard error o	f mean		1 1	± 6·9

An analysis of variance of the 64 uptakes involves the assumption that the error variance is the same for all of them. In fact there are strong indications that the variance increases with decreasing concentration of lime sulphur, and especially that

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the uptakes of the four highest concentrations are much less variable than those of the four lowest. Experience has shown that the validity of conclusions from an analysis of variance is not seriously upset unless the differences in variability are very great, and special investigation of these data indicates that the final estimates obtained would not be much altered if they were based on a variance which itself depended upon the concentration. The standard procedure for the analysis of Latin square data (Fisher, 1944, § 49; Mather, 1943, § 29) has therefore been adopted as a means of estimating the variance per cell. Each row of eight cells in the experimental lay-out contains one cell of each treatment, as also does each column of eight cells, and the variation between the sets of row and column totals is eliminated in the statistical analysis.

The completed analysis of variance is given in Table 35. A total sum of squares of deviations of the 64 measurements about their mean is calculated as

 $57^2 + 84^2 + 87^2 + 130^2 + \ldots + 61^2 + 19^2 - (2907)^2/64 = 79730.$

	D.F.	Sum of squares	Mean square
Rows	7	4,768	
Columns	7	2,808	
Treatments	. 7	56,160	8,023
Error	42	15,994	380.8
Total	63	79,730	

TABLE 35. Analysis of Variance of Data in Table 33

A component of this sum of squares representing the variation in row totals, and having 7 degrees of freedom, is then calculated from the totals shown in Table 33 as

$$(501^2 + 420^2 + \ldots + 355^2)/8 - (2907)^2/64 = 4768,$$

the divisor 8 occurring since each row has eight cells. Similar calculations give components for columns and for treatments, the latter being obtained from totals shown in Table 34 as

 $(37^2 + 61^2 + \ldots + 722^2)/8 - (2907)^2/64 = 56160.$

The difference between these three components and the total is a sum of squares with 42 degrees of freedom which measures the residual variation, and the mean square obtained when this residual is divided by 42 is the estimate of error variance,

$$v = 380.8.$$

A rapid inspection of the data is sufficient to show that the differences between the uptakes of the eight concentrations are much too large to be attributed to random variation. If a test of significance were required, it would consist in comparing the mean squares for 'Treatments' and 'Error'; according to Fisher and Yates's table of the 5 % levels of the variance ratio (1943, Table V), a ratio greater than $2\cdot24$ must be considered indicative of real differences between treatments, and in this analysis the ratio is over 20.

The standard error of the treatment means, each a mean for eight cells, is $\pm \sqrt{(v/8)}$, or ± 6.9 mg. per cell. The means are given in Table 34, and are also shown in Fig. 21 plotted against a logarithmic scale of dosage; on this scale, for convenience, the 0.008 % (= $5^{-3} \%$) concentration has been taken as zero and other doses expressed relative to it by logarithms to base 5, so that

 $x = 3 + \log_5$ (percentage concentration). (10.7)

Sucrose alone is a zero concentration of lime sulphur and therefore has an infinite negative value of x. Though a straight line would fit the points for the seven concentrations of lime sulphur tolerably well, a sigmoid of some type is necessary to show the eventual tapering away to zero uptake at very high concentrations and the approach to the uptake of unadulterated sucrose solution at very low concentrations. A straight line would be an entirely inadequate representation of the relationship much outside the range of concentrations tested, and indeed the points do suggest a sigmoid curve.

Apart from the reversal in the direction of the dose effect, so that increasing the dose decreases the response, the sigmoid required does not look to be markedly different in shape from the normal sigmoid illustrated in Fig. 4, and the frequent success of log dose as a normalizing transformation encourages its use here. In this experiment the test subjects of the earlier discussion are the cells, not the bees, so that there are eight 'subjects' at each concentration; the response, u, is the uptake of liquid from a cell during the course of the experiment. Since u decreases with increasing concentration from the value for sucrose at zero concentration to zero at very high concentrations, for the probit regression line b must be negative.

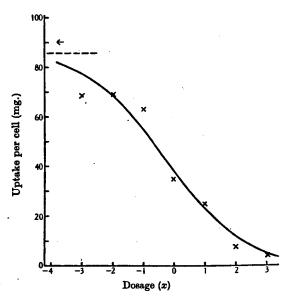


FIG. 21. Uptake of lime sulphur by the honeybee, showing normal sigmoid curve (Ex. 33). \leftarrow indicates uptake of zero concentration. Broken line indicates mean limiting response.

The mean uptake of liquid from the cells containing only sucrose solution is 90 mg. per cell, so that the estimate of the parameter H from these control cells alone is $\hbar = 90$. On the other hand, the two lowest concentrations of lime sulphur tested gave responses of just under 70 mg. per cell, thus suggesting that H is not so great as 90. In obtaining equations (10.4) a provisional value of 85 was taken for H, a guess which proved to be remarkably good. The computations then proceed as in Table 36. Values of u, the mean uptake at dosage x, are copied from Table 34, and p = u/85 is calculated for each. The empirical probits of p are tabulated, and are plotted in Fig. 22, in which figure a provisional regression line is drawn. From this line expected probits, Y, are read, and weights and working probits are determined in the usual manner, except that w must be taken from the Z^2 column of Table V instead of from Table II; thus for the third dosage Y = 4.4, and the weight is therefore $nw = 8 \times 0.111$. The quantities P/Z are obtained from Table II as the values of Q/Z which correspond to (10 - Y).

TABLE 36. Computations on Uptake of Lime Sulphur by Honeybees

z	n	u	p(H=85)	Em- pirical probit	¥	nw	y	x'=P/Z	nwx	nwy	nwx'
$ \begin{array}{r} 3 \\ 2 \\ 1 \\ 0 \\ -1 \\ -2 \\ -3 \\ -\infty \\ -\infty \\ \end{array} $	8 8 8 8 8 8 8	4.6 7.6 25.2 35.0 63.1 69.0 68.5 90.25	0.05 0.09 0.30 0.41 0.74 0.81 0.81	3.36 3.66 4.48 4.77 5.64 5.88 5.88	3·4 3·9 4·4 5·4 5·9 6·4	0·10 0·38 0·89 1·26 1·08 0·57 0·18	3·36 3·69 4·48 4·77 5·63 5·63 5·68 5·67	0·49 0·62 0·82 1·16 1·78 3·07 6·14	$\begin{array}{r} 0.30 \\ 0.76 \\ 0.89 \\ 0.00 \\ -1.08 \\ -1.14 \\ -0.54 \\ - \end{array}$	0-3360 1-4022 3-9872 6-0102 6-0804 3-3516 1-0206	0-0490 0-2356 0-7298 1-4616 1-9224 1-7499 1-1052
						4.46			-0.81	22.1882	7.2535

$\bar{x} = 0.1816, \ \bar{x}' = 1.6263, \ \bar{y} = 4.9749$

Snwx ¹	Snwxx'	Snwx' ²	Snwxy	Snwx'y	Snwy ²
8-2900	- 7.3898	18.0440	- 8 0458	38.6543	112.5612
0.1471	-1.3173	11.7967	<u>-4.0297</u>	36.0857	110.3848
8-1429	-6.0725	6.2473	-4.0161	2.5686	2.1764
	nh	= 8.0000	$n_h(h-H)/h$	H = 0.4941	
		14.2473		3.0627	

Table 36 is completed in the obvious manner, and equations (10.5) written down as

$$8 \cdot 1429b - 6 \cdot 0725 \frac{\delta H}{H} = -4 \cdot 0161,$$

$$- 6 \cdot 0725b + 14 \cdot 2473 \frac{\delta H}{H} = -3 \cdot 0627.$$
(10.8)

The inverse matrix of coefficients is .

$$V = \begin{pmatrix} v_{11} & v_{12} \\ v_{12} & v_{22} \end{pmatrix} = \begin{pmatrix} 0.180029 & 0.076732 \\ 0.076732 & 0.102894 \end{pmatrix}, \quad (10.9)$$

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and by summation of the products of each row of V by the righthand sides of equations (10.8)

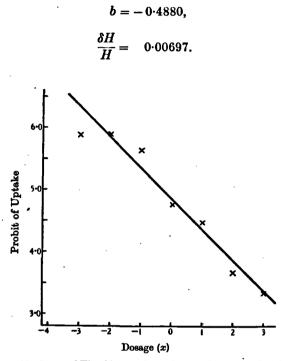


FIG. 22. Data of Fig. 21 transformed to probit scale, showing probit regression line (Ex. 33).

The improved estimate of the maximum response, $H + \delta H$, is .85.59, and, by equation (10.6),

$$a=4.875.$$

The calculations could be repeated, using H = 85.59 and

$$Y = 4.875 - 0.488x \tag{10.10}$$

as a provisional line, but it is easily seen that changes from the first cycle would be negligible by comparison with the experimental errors.

Table 37 shows the result of using equation $(10 \cdot 10)$ to calculate expected probits, expected proportions, and expected uptakes, U, for each x; the sigmoid curve in Fig. 21 has been plotted from these values of U. Comparison with the observed mean responses, u, shows some irregularities but no systematic deviation. A test of significance of the discrepancies may be based upon the sum of squares of differences (u - U). Since each u is a mean of eight observations, $S(u - U)^2$ must be multiplied by 8 to bring it to the same units as those of Table 35; the result, 1663, is a residual sum of squares with 5 degrees of freedom, three parameters having

 TABLE 37.
 Comparison of Observed and Expected

 Lime Sulphur Uptakes

x	Y	Р	U = HP	u
3	3.411	0.056	4.8	<u>4</u> ∙6
2	3·899	0.135	11.6	7.6
1	4.387	0.270	23.1	25.2
0	4.875	0.450	38.5	35.0
-1	5.363	0.642	55.0	63-1
-2	5.851	0.803	68.7	69·0
-3	6.339	0.910	77.9	68.5
- 00		1.000	85-6	90.2

 $S(u-U)^{1} = 207.92.$

been estimated from the eight means. The mean square, 1663/5 = 333, is less than the error mean square in Table 35, and therefore gives no indication of serious deviations from the fitted sigmoid curve. An alternative procedure is to follow the pattern of calculation which led to a χ^2 in §28; the expression

$$S_{yy} - bS_{xy} - \frac{\delta H}{H}S_{x'y} = 0.1952$$
(10.11)

is no longer a χ^2 , but multiplication by H^2 (=7326) will give the residual sum of squares. The result, 1430, differs appreciably from the 1663 obtained by the first method because the basic

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calculations have not been repeated sufficiently often to approximate as closely to the maximum likelihood limiting values as would be required in a critical case. Here the practical conclusion is the same by either test. Had non-systematic heterogeneity of deviations from the fitted curve been found, the mean square with 5 degrees of freedom would have been used in subsequent calculations, in place of $s^2 = 380.8$ with 42 degrees of freedom.

Now
$$V(b) = s_1^2 v_{11} / H^2 = 0.009489,$$
 (10.12)

and $V(H) = s^2 v_{22} = 39.18;$ (10.13)

hence the standard errors of b and H are 0.097 and 6.26 respectively, which confirm that the changes from the provisional values to the results of the first cycle are small by comparison with experimental errors. If the ED 50 is defined as the concentration of lime sulphur producing a 50 % reduction in sucrose uptake, the log ED 50 is found as

$$m = \overline{x} + \left(5 - \overline{y} + \frac{\delta H}{H} \overline{x}'\right) / b \qquad (10.14)$$
$$c = -0.256.$$

For the 5 % level of probability, t = 2.02, and

 $g = t^2 V(b)/b^2 = 0.163,$

too large to permit the assessment of fiducial limits to m by means of a standard error formula. Formula (4.7) must therefore be applied to the expression for $(m-\bar{x})$ given by equation (10.14), using V(b) from equation (10.12),

$$V\left(5-\bar{y}+\frac{\delta H}{H}\bar{x}'\right) = s^2\left(\frac{1}{Snw}+\bar{x}'^2v_{22}\right)/H^2 = 0.026161,$$

d
$$\operatorname{Cov}\left\{\left(5-\bar{y}+\frac{\delta H}{H}\bar{x}'\right),b\right\} = s^2\bar{x}'v_{12}/H^2 = 0.006577.$$

and

The fiducial limits to m are found to be 0.356 and -1.167.

Now equation (10.7) may be written

 $\log_{10}(\% \text{ concentration}) = 0.699x - 2.097,$

from which the ED 50 of lime sulphur is estimated to be 0.0053 %, with 5 % fiducial limits at 0.0142 and 0.0012 %.

Many of the methods of analysis discussed in earlier chapters can be modified for use with quantitative response data of the type under consideration here. In particular, the estimation of the relative potency of two stimuli whose probit regression lines are parallel, the analysis of data involving two or more dose factors, and the study of mixtures, can all be effected by techniques analogous to those used with quantal responses. The chief differences are the change in the weighting coefficient and the frequent necessity of estimating the third parameter, H. Consequently the computations are more laborious than those of ordinary probit analysis, but they are by no means prohibitively so. Additional examples will not be given here, as the assumption of a linear dosage-response relationship is so often good enough; the probit procedure for many types of data should be apparent by analogy with the corresponding quantal response problems. McCallan (1943) has suggested a further modification, for use with certain types of quantitative data, in which an empirical relationship between weighting coefficient and response is estimated as a preliminary to the main analysis.

48. SEMI-QUANTAL RESPONSES

Intermediate between quantal responses and the truly quantitative responses are those which may be described as *semi-quantal*. As noted earlier, Tattersfield *et al.* (1925) classified their insects as dead, moribund, slightly affected, and unaffected, and thus recognized four levels of response instead of the two characteristic of quantal data. They reduced their results to quantal form, however, by assessing toxicity in terms of the percentage of insects which were either moribund or dead, and made no allowance for the subclassification of these insects into two levels or of the remainder into the two levels of unaffected and slightly affected; standard methods of probit analysis are then applicable to the data without modification.

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Better use might be made of the data if the numbers of insects at the four different levels could be combined into a single index of toxic effect. Such a scheme was indeed proposed earlier by Fryer *et al.* (1923); they used the same fourfold classification and, in order to obtain a percentage response at each dose, scored moribund and slightly affected insects as one-half and one-quarter dead respectively. For example, if a batch of ten insects were classified as 4, 1, 3, 2, the proportionate response would be $(4 \times 1 + 1 \times 0.5 + 3 \times 0.25)/10$, or 52.5 %.

This method of scoring the results did not appear to give very much smoother response curves than that subsequently adopted by Tattersfield, and has never been very widely used. Possibly discrimination between moribund and slightly affected is simpler than between other pairs of classifications, so that the information provided by the latter is relatively unreliable. Exact statistical methods appropriate to the analysis of semi-quantal data have not yet been developed, but they would certainly be much more complex than the ordinary probit analysis. On the other hand, probably a sufficiently good approximation for many practical purposes is to form an index such as that described in \cdot the previous paragraph, and to use this as the value of p in an ordinary probit analysis, but always to use an empirical variance for expressing the precision of estimates. The weights derived from equation (3.4) or its generalization (6.3) should give a reasonable indication of the *relative* value to be placed on different observations, but can no longer be regarded as the reciprocals of the true variances; the heterogeneity χ^2 obtained therefore cannot be considered a true χ^2 , but is only a sum of squares from which an estimate of variance can be formed.

Chapter 11

1

RECENT DEVELOPMENTS

49. ALTEBNATIVES TO MAXIMUM LIKELIHOOD ESTIMATION

SINCE the first edition of this book was written, a number of alternatives to the scheme of maximum likelihood computations described in Chapter 4 have been developed, and the author's attention has been drawn to others of which he was previously unaware. Some of these undoubtedly have merits for routine analyses, because they reduce the time needed for calculation. Many, however, involve internal inconsistencies of assumption that will occasionally give trouble, although they may be harmless for most sets of data. The experienced statistician may use a rapid approximate method with safety, because he can recognize when some unusual feature of the data makes the method misleading or inapplicable. Unfortunately, the very simplicity of a method is an attraction to the experimenter who has little knowledge of statistical theory and who must therefore apply his chosen method uncritically. Moreover, at least under the assumption of a normal distribution of log tolerances and probably also for any other reasonable assumption about the form of this distribution, the approximate methods either do not make fully efficient use of the available data or require that the experiment be planned in a manner that does not make the best possible use of the materials. The labour of statistical analysis is usually only a small part of the total labour of an experiment, and, therefore, efficient analysis of a minimal number of observations is generally more economical than non-efficient analysis of a larger number. The argument is sometimes advanced that elaborate statistical analysis of a few observations is not worth while; this idea seems to be based upon a misconception, for scarcity of observations is likely to be a result of high cost of obtaining one observation, just the circumstances in which efficient utilization of each observation is economically most

important. In this book, chief emphasis is placed on efficient methods of analysis, but the reader who has mastered these should have little difficulty in understanding the simpler methods that, whatever their shortcomings, are certainly useful for the rapid preliminary evaluation of data.

Graphical methods due to Litchfield and Fertig (1941) and to de Beer (1945) have been mentioned in Chapters 3 and 4. The Litchfield and Fertig method has been modified by Miller and Tainter (1944), and again by Litchfield and Wilcoxon (1949); in its latest exposition, it could scarcely be said to look simple, but undoubtedly if it were used as a routine it would take less time than the orthodox probit method. All these methods employ a line drawn by eye to fit points plotted for each dose and response, preferably on logarithmic probability paper. Systems of nomographs are then used to simulate the maximum likelihood calculations for variances, fiducial limits, and contributions to a heterogeneity χ^2 ; even without the inevitable inaccuracies of reading from diagrams, the correspondence is not exact, because the fitting of the line cannot take proper account of the working probits and weights. For good data, however, with points close to a straight line and no responses near to zero or 100 %, the results obtained are likely to be very close to those from maximum likelihood estimation.

In §13, a simple arithmetical method has been attributed to Behrens. As there described, the method is in fact that of Reed and Muench (1938), who proposed to use linear interpolation between successive values of S_{x-} and S_{x+} to find the 50 % point. The method originally described by Behrens (1929), and independently but in less detail by Dragstedt and Lang (1928), is closely related but not identical; it makes use of the ratio

$$S_{x-}/(S_{x-}+S_{x+})$$

as a modified response for the dosage x, and interpolates between successive values of this ratio to give the 50 % point. These two methods often mislead users into a belief that they are not merely helping interpolation by a smoothing of the data, but that they are getting results of the same precision as if they had tested

 $(S_{x-}+S_{x+})$ subjects at each dose; Barr and Nelson (1949), for example, state explicitly 'This method enables one to obtain results in terms of a much larger number of animals than originally employed, and, therefore, results are indeed more accurate'. Though these methods may be satisfactory for estimating an LD 50 when the data are well distributed on either side of the 50 % point, they are never valid for the estimation of the LD 90, or any other percentage point; the misunderstanding just noted may have serious consequences if it is used in support of such an estimation. Winder (1947) has drawn attention to the dangers of regarding the Behrens ratio as an estimate of the response rate at any dose. Investigations now in progress indicate that the Reed-Muench form of calculations gives a smaller bias than the Dragstedt-Behrens; both, however, have slightly larger variances than that for Kärber's method, and, since they are no easier to use, they must be judged inferior to Kärber's method.

In §13 was noted a serious objection to the methods mentioned in the last paragraph, namely, that they are applicable only when a range of doses sufficient to span the interval from almost zero to almost 100 % response is used, and this feature is wasteful if an approximation to the LD 50 is known to the experimenter. The moving average method proposed by Thompson (1947) is related to Kärber's method, but is sufficiently different to escape this criticism. For data consisting of experimentally determined response rates, p, at equidistant points on the dosage scale, preferably but not necessarily each based upon the same number of subjects, Thompson would average all possible sets of three successive values of p and plot each average against the middle dose; linear interpolation between the two values nearest on either side of 50 % gives an estimate of the LD 50. (Movingaverage spans other than three can, of course, be used if preferred.) The estimate can be expressed by a formula similar to (3.12). Thompson claims for his method the advantage of estimation 'free from assumption as to the precise type of fundamental curve involved', though presumably symmetry of the log tolerance distribution is important. This method also must not be used for the estimation of any other percentage point. The

efficiency of estimation of the LD 50 has not been studied in detail, though Thompson presents a small amount of empirical evidence that it can stand comparison with the probit method.

The question may reasonably be asked: Why should the maximum likelihood estimation of the parameters be set up as a standard against which alternatives are measured? It must be admitted that no entirely satisfactory answer can be given. General statistical theory proves that, for any formulation of the tolerance distribution in terms of unknown parameters (e.g. that of equation $(2 \cdot 6)$), if the number of observations be large enough, no other method can give estimates of higher precision (smaller variance), though some may be as good. What constitutes 'large enough', however, is unknown. In some simple problems, the optimal property of maximum likelihood estimates persists even in very small samples; in the problem of relating a quantal response to dose, nothing is known about the distribution of the estimates in small samples. In many applications of the probit method, the number of observations is probably sufficient for the large-sample efficiency of maximum likelihood estimation to be nearly achieved, but at present there is no certainty of this. The force of the criticism largely disappears when it is realized that just as little is known about the efficiency of alternative methods of estimation in small samples, that most of these are known to be not fully efficient in large samples, and that they have many other faults, as mentioned above and in \S 13.

Berkson (1946) suggested estimation of the parameters in the tolerance distribution by minimizing χ^2 instead of maximizing the likelihood, and later (1949) developed this method more rigorously. It is known (Cramér, 1946, §30.3; Kendall, 1946, §18.8) to be as efficient as maximum likelihood in large samples; if the correct algebraic formulation of the tolerance distribution is being used, estimates from the two methods and their variances will tend to equality as the size of sample is increased, though in general they will differ for small samples. Minimum χ^2 might therefore seem to have the same status as maximum likelihood for the problems of this book. For samples of moderate size, however, it appears to have at least one disadvantage not shared

by maximum likelihood, namely the instability of χ^2 when some class numbers are small; the extreme instance is that of individual mortality records (§43), for which estimation by minimum χ^2 would seem an unpromising procedure. Nevertheless, there is no *a priori* reason why minimum χ^2 should not be superior to maximum likelihood in small samples, or why some third method should not be superior to either (Tukey, 1949), and investigation of this matter is clearly needed. For the normal log tolerance distribution, all that can at present be said is that minimum χ^2 has no known advantages, and therefore no tables to facilitate its use are provided in this book. For the logistic type of distribution, as Berkson has shown, minimum χ^2 fitting of the logit regression is rather simpler than maximum likelihood, at least if his ingenious approximation be adopted.

50. WADLEY'S PROBLEM

In a study of the effect of temperature on a bacterial suspension, counts of survivors per unit volume may be made, but the actual numbers exposed to treatment in the same units of volume are unknown. If the original number of bacteria per unit volume follows a Poisson distribution with mean N, and a treatment sufficient to kill a proportion P operates at random within a sample, the number of survivors per unit volume will follow a Poisson distribution with mean N(1-P). Analysis of data consisting of counts of survivors, s, at various temperatures would involve the estimation of N as well as of the parameters of the temperature tolerance distribution. A similar problem would arise in experiments on the control of immature stages of fruit flies, in which samples of fruit were treated at various dose levels of some factor, and counts were made of the numbers of flies which survived and developed. The total number of flies treated in any sample, however, could be discovered only by laborious counts on the dissected fruit. An alternative method of obtaining information on the numbers exposed to treatment is to take a parallel sample of untreated fruit, and to use the number of flies developing from this as an estimate of the numbers which would have developed in the other samples in the absence of treatment.

If the distribution of initial numbers were Poisson, the problem would be formally the same as for the bacteria, except that the control samples provide an estimate of N independent of any additional information accruing from comparisons between treated samples at different doses.

Wadley (1949) first stated this problem, and obtained the maximum likelihood solution. Finney (1949d) showed the close relationship to the problem of adjustment for natural mortality discussed in Chapter 6. From a provisional estimate of N, an empirical proportion killed at each dose,

$$p=1-\frac{s}{N},\qquad(11\cdot1)$$

may be assessed, and, on the assumption that once again the tolerance distribution is log normal, the usual probit calculations may be initiated. Working probits are still found as in §16, but, because a Poisson distribution occurs instead of a binomial, the weighting coefficient must be changed to

$$w = Z^2/Q; \tag{11.2}$$

this function is tabulated in Table VIII. Since N is the same for all doses, it may be introduced into totals at the end instead of by calculation of Nw for each dose. If the number of samples per dose varies, each w must be multiplied by the corresponding number. An auxiliary variate

$$x' = -Q/Z \tag{11.3}$$

is required, and may be read from Table II. Revised estimates of the three parameters are then obtained from

$$bS_{xx} + \frac{\delta N}{N} S_{xx'} = S_{xy},$$

$$bS_{xx'} + \frac{\delta N}{N} S_{x'x'} = S_{x'y'},$$

$$a = \bar{y} - b\bar{x} - \frac{\delta N}{N} \bar{x}',$$
(11.5)

and

equations exactly analogous to (6.5) and (6.6), though the change in sign in the definition of x' should be noted. If a sample is tested at zero dose, and gives s_0 survivors, so that s_0 is a direct estimate

of N, $S_{x'x'}$ must be increased by the provisional value of N and $S_{x'y'}$ by $(s_0 - N)$. The cycle of computation may be repeated as often as is required to give a close approach to the maximum likelihood estimates. A test of heterogeneity may be based on

$$\chi^2 = S_{yy} - bS_{xy} - \frac{\delta N}{N}S_{x'y}, \qquad (11.6)$$

in which S_{yy} is understood to have been increased by $(s_0 - N)^2/N$, and the variances and covariance of b, $\delta N/N$ are given by the elements of the inverse matrix of coefficients from equations (11.4). Other results, such as the LD 50 and its fiducial limits, are obtained in the familiar manner. Finney (1949*d*) has given details of a numerical example.

Anscombe (1949) has pointed out that, though a Poisson distribution for N may be appropriate to a well-stirred fluid in which bacteria are suspended, it is usually less satisfactory as a representation of an insect infestation in fruit. For the latter, a negative binomial distribution might be nearer to the truth; this distribution also has the property that counts for treated samples at any one dose will follow the same type of distribution with modified parameters. Unfortunately, however, N is now replaced by two parameters, both of which enter into the new weighting coefficient, and a satisfactory routine for statistical analysis has not yet been developed. Anscombe suggests a modification in experimental procedure that might help to $\bar{}$ overcome the difficulty.

51. A FACTORIAL EXPERIMENT

The merits of factorial design of experiments have been outlined in §29, and §30 indicated how standard methods of analysis might be adopted to a factorial experiment involving qualitative modifications of the stimulus. Potter and Gillham (1946) have reported several experiments of this kind, and Finney (1946) has described the analysis of one of these. A brief account of this analysis follows.

Ex. 34. The effect of temperature on the insecticidal potency of pyrethrins. Potter and Gillham (1946) tested the toxicity of

pyrethrins to adult Tribolium castaneum Hbst. under various conditions of storage before and after spraying. Full details are given in their paper. All that need be noted here is that the insects were stored in either a hot (H) or a cool (C) room before spraying, and in either a hot or a cool room after spraying; tests were made with each of the four possible combinations of storage treatments. Tests were also made, with each of these conditions, using a spray to which 1 % terpineol (T) had been added. The eight series of tests form what is called a 23 factorial system, there being three factors (storage of insects before spraying, storage after spraying, terpineol) each of which has two alternative states (hot or cool for the first two, absence or presence for the third). The eight series are identified by code symbols; thus C.H.T. refers to the tests in which terpineol was added to the spray for insects stored cool before, hot after spraying. In each series, about seventy insects were tested at each of three doses, 0.0195, 0.0130 and 0.0065 % total pyrethrins; the data have been given in full by Finney (1946).

 TABLE 37.1.
 Summary of Calculations for

 Estimation of Eight Regression Lines

Series	Ŧ	ÿ	Snw	S _{ss}	S ₂₉	S _{yy}
H.H.	1.057	4.966	76-0	2.7375	13-179	63.98
C.H.	1.059	5-370	103-9	3.7137	16.478	74.63
H.C.	0.953	6.055	63·3	2.0083	6.075	21.92
C.C.	0.908	6.074	63-6	1-6054	7.974	39.71
H.H.T.	1.148	4-407	92.7	2.2607	10-174	52.30
C.H.T.	1.088	5.003	115.3	3.9490	12.457	43.17
H.C.T.	0-918	6.062	47.4	1-3523	4.309	16-23
C.C.T.	0.864	6.692	24.2	0.3735	1.117	3 ∙58
Total				18.0004	71.763	315.52

(All doses were multiplied by 1000 in order to avoid negative values of x)

The procedure of §20 was used to give eight parallel probit regression lines. Table 37.1 summarizes the calculations. The heterogeneity of the deviations from the lines and the parallelism of the lines were examined, by forming S_{xy}^2/S_{xx} for each series separately and a corresponding expression from the totals and subtracting these from the S_{yy} and ΣS_{yy} . Table 37.2 was thus obtained just as was Table 13. The heterogeneity χ^2 , 18.8, is significant, but the mean square for parallelism is less than the mean square for heterogeneity and raises no doubts about the legitimacy of fitting parallel lines. The heterogeneity factor as ordinarily calculated is 2.35. Here, however, since the two mean squares in Table 37.2 are so similar, it is tempting to combine them in order to gain degrees of freedom, and to use

$$29 \cdot 42/15 = 1 \cdot 961$$

with 15 degrees of freedom, as the factor. Had the parallelism mean square been much larger than the residual, the basis of the analysis that follows would have been destroyed; had it been much smaller, pooling it with the residual would have introduced a danger of biasing the heterogeneity factor. The effect of pooling mean squares in circumstances such as these is a matter needing theoretical investigation, but it can scarcely do much harm, and has considerable advantages, when the constituent mean squares are as nearly equal as in Table 37.2.

 TABLE 37.2.
 Test of Heterogeneity and of Parallelism of Regressions

Nature of variation	D.F.	Sum of squares	Mean square
Parallelism of regressions Residual heterogeneity	7 8	10·61 18·81	1·52 2·35
Total	15	29.42	·

From Table 37.1, the common regression coefficient is estimated as $b = \Sigma S_{m}/\Sigma S_{m}$

$$= 71.763/18.0004$$

= 3.9867,

with the aid of which, and of the means \overline{x} , \overline{y} from Table 37.1, the log LD 50 for each series may be estimated. Moreover

$$g = \frac{1 \cdot 96 \times (2 \cdot 13)^2}{(3 \cdot 987)^2 \times 18 \cdot 00} = 0.031,$$

which is small enough to permit the use of variances.

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Chief interest attaches to comparisons between median lethal doses, rather than to the separate values, since in this manner the relative potencies of different conditions can be assessed. In particular the main effect of each factor must be studied: Is the potency for the average of all series stored hot before spraying different from that of all series stored cool before spraving?, with similar questions for the other two factors. Secondly, the interactions between these factors, the extent to which the effect of one is affected by the state of another, must be examined. A very convenient method of studying main effects and interactions in an experiment of this design (several factors, each at two levels) has been suggested by Yates (1937, §3), and may be adapted to the present purpose. A full account of the method will not be given here, but the main computations are shown in Table 37.3. The values of \bar{y} (Table 37.1) are arranged in order, starting with 'hot before, hot after, no terpineol' and changing each factor in turn according to the system shown in the first column. The first four entries of column (1) are the sums of the four successive pairs of values of \bar{y} , and the last four entries are the differences of these pairs, the first number always being subtracted from the second: thus

> $5 \cdot 370 + 4 \cdot 966 = 10 \cdot 336$, $5 \cdot 370 - 4 \cdot 966 = 0 \cdot 404$.

Column (2) is derived from column (1), and column (3) from column (2) by repetition of this process. A final column shows the result of similar operations on the values of \bar{x} .

Series	Ţ	(1)	(2)	(3)	(3) for \bar{x}
H.H.	4.966	10.336	22.465	44·629	7.995
C.H.	5.370	12.129	$22 \cdot 164$	1.649	- 0.157
H.C	6.055	9.410	0.423	5.137	- 0.709
C.C.	6.074	12.754	1.226	-0.321	-0.041
H.H.T.	4.407	0.404	1.793	-0.301	0.041
C.H.T.	5.003	0.019	3.344	0.803	-0.071
H.C.T.	6.062	0.596	-0.385	1.551	-0.199
C.C.T.	6.692	0.630	0.034	0.419	0.023

The first entry in column (3) is the total of the eight values of \bar{y} , and is of no particular interest. The second can be seen to be the difference between totals of the four 'cool before spraying' values of \bar{y} and the four 'hot before spraying', and is used in comparing the potency for these two conditions. In fact, for the average potency of cool relative to hot

$$M = -\frac{1}{4}(T_x - T_y/b), \qquad (11.7)$$

where T_x and T_y represent the second entries in column (3) for \bar{x} and \bar{y} respectively. The third entries, similarly, give the main effect of cool versus hot after spraying, and the fifth the main effect of the addition of terpineol. The remaining entries give interactions. The fourth, for example, contains the difference between 'cool versus hot before spraying, cool after' and 'cool versus hot before spraying, hot after', that is to say between

and (6.074 + 6.692) - (6.055 + 6.062) = 0.649(5.370 + 5.003) - (4.966 + 4.407) = 1.000

for the \bar{y} and between corresponding expressions for the \bar{x} ; application of equation (11.7) gives a value of M for the interaction of conditions before and after spraying. The sixth and seventh entries give interactions of the two storage conditions separately with terpineol, and the last entry gives the interaction of all three factors. Moreover, for any one of these Mvalues,

$$V(M) = \frac{1 \cdot 96}{16b^2} \left\{ \Sigma\left(\frac{1}{Snw}\right) + \frac{(4M + T_x)^2}{\Sigma S_{xx}} \right\};$$
(11.8)

this formula is analogous to equation (5.7), 1.96 being the heterogeneity factor; the factors of 4 and 16 arise because M is based on an average difference between two sets of four tests. The results of applying equations (11.7) and (11.8) are summarized in Table 37.4.

From Table 37.4, cool storage is seen to have increased the potency of pyrethrins substantially and significantly, especially after spraying. The mean effect of terpineol has been to reduce the potency slightly, but the difference is not significant. There is

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Effect	M	Relative potency
1. Cool versus hot before spraying	0.143 + 0.034	1.39
2. Cool versus hot after spraying	0.499 + 0.042	3.16
3. Terpineol	-0.029 + 0.032	0.94
4. Interaction of 1 and 2	-0.012 ± 0.032	0.97
5. Interaction of 1 and 3	0.068 + 0.033	1.17
6. Interaction of 2 and 3	0.147 + 0.033	1.40
7. Interaction of 1, 2 and 3	0.013 ± 0.032	1.03

TABLE 37.4. Summary of Effects Expressed as Relative Potencies

evidence, however, of positive interaction of the terpmeol effect with cool storage, the after-spraying interaction being significant; this means that, although it had no average effect on potency, terpineol seems to have augmented the benefits of cool storage. Addition of line 6 to line 2 in Table 37.4 gives the value of M for cool versus hot storage after spraying in the presence of terpineol, subtraction of line 2 from line 6 gives the value in the absence of terpineol:

> $0.499 + 0.147 = 0.646 = \log 4.43,$ $0.499 - 0.147 = 0.352 = \log 2.25;$

it is the difference of these two quantities whose significance is demonstrated by comparison of the sixth value of M in Table 37.4 with its standard error. The quantities could be obtained by direct calculation from Table 37.1, using either the first four or the last four lines only and calculating the appropriate T_x , T_y from these by the method of Table 37.3 or otherwise; equations (11.7), (11.8) must be modified in order to take account of the lesser number of series involved:

$$M = -\frac{1}{2}(T_x - T_y/b), \qquad (11.9)$$

$$V(M) = \frac{1 \cdot 96}{4b^2} \left\{ \Sigma' \left(\frac{1}{Snw} \right) + \frac{(2M + T_x)^2}{\Sigma S_{xx}} \right\}, \qquad (11\cdot 10)$$

where Σ' indicates summation over the appropriate four series only.

A convenient and easily understood summary for factorial experiments of this type is shown in Table 37.5. The relative potency for each effect is shown as an average for all series, and also separately averaged for the four series at each level of the other factors in turn. The derivation of the second pair of entries in the last column has been described above, and others follow the same pattern.

	l		Stor	rage			
Relative potency for	Mean	Before a	praying	After s	praying	- Terr	oineol
		Hot	Cool	Hot	Cool	Absent	Present
Cool instead of hot, before spraying	1.39	_		1.43	1.35	1.19	1.63
Cool instead of hot, after spraying	3.16	3.24	3.07			2.25	4.43
Terpineol added to spray	0.94	0.80	1.09	0.67	1.31		

TABLE 37.5. Relative Potencies of Pyrethrins under Various Conditions

52. INCOMPLETE REPLICATION IN COMPLEX EXPERIMENTS

The experiment discussed in §51 may be described as fully replicated, for tests were made at each dose level for all combinations of the factors of storage and terpineol. A similar design might be used in the application of litter-mate control to an experiment with quantal responses. If the test subjects are, say, rats, experience of other quantitative properties suggests that members of the same litter are likely to be less variable in their individual tolerances than members of different litters. An experimenter who proposed to test six dose levels (perhaps three from each of two preparations whose relative potency was to be estimated) might hope to increase the precision of his results by using litters of six rats and assigning to each dose one animal from each litter. On the assumption that the variance within a litter was the same for all litters, the method of §43 might be adapted to the estimation of a series of probit lines (two for each litter if two preparations were involved) constrained to be parallel, the slope being the reciprocal of the standard deviation

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of log tolerances within litters and therefore being steeper than if litter-mate control were not used. An average value of M could then be formed, as in equation (11.7), from the common value of b and the appropriate differences between averages of \bar{x} and of \bar{y} . The calculations would be more laborious than if the litter classifications were ignored and data for all subjects at one dose were pooled, but, for subjects showing real differences between litters, the precision of the relative potency would benefit. Moreover, it must not be forgotten that the design of an experiment always influences its analysis; to incorporate litter-mate restriotions into the design and to ignore them in the analysis would be improper and could be misleading.

Though the analysis suggested, like that used in §51, is a legitimate statistical treatment of the data, it is not the best possible. A litter whose members had unusually low tolerances would be regarded as giving information as valuable as that contributed by one whose tolerances fell in the middle of the dose range tested, yet clearly the observations from the first litter carry less weight. So in Ex. 34, the C.C.T. series showed very high response rates, and consequently a low value of Snw, yet in forming the relative potencies in Table 37.4 it was allowed to play the same part as the C.H.T. series. Of course, account is taken of this in forming V(M), but the unweighted mean of quantities whose variances are different will have a larger variance than a suitably weighted mean.

The difficulty arises in more acute form when there is no longer full replication. Suppose, for example, that tests for the six-dose experiment cannot all be completed on one day. They might be spread over six days in accordance with a Latin square design; thus, if 1, 2, 3, 1', 2', 3', denote the doses of the two preparations, the following scheme might be adopted:

Litter	Day l	Day 2	Day 3	Day 4	Day 5	Day 6
Ι	Ł	ľ	· 3′-	2'	3	2
II	2	1	ľ	3′	2′	3
111	ľ	2′	2	<u> </u>	1	3′
IV	3	2	2′	ľ	3′	1
V	2'	3′	3	1	2	ľ
VI	3′	3	L	2	ľ	21

Six litters would scarcely be enough to give reasonable precision, and the design might be extended by adding further sets of six litters with other Latin square arrangements. Again, if the response were not death but some minor reaction that could be repeated indefinitely without harm to the subject, each row might represent a single animal, instead of a litter, and five or ten tests per day could be made on every animal. The simple six-litter scheme will serve for illustration here. If a quantitative response were measured for each test, differences between litters and between days would be eliminated by an analysis of variance, and all comparisons between doses would be made with the same precision as if they were based entirely on differences within one litter and within one day. This ought still to be true for quantal responses, but the method of analysis is less obvious.

The key lies in the basic formulation of tolerance distributions. Each test gives information on the tolerance for a particular combination of litter and day, information that is inexact by comparison with a hypothetical direct measurement but still information of value. If direct measurements of log tolerance or some other quantitative character were made, the assumptions underlying the analysis of variance would be that the expected value for each combination of litter and day could be expressed as the sum of two parameters, one constant for the litter and the other constant for the day, and that the distribution of measurements about their expectations was normal with a variance constant for the whole experiment. Such assumptions are made regularly in the analysis of quantitative data, and an analysis for quantal data may equally well be based upon them; a complication arises, however, because the orthogonality of the Latin square disappears when unequally weighted probits replace quantitative measurements.

The analysis must begin with the choice of a set of expected probits for the thirty-six combinations of litter and day; these are the values read from provisional regression lines, each modified by addition of a quantity representing the deviation of its litter from the general average and another representing the deviation of its day from the general average. Thus, for a litter

which shows general evidence of low tolerances, all the expected probits will be increased by the same amount. In the first cycle of the iteration, these adjustments may be little better than guesses. Weights and working probits are found from the usual formulae, and weighted totals of y for each litter, each day, and each dose formed. By equation of these totals to similarly weighted totals of expected values, representing a true linear regression of probit on log dose modified by litter and day parameters, estimates of the parameters may be formed. This step is essentially the same as for the 'fitting of constants' in any non-orthogonal experimental design (Yates, 1933), except that the weights are no longer integral numbers of plots but values of nw. From the estimates of the parameters, a new set of expected probits is constructed, and iteration proceeds as in simpler problems. Even for a Latin square, the calculations are laborious. If angles could be used instead of probits (§15), the labour would be much less, since the constancy of w would preserve the orthogonality of the Latin square in each cycle of the calculations and the linear equations for the parameters would degenerate into the evaluation of arithmetic means. At least for experiments in which all expected probits lie between 3.2 and 6.8 (i.e. between 4 and 96 % probability of response), this is likely to be good enough. Even when angles are not satisfactory, one or two cycles of iteration based on angles might begin the calculations; expected angles would be replaced by the corresponding expected probits for the closing stages.

A Latin square of k^2 cells may be regarded as a fractional replicate of a k^3 factorial arrangement, since only 1/k of all combinations of the three factors are used (Finney, 1945b). Analysis is possible only if interactions between factors can be ignored. This condition, sometimes forgotten in other applications of Latin square designs, means here that tolerance differences between litters are assumed to remain the same from day to day; consequently, the fact that dose 2 is tested only on six of the thirty-six possible combinations of litter and day does not prevent the comparison of the responses it produces with corresponding results from other doses tested on other sets of six combinations.

The method of analysis just outlined, however, is essential if the risk of false inference from non-orthogonal comparisons is to be avoided. If all tests had been made simultaneously, so that the day classification did not occur, the same principles could be applied and would ensure that the final estimate of relative potency was the best weighted average from the several litters. The calculations would then become rather simpler, but the method of §51, now applicable because of the complete replication of all combinations of litters and doses, might be preferred by many experimenters, even at the price of loss of precision.

The general problem of the analysis of quantal response data in complex experiments may be regarded as solved, for the principles of analysis described here may be adapted to any design. Whether or not the gain in precision is sufficient to justify the labour of computation for a complex design must be decided by experience, as also must the extent to which approximations can replace the full scheme of analysis. No example of the computations will be given here, because, at the time of writing, they have been insufficiently tried for systematic recommendations to be made.

53. THEORY OF INDEPENDENT ACTION

Recent work by Plackett and Hewlett (1948) throws new light on the concept of the independent action of mixtures of poisons, and the account given in § 38 needs some modification. They point out the inadequacy of equation (8-20), and especially its inability to represent a negative correlation of susceptibilities. Instead, they turn to the basic distribution of dosage tolerances for the two poisons; just as for one poison the distribution of tolerances may often be taken as normal, so for two a bivariate normal distribution may be expected to give reasonable results. Equation (2.6) is thus generalized to

$$dP = \frac{1}{2\pi\sigma_1\sigma_2(1-\rho^2)^{\frac{1}{4}}} \exp\left[-\frac{1}{2(1-\rho^2)} \left\{\frac{(x_1-\mu_1)^2}{\sigma_1^2} - \frac{2\rho(x_1-\mu_1)(x_2-\mu_2)}{\sigma_1\sigma_2} + \frac{(x_2-\mu_2)^2}{\sigma_2^2}\right\}\right] dx_1 dx_2, \quad (11.11)$$

in which x_1 , x_2 are dosages of the two poisons, μ_1 , σ_1 and μ_2 , σ_2 the means and standard deviations of their own tolerance distributions; ρ is the correlation coefficient* between the two tolerances of a subject. When a subject receives a dose of a mixture, it will be killed if either constituent is present in excess of the tolerance and will survive if both doses are less than the corresponding tolerances.

A positive value of ρ means that most subjects with high tolerances for one poison will have high tolerances for the other. Consequently, a mixture may be little more potent than the stronger constituent applied alone. The extreme situation of $\rho = 1$, perfect correlation, gives a mortality for any mixture equal to that of the more toxic constituent alone, and is identical with the case of r = 1 in §38. Illustrations have been given in Figs. 12, 15, 16. If $\rho = 0$, the mortality from a mixture is exactly that expressed by equation (8.18), and adequately illustrated in Figs. 12-16. Intermediate values of ρ will give results similar to, but not identical with, those predicted by substituting ρ for r in equation (8.20). A negative value of ρ means that subjects whose tolerance for one preparation is high will generally have a low tolerance for the other; a mixture may be then much more potent than either constituent alone. Indeed, in the extreme case of $\rho = -1$, perfect negative correlation, equation (8.18) is changed to

 $\begin{array}{c} P = P_1 + P_2 \\ \text{or } 1 \end{array}$ whichever is the less. (11.12)

In other words, for low dozes of a mixture, the mortality will be equal to the sum of the mortalities appropriate to the two constituents applied separately; at or above the dose for which this sum is unity, the mortality will always be complete.

These authors have also investigated more fully the shape of the probit regression curve when $\rho = 0$, for a mixture of two constituents whose separate regression lines have slopes b_1 and b_2 . At low doses, the mixture will give results for which the probits lie practically on a straight line whose slope is the lesser of b_1 , b_2 ;

* The convention that the symbol ρ represents a relative potency must be suspended for this section.

at intermediate doses, the relationship will curve upwards; and at high doses it will again become approximately linear, with a slope of $(b_1^2 + b_2^2)^{\frac{1}{2}}$. The upper section of linearity, however, will generally be well outside the range of mortalities determinable experimentally. This is theoretical confirmation of the empirical results in § 38. For data of the limited range and number obtainable in most experiments, a straight line of slope

$$b = \frac{1}{2} [b_1 + (b_1^2 + b_2^2)^{\frac{1}{2}}] \quad (b_1 < b_2) \tag{11.13}$$

is likely to fit tolerably well, and very extensive data would be needed in order to detect the curvature.

These suggestions effect a great simplification in the theoretical formulation of a law of independent joint action. They may obviously be extended to any number of independently acting constituents. From the practical point of view, the importance of discovering insecticides whose tolerances are negatively correlated is obvious. In a later paper (Hewlett and Plackett, 1950), the same authors consider the estimation of ρ , illustrate the rather heavy calculations required for obtaining the estimate and its standard error, and obtain a test of significance for the deviations of observations from the predictions of correlated independent action. They also produce experimental evidence of a mixture of pyrethrins and D.D.T. behaving in accordance with the predictions in respect of its toxicity to *Tribolium castaneum*, the correlation coefficient being positive for application as a film, negative for a direct spray.

Similar action is the type of joint action that might reasonably be expected for a mixture of related chemical compounds all of which cause reaction or failure of the same physiological system, as in Exs. 20 and 23. Hewlett and Plackett contrast this with *dissimilar action*, in which the constituents of the mixture affect different physiological systems. Independent action is a particular case of dissimilar action. That two poisons whose physiological effects are entirely different happen to have approximately equal variances for their tolerance distributions, and therefore give probit regression lines that seem parallel, should not be regarded as justification for attempting to explain the

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toxicity of a mixture in terms of equations $(8\cdot4)$: in such circumstances, the basic assumption of similar action, that an amount of one poison can be replaced by a fixed proportion of the other without affecting the potency, is *a priori* unacceptable. Equation $(8\cdot23)$ is perhaps equally open to objection; the analysis of data on mixtures of rotenone and pyrethrins in Ex. 27 is an illustration of statistical method rather than the appropriate treatment for these data. A form of dissimilar action, with an interaction between the modes of action of the poisons or a correlation of individual tolerances, might well give predictions of potency for a mixture so like those for similar synergistic or antagonistic action as to be experimentally indistinguishable without very extensive tests.

These developments are at present too new to the author to be discussed adequately here. Hewlett and Plackett have opened up an important new line of investigation, by means of which the collaboration of experimenter and statistician may lead to a much more complete understanding of the toxic effects of mixtures than has been possible in the past.

54. ESTIMATION OF A PERCENTAGE POINT

If the prime object of a series of tests for quantal responses is to estimate the dose corresponding to a particular percentage response, the tests should be planned in order to estimate that dose as precisely as possible. The first problem facing an experimenter is then likely to be how to allocate his available supply of N subjects to different doses in order to maximize the precision of the estimate. No complete solution of the problem can be given, but some general principles can be established.

The easiest situation is that in which the ED 50 is to be estimated, and in practice the same recommendations may be regarded as applicable to neighbouring points such as the ED 45 or the ED 60. If nothing is known about the location of the ED 50 beyond a reasonable confidence that it lies between two widely separated limits, little can be done except to divide the Nsubjects equally between a large number of doses extending over a range rather wider than these limits. When N is large, a pilot

investigation with only a few of the subjects (say N/4 or N/10) might be used to give some idea of the ED 50, the remaining subjects being assigned to an experiment planned as described below. The experimenter who can afford only ten animals and wishes to estimate the mean toxic dose of a new poison or the mean curative dose of a new therapeutic agent, however, is asking for the impossible: discussion of whether he ought to use two groups of five animals or to give every animal a different dose is often vigorous but always unprofitable.

The precision with which the value of x corresponding to a 50 % response rate is estimated depends very much upon the position of the doses tested relative to the ED 50. Consequently. any information on the ED 50 can be used to advantage in the selection of a set of doses for the experiment; this knowledge will be inexact, for if it were good, no experiment would be needed, but the better it is the better will be the results of a carefully planned experiment. Inspection of equation (3.6) shows that, since b is a characteristic of the subjects rather than of the experiment performed upon them, the variance of m is largely determined by the size of Snw. The maximum value of w occurs at Y = 5, and therefore Snw will be maximized by choosing doses as near as possible to the ED 50. To concentrate all efforts on maximizing Snw would be disastrous, however, as tests confined to a single dose, or to several doses close together, would give a zero or small value for S_{xx} ; the precision of estimation of the regression coefficient would be low, and the second term in equation (3 6) large. If the doses are symmetrically spaced about the ED 50, the largest contributions to S_{rr} will come from subjects with Y = 3.42 or Y = 6.58; for increasing the precision of b, therefore, doses expected to give responses of about 6 or 94 % are the most valuable. Unfortunately, at these extremes, w is less than half its maximum value. A suitable compromise would be to divide the subjects equally between doses which previous information suggests as likely to give responses of about 15 and 85 %; contributions to Snw and to S_{xx} are then approximately 70 % of their maxima. Such an experiment would provide no test of the linearity of the regression (a heterogeneity

 χ^2 could be obtained by dividing the subjects into several batches at each dose: §42), and division of subjects between three doses, planned to give 10, 50, 90 % responses, or four doses, planned to give 10, 35, 65, 90 % responses, is often preferable. Unequal division of N between the doses appears to give no advantages that cannot more conveniently be gained by choice of dose levels.

In this argument, equation (3.6) has been assumed applicable, and g small. The smaller the number of subjects, the less precise will be the estimate of b for any one of the schemes just recommended. When N is small enough to bring the value of q above 0.1, there is a case for using doses nearer to the 6 and 94 % level, or even of abandoning the equal division of N, in order to keep g as low as possible and to prevent it from unduly influencing the precision of m; formula (4.6) will have to be used for the fiducial limits of m when g is as large as this. Table 37.6 shows values of Ng for 5 % fiducial limits, values of $Nb^2V(m)$, and values of the standard error of m, for various schemes of two, three, and four dose experimentation. The variance is calculated from equation (3.6), with the supposition that $m = \bar{x}$, as will be true for the ideal situation in which the doses chosen by the experimenter produce exactly the response rates he had predicted; deviations from this ideal will in general increase both q and the variance. Inverse ratios of $Nb^2V(m)$ measure the relative efficiencies of the different schemes for the estimation of the log ED 50, but they must be used with caution as they are applicable only when N is large enough to make g negligible. The column of values of Ng shows that even a two-dose experiment at the 6 and 94 % levels must use over sixty subjects in order to have g small enough to be neglected, and that the formula for V(m) can seldom be used for calculations of fiducial limits unless over 100 subjects are used; if attempts are made to reduce V(m) by the choice of doses near to the ED 50, even 400 subjects may be too few for this. When g is large (say greater than 0.1), variances calculated from equation (3.6) may give misleading evidence on precision, and comparisons between different experimental designs must be made in terms of the widths of fiducial ranges at an agreed probability level (Finney, 1947a).

TABLE 37.6. Values of g and Measures of Precision, in Various Schemes of Experimentation for the Estimation of an ED 50 using a Total of N Subjects

Subjects divided equally between doses giving % responses of	Ng (5%)	$Nb^2V(m)$	S.E. (<i>m</i>)
6. 94	6.3	3.95	1·99 <i>σ</i> /√N
10, 90	6.8	2.92	$1 \cdot 7 1 \sigma / \sqrt{N}$
15, 85	8.4	2.35	$1.53\sigma/\sqrt{N}$
20, 80	11-1	2.04	1·43σ/√N
25, 75	15.9	1.85	$1.36\sigma/\sqrt{N}$
35, 65	41.9	1.66	$1 \cdot 29\sigma / \sqrt{N}$
6, 50, 94	9.5	2.63	1·62σ √N
10, 50, 90	10.3	2.27	$1.51\sigma/\sqrt{N}$
15, 50, 85	12.5	2.02	$1.42\sigma/\sqrt{N}$
20, 50, 80	16.7	1.85	$1.36\sigma/\sqrt{N}$
25, 50, 75	23.8	1.75	$1.32\sigma/\sqrt{N}$
35, 50, 65	62-9	1.63	$1.28\sigma/\sqrt{N}$
6, 35, 65, 94	11.0	2.34	$1.53\sigma/\sqrt{N}$
10, 35, 65, 90	11.8	2.12	$1.46\sigma/\sqrt{N}$
15, 35, 65, 85	13.9	1.95	$1 \cdot 40\sigma / \sqrt{N}$
20, 35, 65, 80	17.6	1.83	$1.35\sigma/\sqrt{N}$
25, 35, 65, 75	23.0	1.75	$1.32\sigma/\sqrt{N}$

(Full explanation in text; σ is variance of tolerance distribution)

If the doses could be chosen on the basis of previous information so as to ensure that \bar{x} was very nearly equal to m, the second term in equation (3.6) would be negligible, and Table 37.6 would show that, for large N, the standard error of m decreases steadily as the doses are concentrated near to the ED 50. Under these conditions, equation (4.6) shows that the effect of g is equivalent to a division of V(m) by (1-g). Hence, as judged by narrowness of the 5% fiducial range, if only 100 subjects are to be used, the fifth of the two-dose schemes, the fourth of the three-dose or four-dose will be the best; if only twenty subjects are to be used. the third of the two-dose schemes and the second of the threedose or four-dose will be the best. This argument gives some idea how a decrease in N pushes the optimal doses further away from the ED 50 on either side. The impossibility of choosing the doses so symmetrically that \bar{x} is equal to m emphasizes this trend, since S_{xx} must be kept moderately large in order to prevent the second term in equation (3.6) from making too great a contribution to V(m).

Though these conclusions have been based upon an assumption of a normal distribution of log tolerances, the general behaviour of such alternative distributions as are likely to be considered for practical use (\S 15) is so similar to the normal, at least in the central region of the distribution, that the same recommendations should prove adequate in a general qualitative sense. A similar investigation could easily be made for any form of distribution that seemed to be of particular interest.

The estimation of an extreme percentage point is more difficult. but is undoubtedly important. The user of an insecticide, for example, is likely to be more interested in the LD 99 than in the LD 50, and in other fields of study the ED 5 may be important. The precision with which the ED 99 can be estimated with the aid of N subjects will be much less than the precision for the ED 50; if tests are made at doses in the neighbourhood of the ED 99, the weights of the observations will be low, yet if tests are made in the central region the second term in equation (3.6)will make a large contribution to the variance. The second procedure, indeed, is scarcely worthy of serious consideration because it will involve extrapolation beyond the range of the observations, an action to be avoided unless the regression relationship fitted to the data is known to be absolutely, or very nearly, correct in its algebraic form. Though the normal distribution is usually adequate to explain the dose-response relationship for a considerable range on either side of the 50 % response, there is rarely any certainty that it is correct, and, if the true probit regression equation has a slight curvature, extrapolatory estimation of a dose such as the ED 99 might be grossly misleading.

Bartlett (1946) drew attention to the importance of estimating an extreme percentage point from observations made in its neighbourhood, and advocated an inverse sampling procedure. His method of obtaining maximum likelihood estimates is presented here in a slightly more general form. It may be adopted for any investigation in which the subjects are tested one at a

time and each can be classed as responding or not responding before the next is tested. As described here, the method is useful for the estimation of a high percentage point, but the same procedure will apply to a low percentage point if the roles of 'response' and 'non-response' are interchanged.

Suppose that subjects are tested one by one at a dosage x, which is expected to give a high response rate, until s have failed to respond, at which stage a total of n has been tested. In this inverse sampling, n is a variable for fixed s, instead of the more usual direct sampling situation of s being a variable for fixed n. An experiment will involve tests of several dosages, with observations of n for each; s need not be a constant for all doses. The general theory stated in Appendix II leads to a probit regression line, by means of an iterative process of the usual type. An empirical response rate

$$p = 1 - q = 1 - \frac{s}{n}$$
 (11.14)

is used to give empirical probits, a provisional regression line, weighting coefficients

$$w = Z^2/PQ^2, \tag{11.15}$$

and working probits

$$y = Y - \frac{Q}{Z} + \frac{Q^2}{Z} \left(\frac{1}{q}\right) \tag{11.16}$$

are determined, and the weighted regression of y on x, with sw (not nw) as the weight per observation, calculated. Table IX shows the minimum working probit,

$$y_0 = Y - \frac{Q}{Z} \tag{11.17}$$

the range, Q^2/Z , and w, from which the working probits and weights for an analysis may rapidly be constructed; the table has entries only for Y = 5.0 and upwards, as lower values of the expected probit are unlikely to occur. After completion of enough cycles of the iteration, the final regression equation,

$$Y = a + bx,$$

is used for the estimation of the required percentage point, by

substitution of the appropriate value for Y, and fiducial limits are given by equation (4.6).

Bartlett suggested that, for the estimation of an extreme percentage point, the inverse sampling method, using a series of doses towards the appropriate tail of the tolerance distribution, would have advantages over direct sampling. He made a numerical study of variances, analogous to that for the ED 50 in Table 37.6, and in some instances found a smaller variance for the inverse method. His investigation, however, seems to have been scarcely fair to direct sampling, for which he considered only experiments with doses symmetrically distributed about the ED 50; an experimenter who wished to estimate the ED 90 would in any event avoid low doses and concentrate on the high ones. In fact, if a second experiment were performed, this time by the ordinary direct method, with the same dosages and with values of n chosen to be equal to those observed in the inverse experiment, the precision of an estimated percentage point would be about the same; this follows because sZ^2/PQ^2 and nZ^2/PQ are approximately equal, and indeed will tend to equality in large samples. Apart from differences due to the small samples used in practice, the two methods would be equivalent.

The real argument for the inverse method is based not on precision but on relevance. If an estimate of an ED 90 is to be of practical use, it must have been estimated from observations on doses that bracket it, and, moreover, the weight of information from doses above the ED 90 should be comparable in amount to that from lower doses; only so can the evils of extrapolation be avoided. Without foreknowledge of the dose-response relationship, satisfactory allocation of the N available subjects between a series of doses is impossible. If instead a series of doses almost certain to bracket the ED 90 is chosen, and tests made by the inverse method, a reasonable balance of information from the different doses will be maintained; the standard error of the rate of non-response, q, will bear an almost constant ratio to q for all doses that give a high response rate (Finney, 1949a). The scheme of experimentation might be to make tests at each dose in turn until one subject has failed to respond, then with the remaining

subjects to make further tests at each dose until s = 2, then to increase until s = 3, and so on until the N subjects have been used. If the last subject responds, and so does not complete a group of tests for one dose, little harm will result from treating all the subjects for that dose as though they related to direct sampling, and using the ordinary formulae for w and y. Even this procedure is not ideal, for Z^2/PQ^2 increases steadily with P and an excessive amount of the total information in the observations may therefore come from the very high doses. This is perhaps a good fault, as it will compensate for the natural tendency of the experimenter to use doses below rather than above the ED 90. The formal precision of the estimated ED 90 may be less than could be obtained in some other way; a more equal division of subjects between the same doses, for example, might give a much greater total weight to the observations, and, even though the increase in $(m - \bar{x})$ made the second term in equation (3.6) larger, the result might be a reduction in V(m). The requirement of relevance, however, must be regarded as more important than formal precision, and even the scheme last mentioned might involve an undesirable extrapolation. In the recommended scheme, the regression line may not be the same as would be obtained from experimentation in the neighbourhood of the ED 50, but it will be an approximation determined from, and relevant to, observations round the percentage point to be studied. The ill effects of using an assumed tolerance distribution which may differ from the true will have been minimized, in a general rather than a rigorous mathematical sense, though deviations from strict normality are likely to be more important for the estimation of an extreme percentage point than for the ED 50.

For many quantal response investigations, of course, the inverse approach would be impracticable. Estimation of the LD 99 of an insecticide by spraying insects one at a time is not likely to prove a popular method, though a modification using small batches of ten or twenty at once might sometimes be worth considering. When the inverse method is impracticable, direct sampling must be used, with values of n guessed as well as possible. In other circumstances, however, as for example in

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the application of destructive tests to certain industrial products, subjects are normally tested singly, and the inverse method could be adopted as easily as the direct.

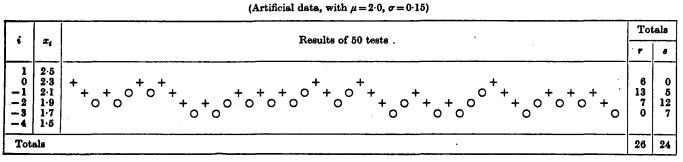
55. STAIRCASE ESTIMATION

The inverse sampling method, described in §54, is useful only if the experimental technique involves testing subjects one at a time, in such a way that the result of any test is known before the next is begun. If, in addition, a change in the level of dose is so little trouble to the experimenter that he is prepared to make every test at a dose different from that of its predecessor, another scheme of experimentation deserves consideration. This *staircase method* is so called because the dose for any test is one step above or one step below the dose for the preceding test.

A series of dosages is chosen, say ..., $x_{-3}, x_{-2}, x_{-1}, x_0, x_1, x_2, x_3, ...,$ where x_0 is believed to be about the ED 50 and the values of x are equally spaced. The result of any test determines the dose for the next test: if the subject responds, the next subject is tested at a dose one step lower, if it fails to respond, the next is tested at a dose one step higher. The first test is made at x_0 ; later tests will tend to be concentrated about the ED 50, because the rule ensures that the further from the ED 50 any test dose happens to be the greater is the probability that the next dose is one step nearer. The staircase method is clearly related to the sequential sampling methods developed in other branches of applied statistics. Dixon and Mood (1948) have given a preliminary report and have proposed an approximate method of statistical analysis which is good enough under certain conditions. The arithmetic of this is very simple, and can best be explained with the aid of an example; no probit transformation is used.

The data in Table 37.7 were obtained artificially, with the aid of a table of random numbers, and purport to relate to a staircase experiment on fifty subjects. The data were in fact derived by sampling from a distribution of tolerances which is normal on the *x*-scale and has true values $\mu = 2.0$, $\sigma = 0.15$ for its parameters. The step between successive doses, to be represented by *d*, is 0.2. The estimation proposed by Dixon and Mood involves first

TABLE 37.7. Example of the Staircase Method





STAIRCASE ESTIMATION

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forming r, s, the total numbers of responses and non-responses at each dose, then N_r , N_s , the totals of these columns, and subsequently working only with the smaller of these two. The next step is to calculate either

or
$$\begin{array}{c} A_r = S(ri), \\ A_s = S(si), \end{array}$$
 (11.18)

and then to estimate the ED 50 (on the dosage scale represented by x) as

$$m = x_0 + d\left(\frac{A_r}{N_r} - \frac{1}{2}\right),$$

$$m = x_0 + d\left(\frac{A_s}{N_s} + \frac{1}{2}\right).$$
(11.19)

OF

Thus, in Table 37.7, N_s is the smaller,

 $\begin{aligned} A_s &= -5 - 2 \times 12 - 3 \times 7 = -50 \\ m &= 2 \cdot 3 - 0 \cdot 2 \times \left(\frac{50}{24} - \frac{1}{2}\right) \\ &= 1 \cdot 983. \end{aligned}$

and

Study of equations (11.19) indicates a similarity between this procedure and Kärber's method (§13).

Dixon and Mood obtain an estimate of σ , the standard deviation of the tolerance distribution, as

$$\sigma_e = 1.620d \left(\frac{NB - A^2}{N^2} + 0.029 \right), \qquad (11.20)$$

where

$$\begin{array}{l} B_r = S(ri^2) \\ B_s = S(si^2) \end{array}$$

$$(11.21)$$

or

is to be substituted in equation (11·20), according as the response or non-response column is to be used. It is noteworthy that, though the right-hand side of equation (11·20) is a linear function of $(NB-A^2)$, it estimates the standard deviation and not the variance of the distribution. Dixon and Mood showed this to be a valid approximation when $(NB-A^2)$ exceeds $0.3N^2$; when $(NB-A^2)$ is less than $0.3N^2$, the estimation of μ remains valid but that of σ breaks down. For the data of Table 37.7

$$B_{s} = 116,$$

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and therefore

$$\sigma_e = 1.620 \times 0.2 \times \left(\frac{24 \times 116 - 2500}{576} + 0.029\right)$$

= 0.169,

there being no trouble about the applicability of equation (11.20). The two estimates, m and σ_e , agree well with the known true values in the population sampled.

The standard error of m is dependent not only on σ but on the ratio d/σ . Dixon and Mood investigated the relationship, and constructed a diagram from which the standard error can be obtained; in the range that is of practical importance, a sufficiently close approximation to their diagram is given by

S.E.
$$(m) = \sigma \left(0.9 + \frac{d}{8\sigma} \right) / N^{\frac{1}{2}}$$
 for $0.2 \leq \frac{d}{\sigma} \leq 2.4$, (11.22)

in which N_r or N_s is to be inserted for N. Another diagram shows the standard error of the estimated standard deviation; this depends upon d/σ in a more complicated manner which may be expressed very roughly as

s.E.
$$(\sigma_e) \simeq \sigma \left(1 \cdot 6 - \frac{d}{6\sigma} \right) / N^{\frac{1}{2}}$$
 for $0 \cdot 6 \leq \frac{d}{\sigma} \leq 2 \cdot 0.$ (11.23)

Outside the limits specified, the standard error of σ_e may be much greater. Equations (11.22) and (11.23) cannot be used directly, since σ is unknown, and the effect of substituting σ_e for σ has not been examined. For the sake of comparison with subsequent calculations, it may be noted that, for the data of Table 37.7,

$$\frac{d}{\sigma_e}=1.18;$$

if σ_e be substituted for σ in equations (11.22) and (11.23), the results are

S.E.
$$(m) = 0.169 \times 1.05 \div \sqrt{24}$$

= 0.036,
S.E. $(\sigma_e) = 0.169 \times 1.40 \div \sqrt{24}$
= 0.048.

and

The first result suggests that limits of about 1.91 and 2.05 might be set to m, the second that limits of about 0.07 and 0.27 might be set to σ , and the breadth of the range for σ clearly throws doubt on the validity of the inference for μ .

The calculations just described may be valuable as a method of rapid estimation from staircase data: they are obviously far from ideal for purposes of exact inference. Dixon and Mood's paper contained also an account of a maximum likelihood procedure, for use when equation (11.20) is not applicable or when extreme values of d/σ make their approximate formulae untrustworthy. This procedure begins with a neglect of a small part of the information, in order to simplify the analysis, but even then the computations are more laborious than for an ordinary probit regression. Yet inspection of the scheme of experimentation shows no reason why the routine of Chapter 4 should not be adopted. At each value of x tested, except for the two extremes, the data give (r+s) independent observations, leading to rresponses, and provide an unbiased estimate of the response rate; the fact that the occasions on which these doses have been used are determined by responses at adjacent values of x does not affect the independence of the observations. The highest and lowest values of x used for tests have the property that, because of the staircase rule, one must show only responses and the other must show only non-responses. This, however, is a small-sample peculiarity, which becomes progressively less important as the total number of tests is increased and the chance occurrence of long sequences of responses or of non-responses pushes the extreme values of x further from the neighbourhood of the ED 50; moreover, the maximum and minimum working probits make some allowance for these extreme responses. As admitted in §49. maximum likelihood estimation has some theoretical flaws when applied to small samples, and there appears to be no reason for supposing that the form of calculation proposed by Dixon and Mood is free from any of the criticisms that may be levelled at the probit method (to which, indeed, it is very closely related). The probit method may be applied to staircase data as reasonably as to any of the other types of data discussed in this book; the

simple approximation described at the beginning of this section may then be regarded as similar to the approximate methods of estimation described in § 13, in that it has some advantages but is of limited applicability.

Application of probit calculations to the totals in the r and s columns of Table 37.7 yields a regression equation

$$Y - 5 = 6 \cdot 094(x - 1 \cdot 980).$$

Thus the ED 50 is estimated as

$$m=1.980$$

and the standard deviation of the tolerance distribution as

$$1/b = 0.164.$$

These are very close to the results of the approximate analysis. If a standard error for m is calculated by equation (3.6) it is found to be 0.034, practically the same as was given by the approximate method. This cannot properly be used, however, because

and consequently

$$b = 6.09 \pm 1.57$$

$$g = 0.254.$$

Formula (4.6) may be used to give 5 % fiducial limits 1.900, 2.056 to the value of m; as noted in § 54, this allowance for g has almost the same effect as a division of the approximate standard error, 0.034, by $(1-g)^{\dagger}$. The almost perfect agreement between these limits and those assigned earlier on the basis of Dixon and Mood's approximate method is no doubt fortuitous, but is gratifying evidence that the approximate method is at least moderately trustworthy. Fiducial limits to σ may be obtained as the reciprocals of fiducial limits to b, and are 0.11 and 0.33; the interval between these is about the same as that between the limits based upon equation (11.23), but both values are a little higher.

As with Bartlett's inverse sampling method for the estimation of an extreme percentage point (§ 54), the merit of the staircase method is not that it is in any inherent quality more precise, but that its rule of experimentation concentrates the observations

where they are most valuable. In the example discussed, the estimate of ED 50 formed is more precise than if the fifty subjects had been allocated in almost equal numbers to the four doses 1.7, 1.9, 2.1, 2.3. Moreover, if nothing had been known originally, except perhaps that the ED 50 almost certainly lay between 1.5 and 2.5, an even wider range of doses, and consequently still lower precision, might have resulted from an attempt to apply the principles suggested at the beginning of § 54. In some situations, the staircase method is the ideal solution to the problem of the pilot investigation, for the first few tests play the part of the pilot in indicating roughly where the ED 50 lies, yet all observations can be combined in the final analysis. If the original uncertainty about the ED 50 is great, there is no reason why a start should not be made with a large dose interval, d, which may be reduced to d/2 or d/4 as soon as the ED 50 has been roughly located; the probit method of analysis applies without modification, and no doubt Dixon and Mood's approximate method, with the obvious slight modifications, would continue to serve well.

Further research on the simple staircase scheme outlined here, and on many variants of the rule for successive doses, is believed to be in progress. For example, a rule that a step down is taken only after two consecutive responses at a dose, whereas a nonresponse is always followed by a step up would concentrate observations at some point above the ED 50. Rules might be found which would give highly efficient allocations of subjects for estimating the ED 90 or the ED 10. Such a modification as that just suggested would not invalidate the probit method, though this need not be true for all staircase schemes, and approximate estimation formulae may also be developed. The account given here is no more than an introduction to a new field.

56. THE MEANING OF STANDARD ERRORS

A warning needs to be given against the uncritical acceptance of standard errors, fiducial limits, or other measures of precision, as though they indicated the variation that might be encountered in *any* repetition of an experiment. Because of their emphasis on

analytical technique rather than on interpretation, text-books of statistics tend to leave the reader with the impression that an estimate has only one standard error; the explanation of fiducial limits in 12 may be open to a similar misunderstanding. In reality, even though only one standard error may be assessable from a particular set of data, the estimate will have different standard errors appropriate to the different kinds of comparison in which it may be used.

The temptation to regard a percentage effective dose estimated from one sample of subjects, whether animal or human, as a 'normal value' typical of the species, must not be bolstered up with argument from a standard error based on the internal variation of the sample. If any impression has been given that, for example, the value of 4.85 ± 0.25 mg./l., obtained in Ex. 6 for the LD 50 of rotenone to *Macrosiphoniella sanborni*, is an estimate of a 'normal value' for this species of insect, that impression must be corrected; it is necessarily only an indication of what would be found in repeated tests of the same stock of *M. sanborni* under the same conditions of testing.

This very limited interpretation must always be attached to statements about the LD 50 or ED 50 of a single material. Tests performed in a London laboratory may be used to estimate the LD 50 and its standard error, and the results may be valid for future work in that laboratory; they could not be expected to give guidance on the outcome of similar tests in New York, where necessarily the subjects would be obtained from a different source, would be genetically different, and would have different nutritional and environmental experience. There is no more reason to suppose that their tolerances would be entirely independent of these conditions than that their weights or other measurements of size would be independent of them; small differences in experimental technique between different laboratories may also affect results. A standard error calculated from the internal evidence of one experiment relates only to repetition under the same conditions, using either the same subjects and materials or others that can be regarded as random samples from the same population as were those in the original experiment. The LD 50 may have one

standard error between determinations by one worker, another standard error between workers using the same apparatus and stock of animals in one laboratory, and yet another between determinations in different laboratories. Comparison of estimates obtained from several experiments by one worker, or in one laboratory, will assess only the variations to be expected from day to day under those conditions and will still not be related to variations between laboratories or between different stocks of subjects. However extensive the experimentation on one population, no statistical analysis can demonstrate the applicability of the conclusions to a different population.

At first sight, this argument might appear to deny the usefulness not merely of statistical analysis but of experimentation. The extent to which past experience can form a logical basis for future action is too great an issue for discussion here. Much of the present difficulty seems to be removed by adoption of comparative experiments. As Anscombe (1948) has pointed out, even though the absolute performance of a particular treatment varies erratically with changes in experimental conditions, the relative performance of two treatments may be much less disturbed by these changes. The relative toxicity of two chemically related insecticides, for example, at least for a particular species of insect, may be a characteristic of the materials, rather than of the circumstances of any one experiment. There is some evidence of this, but the truth-if truth it be-is an empirical one, and is not derived from statistical theory. A full study of the problem would involve lengthy experimentation into the effect of environment on the responses of insects to insecticides. In so far as these experiments demonstrated consistent differences, they could form a reasonable basis for inference relating to the future, while their inexplicable variations would warn the experimenter against any too general application of absolute standards.

In one class of experiment to which the methods of this book are relevant, a theoretical reason exists for believing a numerical result to be applicable more widely than in the circumstances of the particular experiment that produced it. If two preparations contain the same effective constituent, but this is diluted to

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different extents by constituents that do not affect the response, an estimation of relative potency constitutes an *analytical dilution assay*; the subjects are used purely as an aid to the measurement of the relative amounts of the effective constituent in the two preparations, and, though the choice of subjects and experimental conditions will influence the precision, it should not limit the meaning of the estimate. The result is an estimate of **a** property which, in theory, is independent of the species of subject or other circumstances of its determination; empirical verification of this statement is still desirable, but should be undertaken as a check on the validity of fundamental assumptions about the nature of the preparations rather than as a study of other sources of variation.

In spite of the title of § 1, this book is an exposition of statistical methods associated with the probit transformation and not an account of the theory of biological assay. The logic underlying the last paragraph has been discussed at length by Jerne and Wood (1949); statistical techniques needed in many assay problems have been described by Burn, Finney and Goodwin (1950) and Emmens (1948).

Appendix I

THE COMPUTING OF PROBIT ANALYSES

THE chief hindrances to the more widespread adoption of the probit method for the statistical analysis of quantal response data are probably the apparent complexity of the mathematical theory and the apparent laboriousness of the computations. The more technical details of the theory (Appendix II) are admittedly difficult, especially for the many biologists who lack mathematical training. In Chapters 2 and 3 an attempt has been made to present a reasonably simple account of the aim and underlying principles of the method, in a form which the reader will be able to appreciate even though he has to take the theory on faith. For many routine purposes the computations can be largely replaced by the graphical analysis described in Chapter 3, relative potencies and other parameters being estimated by measurement on the diagrams. When the complexity of the data, or the desirability of using extensions of the probit method (such as have been described in later chapters), makes necessary arithmetical rather than graphical estimation, the labour can be much reduced and computational accuracy much increased by orderly arrangement and systematic working procedure. In this appendix is given an example of the computing required for the simplest type of analysis, the fitting of a single probit regression line; the arrangement is that used throughout this book, but the steps are set out in greater detail and recommendations for adequate checking are made. With modifications appropriate to the various circumstances, the same procedure may be used in computing any of the more complicated probit regression equations that have been described.

The computer is assumed to be working with a calculating machine; the same results can be obtained by pen-and-paper calculations, but almost any machine intended for general computing will materially improve both accuracy and speed. Before buying a machine it is well to take expert advice on the most suitable type, though to some extent personal preference must decide between the products of different manufacturers. If the machine is to be used frequently, an electrically operated model is desirable, and a 10-figure rather than an 8-figure keyboard is an advantage that usually outweighs the additional cost.

The reader should find little difficulty in becoming proficient at the arithmetical processes described in this book, but the only route to accuracy and efficiency is by way of experience. A machine is not a complete safeguard against arithmetical errors, and carelessness will lead to wrong answers just as certainly as in non-mechanized calculations. One of the most frequent sources of error lies in copying from paper to machine or from machine to paper; inversions of the order of two digits and similar mistakes are particularly easily made. Computations should therefore be planned so as to reduce to a minimum the necessity for copying on to paper figures that have later to be restored to the machine. The surest means of preventing errors is to have all work checked by another computer on another machine, but, by the application of full checks at every stage, one computer should be able to carry out the work satisfactorily. The chief danger is that any misreading of figures, wrong setting of the machine, or faulty working of the machine may be repeated in checking; in order to guard against this, where a section of the computations cannot be checked by an independent path and the steps have therefore to be duplicated, the order of setting the machine should be changed so as to avoid the complete repetition of every detail. For example, a column may be summed by starting from the bottom instead of from the top, or a product found by interchanging the roles of multiplicand and multiplier.

Inexperienced computers frequently carry a far greater number of digits in their computations than is warranted by the accuracy of the original data, and present their results to six places of decimals when three at most are justifiable. In machine calculations an increase in the number of digits does not increase the labour to the same extent as in pen-and-paper work, and undoubtedly it is sometimes easier to carry an additional decimal place than to decide how many can be justified. Nevertheless the saving of time through working with fewer digits may be considerable in a long series of analyses; it should further be remembered that copying errors are less common with four digit numbers than with seven. The general practice in the numerical examples of this book has been to cut out all unnecessary digits from the early stages of an analysis (i.e. in dosages, working probits, weights, etc.), to retain digits fairly fully at intermediate stages (sums of squares and products, elements of matrices, etc.) as an aid to checking, but to present results shorn of superfluous digits and free of any spurious appearance of accuracy. Unless doses have been measured to within 0.1 % of their true values, two or three significant digits in the dosage, x, are sufficient. Percentage kills

based on batches of 200 individuals or less need not be expressed more precisely than to the nearest 1%, thus enabling the tables of this book to be used without interpolation; there is little to be gained by calculating the kills to greater accuracy than the nearest 0.1% unless the batches contain more than 2000 individuals.

Martin (1940, Table 3) has given data on the toxicity to Aphis rumicis of an ether extract of Derris malaccensis; the original observations of the experiment have been reconstructed from his figures in order that they may be used in an example of the typical probit analysis. Table 38 shows the greater portion of the computations for estimating a linear regression equation which will relate the probit mortality to the log concentration.

 TABLE 38. Computations for the Fitting of a Probit

 Regression Equation

λ	z	5	7	p'	<i>p</i> (<i>C</i> =2)	Em- pirical probit	¥	n 10	y	nwI	nwy
520 390 260 130 65 0	2·72 2·59 2·41 2·11 1·81	49 45 44 52 47 48	49 44 38 15 4 1	100 98 86 29 9 2	100 98 86 28 7 0	^{co} 7-05 6-08 4-42 3-52 	7·5 6·9 6·1 4·7 3·3	2·4 6·8 17·4 30·4 6·7 	7.85 7.03 6.08 4.43 3.57	6·528 17·612 41·934 64·144 12·127 	18·840 47·804 105·792 134·672 23·919

 $1/Snw = 0.0156986, \ \bar{x} = 2.2346, \ \bar{y} = 5.1967.$

Snwx ²	Snwxy	Snwy ²
321.72589	757-4672	1809-159
318-08633	739.7180	1720-234
3.63956	17.7492	88.925
		86-558

The various steps in building up Table 38 and in completing the estimation of the probit regression line are as follows:

1. In the column headed λ enter, in suitable units (here milligrams of dry root per litre of spray fluid), the doses tested, arranging these in descending order from the highest to the controls or zero concentration.

2. In the column headed x enter the logarithms of λ , to base 10, correct to two places of decimals. The doses may be multiplied or divided by a power of 10 throughout in order to make x take small

positive values, but a compensating adjustment must be made in later stages of the analysis if the results are to be expressed in the original units of dose.

3. In the columns headed n and r enter for each dose the number of insects tested and the number badly affected, moribund, or dead (Martin's B+M+D, which he takes as the 'kill').

4. Check that steps 1, 2 and 3 are correct, beginning each check from the bottom of the column.

5. Calculate the percentage kill, p' = 100r/n, to the nearest whole number; this step is most expeditiously performed on a slide rule. If *n* exceeds 200 for many of the doses, give the percentages to one decimal place.

6. The percentage kill among the controls is small (c = 2), and is estimated from about as many insects as the kill for other doses; the approximate analysis discussed in § 27 will therefore be sufficiently accurate. The population value, C, is taken as equal to c, and the adjusted kills calculated accordingly as

$$p = \frac{p'-2}{98} \times 100$$

= 1.020p'-2.0:

p also is taken correct to the nearest whole number (or to 1 decimal place if n is generally greater than 200). A slide rule can be used for the multiplication.

7. Check step 5, multiplying p' by n to give 100r.

8. Check step 6 from the relation

$$p' = 0.98p + 2.$$

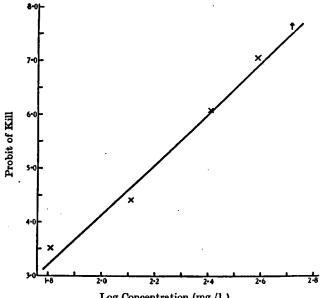
9. Enter the probits of p in the 'empirical probit' column, reading the values from Table I or Table 1, to 2 decimal places.

10. Plot the empirical probits against x, as shown in Fig. 23; draw a provisional straight line to fit the points, placing the line by eye and allowing for one point above the line at x = 2.72.

11. For each of the dosages used in the experiment read the value of the ordinate to the provisional line. These are the expected probits, Y, and are entered in the appropriate column of Table 38, correct to 1 decimal place; greater accuracy in Y is unnecessary unless n is very large.

12. Check steps 9, 10 and 11.

13. From the column C = 2 of Table II, read the weighting coefficient for each Y in Table 38, multiply by the corresponding n, and enter, to I place of decimals, in the column nw. Two or three significant digits in nw will generally be sufficient, and a good working rule is to use 2 places of decimals when n is less than 20, 1 place when n is between 20 and 200, and the nearest whole number for higher values



Log Concentration (mg./1.)

FIG. 23. Relationship between dosage of D. malaccensis and probit of kill of A. rumicis, showing probit regression line.

of n, modifying this rule so as to give the same number of decimal places for each dose throughout one analysis. In exceptional circumstances, such as the occurrence of a very wide discrepancy between the provisional line and an observation at either extreme of the range, a greater number of decimal places in nw may be desirable for the one dose. These recommendations assume that C is not very large; more decimal places may be required to give a reasonable number of significant digits when a high value of C reduces the values of wconsiderably.

14. From Table IV enter the working probit, y, corresponding to each p (not p') and Y. Thus from the last page of this table, the last

line of the column headed 7.5 gives y = 7.85 for p = 100; from the preceding page the column headed 6.9 gives y = 7.03 for p = 98, and so on. Use 2 decimal places if *n* is generally less than 200, otherwise 3. If Y is less than 2.0 or greater than 7.9, determine y from Table III as described in § 16.

15. Check steps 13 and 14; 13 may be checked satisfactorily with a slide rule. It will be observed that y usually, but not always, differs less from the empirical probit than does Y.

16. Place the first value of nw on the keyboard of the machine, multiply by the corresponding x, and enter the product in the column nwx; leave the keyboard unaltered but clear the product from the machine, multiply by y, and enter in the column nwy. Clear the machine and repeat for each dosage, entering each result to its full number of decimal places. No entries are made in the line for $\lambda = 0$, as the controls are used only for estimating C.

17. Place the last value of x on the right-hand side of the keyboard, unity at the left-hand side, and multiply by nw; without clearing the result, repeat with the next to the last value of x and so work to the top of the x column, thus accumulating the products at the right-hand side of the result register and the sum of nw at the left hand. Enter the totals

$$Snw = 63.7,$$

$$Snwx = 142.345,$$

in Table 38, and check the latter by addition of the *nwx* column from the top. Before *Snwx* is cleared from the machine, divide it by *Snw* to give \bar{x} to 4 places of decimals, and enter \bar{x} in Table 38.

18. Repeat these operations using y in place of x, thus checking the total Snw and also obtaining

$$Snwy = 331.027;$$

again check, by adding the *nwy* column from the top, and before clearing divide by 63.7 to give \bar{y} to 4 places of decimals. Enter \bar{y} in Table 38.

19. Find the reciprocal of Snw, either from tables or by division, and enter at the bottom of Table 38 to at least 7 decimal places. Additional accuracy is needed here, as this quantity is to be multiplied by large numbers.

20. Set 1/Snw on the keyboard and check that multiplication by Snw gives unity; clear the result and multiply by Snwx, so checking the values of \bar{x} ; clear the result and similarly check \bar{y} .

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21. Square Snwx and divide by Snw, multiply Snwx and Snwy and divide by Snw, square Snwy and divide by Snw; the answers 318.08633, 739.7180 and 1720.234, are to be entered in Table 38 as shown. Check by squaring the second, dividing by the first, and obtaining the third as the quotient; the first being still on the keyboard, multiply by 63.7 and divide by 142.345 to give 142.345 as a quotient. These checks may fail in the last digit as a result of rounding off. The parts played in the checks by the first and third of the three calculated quantities should be interchanged when the third has the greater number of significant digits.

22. Set the first entry for nwx on the keyboard, multiply by x, and repeat with successive lines of Table 38, accumulating the total so as to give

$$Snwx^{2} = 6.528 \times 2.72 + 17.612 \times 2.59 + \dots + 12.127 \times 1.81$$

= 321.72589.

Enter this figure in the appropriate position in the lower part of the table. Multiply the last value in the nwx column, which is already on the keyboard, by y and repeat with successive entries up the table, to obtain

$$Snwxy = 757.4672;$$

enter this figure. Multiply nwy by x down the table, in order to check Snwxy. Before clearing the machine, subtract from Snwxy the second of the three quantities calculated in step 21, without the machine, to give $S_{ry} = 17.7492;$

enter in Table 38 and chèck the difference by means of the machine.
Multiply *nwy* by
$$y up$$
 the table to give $Snwy^2$, and enter in Table 38.

23. Check $Snwx^2$ by accumulating products of nwx and x up the table. Before clearing, subtract the first of the quantities calculated in step 21, without the machine, to give

$$S_{xx} = 3.63956;$$

enter in the table and check with the machine. Similarly check $Snwy^2$ down the table, derive $S_{uv} = 88.925$,

enter in the table, and heck.

24. Compute $b = 17.7492 \div 3.63956$ = 4.8767.

25. Obtain the equation for the probit regression line as

$$Y = 5 \cdot 1967 + 4 \cdot 8767(x - 2 \cdot 2346)$$

= -5 \cdot 70 + 4 \cdot 88x;

3 places of decimals are almost always sufficient for the coefficients of this equation, and consideration of the standard errors to which the parameters are subject often suggests, as here, that only 2 are justified.

'26. Compute $(17.7492)^2 \div 3.63956$, and subtract the result from S_{yy} , giving $\chi^2_{131} = 2.37$;

reference to Table VI shows this not to be indicative of significant heterogeneity, and variances may therefore be derived from true weights without any heterogeneity factor. When a significant χ^2 is obtained at this stage, modify subsequent steps as in Exs. 7 and 9.

27. To check b, set 3.63956 and multiply by 4.8767 to obtain 17.7492.

28. The variance of b is $1/S_{xx}$. Find the standard error of b as the square root of this reciprocal, and check by calculating the reciprocal of $\sqrt{S_{xx}}$; a slide rule or tables of square roots and reciprocals may be used. The result is b = 4.88 + 0.52.

and the size of the standard error shows that there is no need to quote **b** to more than 2 decimal places.

29. Check step 25. Calculate values of Y for three values of x (say x = 1.5, 2.0 and 2.5), and plot Y against x on Fig. 23. The three points should be collinear and they define the probit regression line. This line is almost indistinguishable from the provisional line and has therefore not been drawn separately in the figure. Since the agreement is so good, there is no need to carry out a second cycle of computations.

30. Find the log LD 50 as the value of x which gives Y = 5:

$$m = (5+5.70)/4.88$$

= 2.193.

The antilogarithm of m is the estimated LD50, 156 mg./l.

31. Calculate

$$V(m) = \frac{1}{(4.877)^2} \left\{ 0.0157 + \frac{(2.193 - 2.235)^2}{3.640} \right\}$$

= (0.0157 + 0.0005)/23.79
= 0.00068,

whence the standard error of m is ± 0.026 .

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32. Calculate $g = t^2 V(b)/b^2$ for the probability level at which fiducial limits are required; for the 5% level t = 1.96 (Table VII), so that

$$g = (1.96)^2/3.640 \times 23.79$$

= 0.044.

33. Since g is small, the fiducial limits to m may be taken at a distance 1.96×0.026 on either side of m, these therefore being 2.244 and 2.142. The corresponding concentrations are 175 and 139 mg.71. respectively.*

34. Check steps 30-33.

35. The conclusion from the analysis of these data is that the median lethal dose of Martin's ether extract of *Derris malaccensis* to *Aphis rumicis*, under the conditions of the experiment, has a maximum likelihood estimate of 156 mg./l.; with a fiducial probability of 95 %, the true value of the median lethal dose may be expected to lie between 175 and 139 mg./l.

The above is a detailed account of a systematic arrangement of the computations suitable for use when a single computer is responsible for the whole analysis. The plan need not be followed exactly and no doubt personal taste will suggest modifications. There is necessarily a conflict between the desirability of checking a result before too many further calculations have been based upon it and the desirability of delaying a check for some time so as to reduce the risk of unconscious repetition of mistakes. The computer's aim should be to carry out the original calculations correctly, and checking should be looked upon as a verification of correctness rather than as means of discovering errors. Where a result can be checked independently of its first calculation the check may be made immediately, but where the only check is to repeat the same processes a reasonable interval should be allowed to elapse first. In the arrangement of the computations that has just been described, notes on checks have been inserted, but generally the reader has been left to devise his own methods for these; they should not be made earlier than the points indicated, and preferably the original working should be so accurate that they can safely be left until much later. When a second computer is available to assist in the analysis, he should be responsible for all checks.

* Fiducial limits to m calculated from the exact equation (4.6) are 2.243 and 2.139, and the concentrations are 175 and 138 mg./l. respectively, thus confirming the statement that g is too small for the exact formula to be needed.

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The reader who adopts either this or a similar arrangement of the work for the simple type of data should have no difficulty in adapting and extending it to the more complex analyses discussed in Chapters 5–10. For example, when relative potencies are being estimated for several poisons tested in a single experiment, the provisional lines (step 10) will be drawn parallel if there seems any possibility of data agreeing satisfactorily with the hypothesis that the probit regression lines have equal slopes. Thereafter the computations proceed as outlined above, for each poison separately, until the values of S_{xx} , S_{xy} and S_{yy} have been obtained for each. A test of parallelism and, if departures from parallelism are not significant, subsequent estimation of relative potencies based on a common regression coefficient follow as described in § 20. The computations are analogous to those in steps 24–34 above.

Appendix II

MATHEMATICAL BASIS OF THE PROBIT METHOD

MANY who are concerned with the use of the probit transformation in the analysis of numerical data will be content to accept the technique without explanation of the mathematical and statistical theory. In Chapters 1-11 of this book, the practical applications have been developed, and the computational processes illustrated, with a minimum of theoretical argument. For the benefit of the mathematically minded reader, this appendix gives a brief outline of the derivation of the equations of estimation which have been used elsewhere. The appendix is not intended as a complete account, but the whole theory of the probit method can be developed on the lines set out below; the mathematical statistician should have little difficulty in discovering for himself how the equations that follow lead to the computations described earlier, or in modifying these equations in order to obtain the analogous techniques for the alternative distributions of tolerance discussed in § 15. The essential features of the maximum likelihood estimation of the parameters in the quantal response problem were contained in a note by Fisher (1935), though the first adequate formal presentation appears to have been that of Garwood (1941); Finney (1949d) has given a more general result. As emphasized in §49, the known optimal properties of maximum likelihood estimation relate to large samples, and some alternative may be superior in samples of finite size. No other method has yet been demonstrated to have an advantage, and only maximum likelihood will be considered here. Berkson's method of minimizing χ^2 (1949) is in many respects similar, and the construction of the iterative process may be of interest in connexion with problems other than those of quantal responses.

Suppose that a tolerance distribution is represented by equation (2-1), so that the probability of response to a dose λ_0 is *P*, as defined by equation (2-2). If a batch of *n* subjects is exposed to the stimulus at a dose λ_0 , and if the subjects react independently of one another, the probability of *r* responses is given by the binomial distribution (§7) as

$$P(r) = \frac{n!}{r!(n-r)!} P'Q^{n-r}.$$
 (II, 1)

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Suppose that a series of k doses is tested in an experiment; then the probability of a particular set of numbers killed in each group is proportional to e^{L} , where

$$L = Sr \log P + S(n-r) \log Q, \qquad (II,2)$$

and S denotes summation over all doses. The quantity e^L , or, more strictly, a quantity proportional to it but having a maximum value of unity, has been called by Fisher (1922) the *Likelihood* of the observations.

Now P, Q are functions of the dose which contain certain parameters, and the problem confronting the statistician is that of estimating the parameters from the experimental data. Fisher (1922, 1925) has shown that estimates of the parameters which maximize the likelihood are *asymptotically efficient* (Cramér, 1946, §33·3; Kendall, 1946, §17·27). The likelihood is a maximum when L is a maximum; hence, if θ is a parameter of the distribution of individual tolerances, the maximum likelihood estimate of θ must satisfy the equation

$$0 = \frac{\partial L}{\partial \theta} = S \frac{r}{P} \frac{\partial P}{\partial \theta} + S \frac{n-r}{Q} \frac{\partial Q}{\partial \theta}$$
$$= S \frac{n(p-P)}{PQ} \frac{\partial P}{\partial \theta}, \qquad (II,3)$$

where

$$p = r/n \tag{II, 4}$$

is an empirical estimate of the value of P at a particular dose. If the tolerance distribution involves more than one unknown parameter, a set of equations of the form of (II, 3) must be satisfied simultaneously.

Direct solution of such equations is seldom possible, but iterative methods can be made to give successive approximations converging to the solutions. It will be sufficient to illustrate the procedure for two parameters, θ and ϕ . Suppose that θ_1 , ϕ_1 are first approximations to the solutions of the equations (II, 3), obtained perhaps by rough graphical or arithmetical estimation. By the Taylor-Maclaurin expansion, to the first order of small quantities

$$\frac{\partial L}{\partial \theta_1} + \delta \theta \frac{\partial^2 L}{\partial \theta_1^2} + \delta \phi \frac{\partial^2 L}{\partial \theta_1 \partial \phi_1} = 0, \\ \frac{\partial L}{\partial \phi_1} + \delta \theta \frac{\partial^2 L}{\partial \theta_1 \partial \phi_1} + \delta \phi \frac{\partial^2 L}{\partial \phi_1^2} = 0,$$
 (II, 5)

where the addition of the suffix to θ , ϕ indicates that the first approximations are to be substituted after differentiation. The second-order

differential coefficients $\frac{\partial^2 L}{\partial \theta_1^2}$, $\frac{\partial^2 L}{\partial \theta_1 \partial \phi_1}$ and $\frac{\partial^2 L}{\partial \phi_1^2}$ may be simplified by putting p = P after differentiation, in order to give expected instead of empirical values (cf. Fisher, 1944, §57.2). Equations (II, 5) then become

$$\delta\theta S \frac{n}{P_1 Q_1} \left(\frac{\partial P_1}{\partial \theta_1}\right)^2 + \delta\phi S \frac{n}{P_1 Q_1} \left(\frac{\partial P_1}{\partial \theta_1}\frac{\partial P_1}{\partial \phi_1}\right) = S \frac{n(p-P_1)}{P_1 Q_1} \frac{\partial P_1}{\partial \theta_1},$$

$$\delta\theta S \frac{n}{P_1 Q_1} \left(\frac{\partial P_1}{\partial \theta_1}\frac{\partial P_1}{\partial \phi_1}\right) + \delta\phi S \frac{n}{P_1 Q_1} \left(\frac{\partial P_1}{\partial \phi_1}\right)^2 = S \frac{n(p-P_1)}{P_1 Q_1} \frac{\partial P_1}{\partial \phi_1}.$$
 (II, 6)

The solutions, $\delta\theta$, $\delta\phi$, are adjustments to θ_1 , ϕ_1 , which give improved approximations, $\theta_2 = \theta_1 + \delta\theta$, $\phi_2 = \phi_1 + \delta\phi$. Further adjustments are obtainable by recalculating the equations with θ_2 , ϕ_2 in place of θ_1 , ϕ_1 , and the process may be repeated until the latest set of adjustments is negligible. General theory of statistical estimation, indeed, states that if the first approximations are of non-zero efficiency, the first cycle of computation will yield fully efficient estimates. This, however, is strictly true only for large samples, and also is likely to be disturbed by the subjective element in determining θ_1 , ϕ_1 from the graphical process usual in probit analysis. The wisest course, therefore, is to iterate the calculations until the solutions of (II, 3) are approached; care and experience in the choice of first approximations will usually ensure that two cycles give a numerical accuracy sufficient for practical purposes.

If $\hat{\theta}$ is the estimate of a single parameter, obtained as a solution of the maximum likelihood equation (II, 3), its variance is (Cramér, 1946, § 33·3; Fisher, 1922; Kendall, 1946, § 17·25)

$$V(\hat{\theta}) = -1 / \frac{\partial^2 L}{\partial \hat{\theta}^2}, \qquad (\text{II}, 7)$$

in which expression $\hat{\theta}$ is to be substituted for θ after differentiation. When more than one parameter must be estimated, the variances and covariances of the estimates are the elements of the inverse matrix of second differential coefficients; thus, for two parameters,

$$V = \begin{pmatrix} -\frac{\partial^2 L}{\partial \hat{\theta}^2} & -\frac{\partial^2 L}{\partial \hat{\theta} \partial \hat{\phi}} \\ -\frac{\partial^2 L}{\partial \hat{\theta} \partial \hat{\phi}} & -\frac{\partial^2 L}{\partial \hat{\phi}^2} \end{pmatrix}^{-1}$$
(II, 8)

This matrix, but with the previous set of approximations instead of $\hat{\theta}$, $\hat{\phi}$, is inverted as part of the last cycle of computations, since the

differential coefficients appear as the coefficients of $\delta\theta$, $\delta\phi$ in equations (II, 6). In practice, the variances and covariances may be taken as approximately equal to the elements of this inverse, so as to avoid the necessity of recalculation. A χ^2 test for the heterogeneity of the departures of the observations from the tolerance distribution specified by the parameters then follows.

The equations discussed above are of general applicability, whatever the form of P, and may easily be extended to the estimation of a greater number of parameters. Toxicity test data, and other quantal response problems, require their simplification to specialized forms suitable for computation. The most important form is that for the estimation of the parameters of the tolerance distribution given by equation (2.6), or

$$P = \frac{1}{\sigma \sqrt{(2\pi)}} \int_{-\infty}^{x} \exp\left[-\frac{1}{2\sigma^2} (x-\mu)^2\right] dx, \qquad (\text{II}, 9)$$

where x measures the dosage on a logarithmic or other normalizing scale. This distribution has been shown in §9 to be equivalent to a linear relationship between x and the probit of P, say

$$Y = \alpha + \beta x, \qquad (II, 10)$$

in which the new parameters, α , β are related to the old μ , σ by

$$\mu = (5 - \alpha)/\beta,$$

$$\sigma = 1/\beta,$$
(II, 11)

and Y is defined by equation (3.1), namely

$$P = \frac{1}{\sqrt{(2\pi)}} \int_{-\infty}^{Y-5} e^{-\frac{1}{2}u^2} du.$$
$$\frac{\partial P}{\partial Y} = \frac{1}{\sqrt{(2\pi)}} e^{-\frac{1}{2}(Y-5)^2} = Z,$$

Now

the ordinate to the normal curve at the point whose abscissa is (Y-5), and therefore

$$\frac{\partial P}{\partial \alpha} = Z, \quad \frac{\partial P}{\partial \beta} = xZ.$$
$$Y = a_1 + b_1 x \qquad (II, 12)$$

Hence, if

is a first approximation to the maximum likelihood estimate of

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equation (II, 10), adjustments to a_1 and b_1 are given by equations (II, 6) or

$$\delta aS \frac{nZ^{2}}{PQ} + \delta bS \frac{nZ^{2}}{PQ} x = S \frac{nZ^{2}}{PQ} \left(\frac{p-P}{Z}\right),$$

$$\delta aS \frac{nZ^{2}}{PQ} x + \delta bS \frac{nZ^{2}}{PQ} x^{2} = S \frac{nZ^{2}}{PQ} x \left(\frac{p-P}{Z}\right),$$
(II, 13)

in which Z, P, Q are determined from equation (II, 12). If the weighting coefficient, w, is defined by

$$w = Z^2/PQ, \qquad (II, 14)$$

these last equations are seen to be equations for the estimation of the weighted linear regression of (p-P)/Z on x, the weight nw being assigned to each value of (p-P)/Z. A convenient method of solution is to introduce the working probit

$$y = Y + \frac{p - P}{Z}, \qquad (\text{II}, 15)$$

and to add SnwY to each side of the first, SnwxY to each side of the second of equations (II, 13). The weighted linear regression equation of y on x is then calculated in the form

$$Y = \bar{y} + b(x - \bar{x}), \qquad (\text{II}, 16)$$

where

$$\bar{x} = Snwx/Snw, \bar{y} = Snwy/Snw,$$
 (II, 17)

(II, 18)

and

 $b = S_{xy}/S_{xx};$ equation (II, 16) may be seen to be a revised version of (II, 12), with

$$a_2 = a_1 + \delta a = \bar{y} - b\bar{x},$$

$$b_2 = b_1 + \delta b = b.$$

The iteration may continue with a new cycle of computations based on

$$Y = a_2 + b_2 x_1$$

By equation (II, 8), which here is analogous to the ordinary formulae for weighted linear regression,

$$V(\bar{y}) = 1/Snw,$$
 (II, 19)

$$V(t) = 1/S_{xx},\tag{II,20}$$

provided that the maximum likelihood estimates have been approached sufficiently closely. These results are, strictly, applicable only for

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large samples, and research into the appropriate formulae for small samples is needed; the rapidity with which increase in the number of observations improves the approximation of a normal distribution to a binomial suggests that, when the total number of subjects tested is not very small, these variance formulae will be trustworthy.

Homogeneity of the experimental materials and mutual independence of the subjects have so far been assumed. If the n subjects in a batch do not react independently to the dose applied, but depart from the fitted equation (II, 16) more markedly than can be attributed to random variation, without showing any regularity such as would suggest the need for a different specification of the tolerance distribution, the weights of the observations must be decreased, and the variances correspondingly increased. This situation will in general be indicated by a significantly large value of the heterogeneity χ^2 , whose calculation is described in §§ 17, 18. Provided that no classes have had to be amalgamated (cf. Ex.7), χ^{2} will have (k-2) degrees of freedom, two less than the number of batches tested; $\chi^2/(k-2)$ is then the factor by which the variances must be increased. In a more complex analysis, the heterogeneity factor remains the mean square derived from the heterogeneity χ^2 . When a heterogeneity factor has to be used, all variances and standard errors must be considered in relation to a t-distribution, not a normal, with degrees of freedom equal to those of χ^2 . This procedure is not peculiar to probit analysis but is the normal statistical practice of using an empirical variance instead of a theoretical when the data give evidence of the occurrence of variation other than that included in the theoretical value.

The problem of adjustment for a natural response rate (§ 28) which supplements the responses due to the stimulus provides an instructive example of the generalization of the method just described (Finney, 1949 b, d). When an average natural response rate, C, operates independently of the stimulus, the total response rate expected at any dose is as given in equation (6.1),

$$P'=C+P(1-C).$$

The probabilities P', Q' replace P, Q in (II, 1), and the prototype of the maximum likelihood equations is

$$0 = \frac{\partial L}{\partial \theta} = S \frac{n(p' - P')}{P'Q'} \frac{\partial P'}{\partial \theta}.$$
$$\frac{\partial P'}{\partial C} = Q, \quad \frac{\partial P'}{\partial P} = (1 - C),$$
$$p' - P' = (p - P)(1 - C).$$

Now

and also.

In general, C also has to be estimated. If S is used for summation over all non-zero doses, and the n_c subjects tested at zero dose showed a response rate c, the equations corresponding to (II, 3) may be written

$$\frac{\partial L}{\partial \alpha} = S \frac{nZ^2(1-C)}{Q\{C+P(1-C)\}} \left(\frac{p-P}{Z}\right) = 0,$$
$$\frac{\partial L}{\partial \beta} = S \frac{nZ^2x(1-C)}{Q\{C+P(1-C)\}} \left(\frac{p-P}{Z}\right) = 0,$$
$$\frac{\partial L}{\partial C} = \frac{n_c(c-C)}{C(1-C)} + S \frac{n(p-P)}{C+P(1-C)} = 0.$$

Differentiate again, and equate observed values with expected; if

$$w = \frac{Z^{2}}{Q\left\{P + \frac{C}{1 - C}\right\}}$$
$$\therefore x' = Q/Z$$

and

be introduced, as in equations (6.3), (6.4), the coefficients required for equations (II, 5) may be written

$$\begin{aligned} \frac{\partial^2 L}{\partial \alpha^2} &= -Snw, \\ \frac{\partial^2 L}{\partial \alpha \partial \beta} &= -Snwx, \\ \frac{\partial^2 L}{\partial \beta^2} &= -Snwx^2, \\ \frac{\partial^2 L}{\partial \alpha \partial C} &= -\frac{1}{1-C}Snwx', \\ \frac{\partial^2 L}{\partial \beta \partial C} &= -\frac{1}{1-C}Snwx', \\ \frac{\partial^2 L}{\partial \beta^2 C} &= -\frac{1}{1-C}Snwxx', \\ \frac{\partial^2 L}{\partial C^2} &= -\frac{n_c}{C(1-C)} - \frac{1}{(1-C)^2}Snwx'^2. \end{aligned}$$

If the working probit is again introduced, the equations for a, b, δC may be reduced to the form most suitable for computing, equations (6.5), (6.6); these equations are formally the same as for a multiple linear regression of y on x, x', and the analysis may proceed as illustrated in Ex. 17.

Estimates of the parameters for other expressions of the probability function, P, can be derived in a similar manner. Other examples in

which the basic distribution (II, 9) is retained but modified have been given in §§ 45, 47, 50, and the reader should have no difficulty in developing the equations required for these problems. The analogue of the probit method appropriate to any of the alternative tolerance distributions mentioned in § 15 can be found by alteration of (II, 9) to an integral based on the new distribution. This has been illustrated elsewhere (Finney, 1947*a*) for the logistic distribution, for which it leads to the maximum likelihood estimates in Berkson's (1944) logit method. The logistic sigmoid might be more appropriate than the normal for the problem of Ex. 33; it could be estimated without any change in the form of equations (10·1)-(10·6), except that P, Q, Z, w, x', y, must be interpreted in terms of the logistic function:

$$P = e^{Y}/(1+e^{Y}),$$
$$Z = e^{Y}/(1+e^{Y})^{2},$$

with the other quantities related to P, Z by the usual formulae. Thus the method of §47 can be used in the fitting of a logistic curve with unknown upper limit to observations with equal variances, and can easily be modified so as to take account of unequal variances. Since the logistic and normal sigmoids are so similar in shape over a wide range of values of P, a logistic analysis of the data of Ex. 33 would not in fact have given results appreciably different from those of the probit analysis.

It must not be forgotten that the probit transformation is only a convenient trick for representing the sigmoid (II, 9) by a straight line, and that probit analysis is only one convenient method for the solution of equations such as (II, 3) for the normal distribution of tolerances. Neither the transformation nor the method of analysis have any fundamental relationship to the problems in which they are used, beyond what is implied by this statement. The chief advantage of probit analysis over other methods of solving the same equations is that it enables the computational methods familiar in linear regression analysis to be applied in what is not really a linear regression problem. The wide range of tables now available reduces to a minimum the need for the calculation of special functions in any one problem; the procedure is a simple, easily checked routine that can be performed by workers who would be at a loss if merely ordered to solve equation (II, 3).

Nevertheless, methods other than those of this book are in some respects superior. Black (1950) suggests an alternative method of tabulating maximum and minimum working probits which, while

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leading to what is mathematically the same analysis, may be more rapid in use than the usual, since the nwy column is built up directly. The gain is at the expense of the graphical representation, and the linear regression analogy is left a little less clear, but, at least for some types of data, it has advantages. Black's proposal, in fact, is equivalent to assigning a weight and a score to each subject, the score being a maximum or minimum function of the expected Y according as the subject does or does not respond: the analogy between probit analysis and linear scoring methods used in the estimation of genetic parameters (Finney, 1949c) is thus apparent. Garwood (1941) compared the use of expected values of the second differential coefficients in equations (II, 5) with the use of empirical values calculated from the complete expressions for these differential coefficients. The two methods must converge to the same estimates of the parameters, though not necessarily to identical values for the variances. Garwood found that the empirical coefficients gave a quicker convergence, in the sense that fewer cycles were needed in order to attain a specified closeness of approach to the limiting values; this itself is an empirical conclusion, based on trial of several sets of data, though it is likely to be generally true. The advantage of the empirical coefficients is counterbalanced by their greater algebraic complexity and the consequent greater labour of calculating them, so that the time required for computing one cycle becomes much greater than for the expected coefficients. This difficulty seemed to condemn the 'empirical' so long as the only tables available were of the type of those in this book for the 'expected'. Recent work by Cornfield and Mantel (1950) alters this situation by suggesting an improved computing scheme for the empirical coefficients and providing tables for use in it; the method is still more laborious, cycle for cycle, than that used in this book, but the advantage in speed of convergence may now be sufficient to outweigh this disadvantage. Extensive comparisons of the two will be necessary before a final decision is made as to which is best for routine use.

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%	0.0	0.1	0.5	0.3	0.4	0.2	0.6	0·7	0.8	0.9	1	2	3	4	5	264
0	-	1.9098	2.1218	2.2522	2.3479	2.4242	2.4879	2.5427	2.5911	2.6344	-					
1	2.6737	2·7 096	2.7429	2.7738	2·8027	2 ·8299	$2 \cdot 8556$	2.8799	$2 \cdot 9031$	$2 \cdot 9251$						
2 3	2.9463	2.9665	2.9859	3 ∙0046	3 ·0226	3 ∙0400	3 ∙0569	3.0732	3 ∙0890	3.1043	1 1	For m	ore de	etail a	ee	
3	3.1192	3.1337	3.1478	3.1616	3·17 50	3 ·1881	3.2009	3.2134	3.2256	3.2376				95-10		
4	3.2493	3.2608	3.2721	3.2831	3 ·2940	3.3046	3.3151	3.3253	3.3354	3.3454		·			Ū.	
5	3 ·3551	3.3648	3.3742	3.3836	3.3928	3.4018	3.4107	3.4195	3.4282	3.4368	9	18	27	36	45	
6	3.4452	3·4 536	3.4618	3.4699	3.4780	3·4859	3.4937	3.5015	3.5091	3.5167	8	16	24	32	40	
7	3.5242	3.5316	3·5389	3.5462	3.5534	3.5605	3.5675	3.5745	3.5813	3.5882	7	14	21	28	36	
8	3.5949	3 ·6016	3.6083	3.6148	3.6213	3.6278	3.6342	3.6405	3.6468	3.6531	6	13	19	26	32	
9	3 ·6592	3 ·6654	3.6715	3.6775	3.6835	3.6894	3.6953	3.7012	3 ·7070	3.7127	6	12	18	24	30	
10	3.7184	3.7241	3 ·7298	3 ·7354	3.7409	3.7464	3.7519	3.7574	3.7628	3 ·7681	6	11	17	22	28	
11	3.7735	3 ·7788	3 ·7840	3 ·7893	3.7945	3.7996	3.8048	3.8099	3.8150	3.8200	5	10	16	21	26	
12	3.8250	3.8300	3.8350	3.8399	3.8448	3.8497	3.8545	3.8593	3.8641	3.8689	5	10	15	20	24	
13	3.8736	3.8783	3.8830	3.8877	3.8923	3.8969	3.9015	3.9061	3.9107	3.9152	5	9	14	18	23	
14	3.9197	3.9242	3.9286	3.9331	3.9375	3.9419	3.9463	3.9506	3.9550	3.9593	4	9	13	18	22	
15	3.9636	3.9678	3.9721	3.9763	3 ·9806	3.9848	3 ∙9890	3 ·9931	3.9973	4.0014	4	8	13	17	21	
16	4.0055	4.0096	4.0137	4.0178	4.0218	4.0259	4.0299	4.0339	4.0379	4.0419	4	8	12	16	20	
17	4.0458	4.0498	4.0537	4.0576	4.0615	4.0654	4.0693	4.0731	4.0770	4.0808	4	8	12	16	1 9	
18	4.0846	4.0884	4.0922	4.0960	4.0998	4.1035	4.1073	4.1110	4.1147	4.1184	4	8	ñ	15	19	
19	4.1221	4.1258	4.1295	4.1331	4.1367	4.1404	4.1440	4.1476	4.1512	4.1548	4	7	ii	15	18	
20	4.1584	4.1619	4.1655	4.1690	4·1726	4.1761	4·1796	4.1831	4.1866	4-1901	4	7	11	14	18	
21	4.1936	4 ·1970	4.2005	4.2039	4.2074	4.2108	4.2142	4.2176	4.2210	4.2244	3	7	10	14	17	
22	4.2278	4.2312	4.2345	4.2379	4.2412	4.2446	4.2479	4.2512	4.2546	4.2579	3	7	10	13	17	
23	4.2612	4.2644	4.2677	4.2710	4.2743	4.2775	4.2808	4.2840	4.2872	4.2905	3	ż	10	13	16	
24	4·2937	4.2969	4.3001	4.3033	4.3065	4.3097	4.3129	4.3160	4.3192	4.3224	3	6	10	13	16	
25	4.3255	4.3287	4.3318	4.3349	4.3380	4.3412	4.3443	4.3474	4.3505	4.3536	3	6	9	12	16	
26	4.3567	4.3597	4.3628	4.3659	4.3689	4.3720	4.3750	4.3781	4.3811	4.3842	3	6	9	12	15	
27	4.3872	4.3902	4.3932	4.3962	4.3992	4.4022	4.4052	4.4082	4.4112	4.4142	3	6	9	12	15	
28	4.4172	4.4201	4.4231	4.4260	4.4290	4.4319	4.4349	4.4378	4.4408	4.4437	3	6	9	12	15	
29	4.4466	4.4495	4.4524	4.4554	4.4583	4.4612	4.4641	4.4670	4.4698	4.4437	3	6	9	12	14	

.

TABLE I. Transformation of Percentages to Probits

. . '

30	4.4756	4.4785	4.4813	4.4842	4-4871	4.4899	4.4928	4-4956	4.4985	4.5013	3	6	9	· 11	14
31	4 ·5041	4.5070	4 ·5098	4.5126	4.5155	4.5183	4.5211	4·5239	4.5267	4.5295	3	6	8	11	14
32	4.5323	4.5351	4.5379	4.5407	4.5435	4.5462	4.5490	4.5518	4.5546	4.5573	3	6	8	īī	14
33	4.5601	4.5628	4.5656	4.5684	4.5711	4.5739	4.5766	4.5793	4.5821	4.5848	3	5	8	ii	14
34	4.5875	4.5903	4.5930	4.5957	4.5984	4.6011	4.6039	4.6066	4.6093	4.6120	3	ğ	8	ii	14
	# 0070	* 0000	4 0000	± 0001	10001	4.0011	#.0039	#·0000	4.0083	4.0120	3	0		11	14
35	4.6147	4.6174	4.6201	4.6228	4 ·6255	4 ·6281	4.6308	4.6335	4.6362	4.6389	3	5	8	11	13
36	4 ·6415	4.6442	4·6469	4.6495	4.6522	4 ·6549	4.6575	4.6602	4.6628	4.6655	3	5	8	11	13
37	4.6681	4·6708	4.6734	4.6761	4.6787	4.6814	4.6840	4.6866	4.6893	4.6919	3	5	8	īī	13
38	4.6945	4.6971	4.6998	4.7024	4.7050	4.7076	4.7102	4.7129	4.7155	4.7181	3	5	8	10	13
39	4.7207	4.7233	4.7259	4.7285	4.7311	4.7337	4.7363	4.7389	4.7415	4.7441	3	5	8	10	13
•••		1 1200	* 1800	21200	17011	± 1001	#'/JUJ	7.1009	4.1410	4.1441	0	0	0	10	19
40	4.7467	4.7492	4.7518	4.7544	4.7570	4.7596	4.7622	4.7647	4.7673	4.7699	3	5	8	10	13
· 41	4.7725	4.7750	4.7776	4.7802	4.7827	4.7853	4.7879	4.7904	4.7930	4.7955	3	5	8	10	13
42	4.7981	4.8007	4.8032	4.8058	4.8083	4.8109	4.8134	4.8160	4.8185	4.8211	3	5	8	10	13
43	4.8236	4.8262	4.8287	4.8313	4.8338	4.8363	4.8389	4.8414	4.8440						
44	4.8490	4.8516	4.8541	4.8566						4.8465	3	5	8	- 10	13
**	4.0490	4.9910	4.0041	4.9000	4 ·8592	4.8617	4 ·8642	4.8668	4 ·8693	4.8718	3	5	8	10	13
45	4 ·8743	4.8769	4.8794	4.8819	4.8844	4.8870	4.8895	4.8920	4.8945	4-8970	3	5	8	10	13
46	4.8996	4.9021	4.9046	4.9071	4.9096	4.9122	4.9147	4.9172	4.9197	4.9222	3	5	8	- 10	13
47	4.9247	4.9272	4.9298	4.9323	4.9348	4.9373	4.9398	4.9423	4.9448	4.9473	3	5	8	10	13
48	4.9498	4.9524	4.9549	4.9574	4.9599	4.9624	4.9649	4.9674	4.9699	4.9724	3	5	8	10	13
49	4.9749	4.9774	4.9799	4.9825	4.9850										
Ŧ	10110	1.0111	#.0100	1.9020	4.9000	4 ·9875	4 ·9900	4 ·9925	4 ∙9950	4 ·9975	3	5	8	10	13
50	5.0000	5.0025	5.0050	5.0075	5-0100	5.0125	5.0150	5.0175	5.0201	5.0226	3	- 5	8	10	13
51	5 ·0251	5.0276	5.0301	5.0326	5.0351	5.0376	5.0401	5.0426	5.0451	5.0476	3	5	8	10	13
52	5.0502	5.0527	5.0552	5.0577	5.0602	5.0627	5.0652	5.0677	5.0702	5.0728	3	5	8	ĩŏ	13
53	5.0753	5.0778	5.0803	5.0828	5.0853	5.0878	5.0904	5.0929	5.0102	5.0979	3	5	8	10	13
54	5.1004	5.1030	5.1055	5.1080	5.1105										
	0.1004	0.1030	0.1009	0.1090	9.1109	5.1130	5.1156	5-1181	5.1206	5.1231	3	5	8	10	13
55	5.1257	5 ·1282	5.1307	5.1332	5.1358	5.1383	5.1408	5.1434	5 ·1459	5.1484	3	5	8	10	13
56	5.1510	5.1535	5.1560	5.1586	5.1611	5.1637	5.1662	5.1687	5.1713	5.1738	3	5	8	ĩŏ	13
57	5.1764	5.1789	5.1815	5.1840	5-1866	5.1891	5.1917	5.1942	5.1968	5.1993	3	5	8	10	13
58	5.2019	5.2045	5.2070	5.2096	5.2121	5.2147	5.2173	5·1942 5·2198							
59	5.2275	5·2301	5.2327						5·2224	5.2250	3	5	8	10	13 N
00 1	0-2210	0.7901	0.2971	5 ·2353	5.2378	5 ·2404	5.2430	5.2456	5·2482	5.2 508	3	5	8	10	13 N 13 G

TABLE I (cont.)

%	0.0	0.1	0.2	0.3	0-4	0.2	0.8	0.7	0.8	0.8	1.1	2	8	4	5	266
60	5.2533	5.2559	5.2585	5·2611	5.2637	5.2663	5·2689	5.2715	5.2741	5.2767	3	5	8	10	13	φ.
61	5.2793	5.2819	5.2845	5.2871	5.2898	5.2924	5.2950	5.2976	5.3002	5.3029	3	5	8	īŏ	13	
62	5.3055	5.3081	5.3107	5.3134	5.3160	5.8186	5.3213	5.3239	5.3266	5.3292	l ă	Ř	š	ĩĭ	13	
63	5.3319	5.3345	5.3372	5.3398	5.3425	5.3451	5.3478	5.3505	5.3531	5.3558	8	5 5	8	ii	13	
64	5.3585	5.8611	5.3638	5.3665	5.3692	5.3719	5.3745	5.3772	5.3799	5.3826	8	5	8	ii	13	
65	5-3853	5-3880	5-3907	5.3934	5.3961	5-3989	5-4016	5.4043	5-4070	5-4097	8	5	8	11	14	
66	5.4125	5.4152	5.4179	5.4207	5-4234	5.4261	5.4289	5.4316	5.4344	5.4372	3	5	8	ii	14	
67	5.4399	5.4427	5.4454	5.4482	5.4510	5.4538	5.4565	5.4593	5.4621	5.4649	8	ĕ	8	ii	14	
68	5.4677	5.4705	5.4733	5.4761	5.4789	5.4817	5.4845	5.4874	5.4902	5.4930	3	ĕ	8	ii	14	
69	5.4959	5.4987	5.5015	5.5044	5.5072	5.5101	5.5129	5.5158	5.5187	5.5215	3	6	9	ii	14	
70	5.5244	5.5273	5.5302	5-5330	5.5359	5.5388	5.5417	5.5446	5.5476	5.5505	3	6	9	12	14	
71	5.5534	5.5563	5.5592	5.5622	5.5651	5.5681	5.5710	5.5740	5.5769	5.5799	8	ő	ğ	12	15	
72	5.5828	5.5858	5.5888	5.5918	5.5948	5.5978	5.6008	5.6038	5.6068	5.6098		ĕ	ő	12	15	
73	5.6128	5.6158	5.6189	5.6219	5.6250	5.6280	5.6311	5.6341	5.6372	5.6403	3	ĕ	9	12	15	
74	5 6433	5.6464	5.6495	5.6526	5.6557	5.6588	5.6620	5.6651	5.6682	5.6713	8	ĕ	9	12	16	
75	5.6745	5.6776	5-6808	5.6840	5.6871	5-6903	5.6935	5.6967	5.6999	5.7031	8	6	10	13	16	
76	5.7063	5.7095	5.7128	5.7160	5.7192	5.7225	5.7257	5.7290	5.7323	5.7356	.8	7	10	13	16	
77	5.7388	5.7421	5.7454	5.7488	5.7521	5.7554	5.7588	5.7621	5.7655	5.7688	3	ż	ĩŏ	13	17	
78	5.7722	5.7756	5.7790	5.7824	5.7858	5.7892	5.7926	5.7961	5.7995	5-8030	3	-	iŏ	14	17	
79	5.8064	5.8099	5.8134	5.8169	5.8204	5.8239	5.8274	5-8310	5-8345	5.8381	4	ż	iĭ	14	18	
80	5.8416	5.8452	5.8488	5.8524	5-8560	5-8596	5-8633	5-8669	5.8705	5.8742	4	7	11	14	18	
81	5.8779	5.8816	5.8853	5.8890	5.8927	5-8965	5.9002	5.9040	5.9078	5-9116	4	ż	ii	15	19	
82	5.9154	5.9192	5.9230	5.9269	5.9307	5.9346	5.9385	5.9424	5.9463	5.9502		8	12	15	19	
83	5.9542	5.9581	5.9621	5.9661	5.9701	5.9741	5.9782	5.9822	5.9863	5.9904	4	8	12	16	20	
84	5.9945	5.9986	6.0027	6.0069	6.0110	6.0152	6.0194	6.0237	6.0279	6.0322	4	8	13	17	21	
85	6.0364	6.0407	6.0450	6.0494	6.0537	6-0581	6.0625	6 ∙0669	6.0714	6.0758	4	9	13	18	22	
86	6.0803	6.0848	6.0893	6.0939	6.0985	6.1031	6.1077	6.1123	6.1170	6.1217	5	ğ	14	18	23	
87	6.1264	6.1311	6.1359	6.1407	6.1455	6.1503	6.1552	6.1601	6.1650	8-1700	5	10	15	19	24	
88	6.1750	6.1800	6.1850	6-1901	6.1952	6.2004	6.2055	6.2107	6-2160	6-2212	5	10	15	21	26	
89	6-2265	6.2319	6.2372	6.2426	6.2481	6.2536	6-2591	6.2646	6-2702	6-2759	5	iĭ	16	22	27	

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90	6-2816	6.2873	6-2930	6-2988	6.3047	6-3106	6.3165	6 ∙ 3 225	6.3285	6·3346	16	12	18	24	29
91	6·34 08	6.3469	6.3532	6 ∙3595	6.3658	6.3722	6.3787	6.3852	6.3917	6.3984	6	13	19	26	32
92	6.4051	6.4118	6.4187	6·4255	6.4325	6.4395	6-4466	6.4538	6.4611	6.4684	1 7	14	21	28	35
93	6.4758	6.4833	6.4909	6.4985	6.5063	6.5141	6.5220	6.5301	6.5382	6.5464	8	16	24	31	39
94	6.5548	6.5632	6.5718	6.5805	6.5893	6.5982	6.6072	6.6164	6.6258	6.6352	9	18	27	36	45
1														•	
95	6.6449	6 ∙6546	6 ∙6646	6.6747	6 ∙6849	6·6954	6 ∙7060	6.7169	6·7279	6.7392					
	97	100	101	102	105	106	109	110	113	115					
96	6.7507	6.7624	6.7744	6.7866	6.7991	6.8119	6.8250	6.8384	6.8522	6.8663					
	117	120	122	125	128	131	134	138	141	145	1				
97	6.8808	6.8957	6.9110	6.9268	6·9431	6.9600	6.9774	J-9954	7.0141	7.0335	1				
	149	153	158	163	169	174	- 180	187	194	202	1				
•					200		100	107	101	202	I				
%	0.00	0.01	0.02	0.03	0.04	0.05	0.06	0.07	0.08	0.09	1	2	3	4	5
	· · · · · · · · · · · · · · · · · · ·														
98-0	7.0537	7.0558	7.0579	7.0600	7.0621	7.0642	7.0663	7.0684	7.0706	7.0727	2	4	6	8	11
98·1	7.0749	7.0770	7.0792	7.0814	7.0836	7.0858	7.0880	7.0902	7.0924	7.0947	2	4	7	9	11
98-2	7.0969	7.0992	7.1015	7.1038	7.1061	7.1084	7.1107	7.1130	7.1154	7-1177	2	5	7	ğ	12
98.3	7.1201	7.1224	7.1248	7.1272	7.1297	7.1321	7.1345	7.1370	7.1394	7.1419	2	5	7	10	12
98-4	7.1444	7.1469	7.1494	7.1520	7.1545	7.1571	7.1596	7.1622	7.1648	7.1675	3	5	8	īŏ	13
	•						1 2000			. 10.0	1	Ŭ	Ŭ		
98·5	7.1701	7.1727	7.1754	7.1781	7.1808	7.1835	7.1862	7.1890	7.1917	7.1945	3	5	8	11	14
98.6	7.1973	7.2001	7.2029	7.2058	7.2086	7.2115	7.2144	7.2173	7.2203	7.2232	3	ě	9	12	14
98.7	7.2262	7.2292	7.2322	7.2353	7.2383	7.2414	7.2445	7.2476	7.2508	7.2539	3	ĕ) 9	12	15
98.8	7.2571	7.2603	7.2636	7.2668	7.2701	7.2734	7.2768	7.2801	7.2835	7.2869	3	ž	1Ŏ	13	17
98.9	7.2904	7.2938	7.2973	7.3009	7.3044	7.3080	7.3116	7.3152	7.3189	7.3226	4	7	ñ	14	18
			. 2010		10011	1 0000	1.0110	1.0102	1 0100	1.0420	-	•		17	10
99.0	7.3263	7.3301	7.3339	7.3378	7-3416	7.3455	7.3495	7.3535	7.3575	7.3615	4	8	12	16	20
99.1	7·3 656	7.3698	7.3739	7.3781	7.3824	7.3867	7.3911	7.3954	7.3999	7.4044	4	ğ	13	17	22
99.2	7.4089	7.4135	7.4181	7.4228	7.4276	7.4324	7.4372	7.4422	7.4471	7.4522	5	10	14	19	24
99·3	7.4573	7.4624	7.4677	7.4730	7.4783	7.4838	7.4893	7.4949	7.5006	7.5063	5	ĩĭ	16	22	27
99.4	7.5121	7.5181	7.5241	7.5302	7.5364	7.5427	7.5491	7.5556	7.5622	7.5690	6	13	19	25	32
				10004	1 0004	10141	1.0401	1.0000	1.0022	1.0090	U U	10	10	20	04
99-5	7.5758	7.5828	7.5899	7.5972	7.6045	7.6121	7.6197	7.6276	7.6356	7.6437					
99·6	7.6521	7.6606	7.6693	7.6783	7.6874	7.6968	7.7065	7.7164	7.7266	7.7370	1				
99·7	7.7478	7.7589	7.7703	7.7822	7.7944	7.8070	7.8202	7.6338	7.8480	7.8627					
99.8	7.8782	7.8943	7.9112	7.9290	7.9478	7.9677	7.9889	8.0115	8.0357	8.0618					
99.9	8.0902	8.1214	8.1559	8.1947	8.2389	8.2905	8.3528	8.4316	8.5401	8.7190					
- '						0	3 0043	P 1010	J UIVI	3 1100	+				

I am indebted to Professor R. A. Fisher and Dr F. Yates, and also to Messrs Oliver and Boyd, Ltd. of Edinburgh, for permission to reprint Table I from Table IX of their book *Statistical Tables for Biological*, *Agricultural and Medical Research*.

•	TABLE II.	The	Weighting	Coefficient	and Q/Z	
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Percentage natural mortality, C

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Y	Q/Z	0	1	2	3	4	5	6	7	. 8	9	10
1.1	5034	·00082	·									
1.2	3425	+00118	·00001	_			<u> </u>	_	_			_
1.3	2354	-00167	·00002	·00001	·00001					_		
- 1-4	1634	·00235	-00004	.00002	·00001	-00001	·00001	-00001		_		
1.2	1146	·00327	.00007	·00004	.00002	·00002	·00001	·00001	·00001	·00001	•00001	·00001
1.6	811-2	.00451	.00015	·00007	•00005	·00004	·00003	·00002	·00002	-00002	·00002	.00001
1.7	580-2	·00614	-00028	·00014	.00009	.00007	.00006	.00005	·00004	·00003	-00003	·00003
1.8	419-1	+00828	·00053	·00027	+00018	·00013	.00011	•00009	.00007	-00006	.00006	.00005
1.9	305-8	·01105	.00097	.00050	·00034	.00025	.00020	·00017	00014	·00012	.00011	·00010
2.0	225.3	• •01457	·00172	.00090	•00061	·00046	·00036	·00030	·00026	·00022	.00020	-00017
2.1	167-69	•01903	·00297	·00159	·00108	·00082	-00065	·00054	·00046	·00040	.00035	·000 31
2.2	126.02	·02458	·00496	·00274	.00188	·00142	·00114	-00095	·00081	+00070	·00062	.00055
2.3	95.63	•03143	·00803	·00456	·00317	·00241	·00194	.00162	·00138	.00121	-00106	.00095
2.4	73.28	-03977	·01256	.00739	·00521	-00400	.00324	.00271	.00232	·00202	.00179	·00160
2.2	56.70	04979	·01895	-01161	.00832	·00646	·00525	·00441	•00379	.00332	·00294	·00264
2.6	44.288	·06168	·02763	01768	·01292	·01014	.00831	.00702	•00606	·00531	·00472	·00424
2.7	34.923	·07564	·03895	·02605	·01947	·01548	·01280	·01088	·00943	.00830	.00740	-00666
2.8	27.797	•09179	·05316	·03719	·02847	-02297	·01918	·01642	·01431	.01265	·01131	·01021
2.9	22.330	·11026	·07044	·05147	·04037	.03309	·02794	·02411	.02115	·01879	.01687	.01527
3.0	18-101	·13112	•09080	·06912	·05557	·04631	·03957	·03445	·03043	·02719	·02452	-02228
3.1	14.802	·15436	·11419	.09023	·07432	·06298	-05449	·04790	·04263	·03832	·03473	·03170
3∙2	12.211	·17994	·14046	11476	.09670	·08332	.07300	·06481	.05814	·05261	.04795	-04397
3.3	10.159	·20774	·16935	14249	·12263	·10736	·09525	.08541	-07726	.07039	. 06453	·05947
3.4	8.521	·23753	·20056	17308	·15184	·13494	·12116	·10973	·10008	·09182	08469	07846
3.2	7.205	·26907	·23373	20611	·18392	·16571	·15050	·13760	·12652	·11690	·10848	·10103

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3.6	6-1394	-30199	·26842	·24107	·21836	·19921	·18283	·16867	·15631	-14541	·13575	.12711
3.7	5-2705	·33589	·30415	-27741	·25456	·23482	·21759	·20242	·18896	·17694	·16614	·15639
3.8	4.5571	·37031	·34043	-31453	·29186	·27187	·25409	·23819	·22387	·21092	·19915	·18840
3.9	3.9676	·40474	·37669	•35181	·32960	·30964	·29161	·27524	·26031	·24665	·23409	·22250
4.0	3.4770	· 4 3863	•41237	•38864	·36707	·34739	·32937	·31279	·29749	·28334	·27020	·25797
4-1	3.0665	·47144	-44691	·42438	·40362	·38441	·36661	·35005	·33460	·32017	·30666	·29397
4.2	2.7206	·50260	·47973	·45844	· 43 858	·42000	·40259	·38623	•37085	·35634	·34264	· 3 2969
4.3	2.4276	•53159	·51029	•49024	·47134	·45350	· 4 3662	·42063	·40546	·39105	·37735	·36430
4.4	2.1780	·55788	·53806	51924_	·50134	·48430	•46805	·45255	·43774	·42357	·41002	·39702
4 ·5	1.9640	•58099	·56257	•54495	·52806	•51187	•49633	•48140	•46705	· 4 5325	•43996	·42716
4.6	1.7797	·60052	··58341	·56694	·55106	·53574	·52095	·50666	·49286	·47951	·46659	·45409
4.7	1.6202	·61609	·60022	•58485	•56996	·55551	·54150	·52790	·51470	·50187	•48941	·47729
4.8	1.4814	·62742	·61271	59840	·58446	·57089	·55766	·54478	·53221	•51996	·50801	·49635
4.9	1.3599	63431	·62069	·60737	59436	·58164	·56921	·55704	·54514	· 53 350	·52210	·51094
5.0	1.2533	· 6 3662	62401	•61165	•59953	•58765	·57599	·56455	•55332	·54230	·53149	·52087
5.1	1.1593	·63431	·62266	61120	·59994	•58886·	·57796	·56724	·55669	·54631	·53609	·52604
5.2	1.0759	·62742	·61667	:60607	·59562	·58532	·57516	·56515	·55527	·54553	·53592	·52644
5.3	1.0018	·61609	·60618	59639	·58672	·57717	·56773	·55841	·54919	·54008	-53108	·52219
5.4	0.9357	·60052	·59140	•58238	·57346	·56462	•55588	·54722	•53866	·53018	·52178	·51347
5.5	0·876 4	•58099	·57263	-56434	·55612	·54797	•53990	·53189	•52396	·51609	•50829	·50056
5.6	0.8230	-55788	·55022	·54262	·53507	·52759	·52015	·51278	·50545	·49818	•49097	· 48380
5.7	0.7749	·53159	·52460	·51765	·51075	·50389	· 497 08	·49030	·48357	·47688	·47024	·46363
5.8	0.7313	·50260	·49624	48992	·48363	·47737	·47114	·46495	·45879	·45266	·44657	·44050
5.9	0.6917	·47144	·46567	·45993	·45422	·44853	•44287 .	·43723	·43162	·42603	·42047	·41493
6.0	0.6557	•43863	•43343	42824	·42308	·41793	-41281	·40770	•40261	·39754	·39249	·38746
6-1	0.6227	40474	·40006	·39540	·39075	·38612	·38150	·37690	·37231	·36774	·36318	·35863
6 ∙2	0.5926	·37031	·36613	·36196	·35781	·35366	·34952	·34540	·34128	$\cdot 33718$	·33308	·32900
6.3	0.5649	·33589	·33218	·32847	·32477	·32108	·31740	$\cdot 31372$	·31006	·30640	·30274	·29910
6.4	0.5394	-30199	·29871	·29543	·29216	·28890	·28564	·28238	$\cdot 27913$	·27589	·27266	$\cdot 26942$
6.5	0.5158	·26907	·26619	·26331	·26044	·25757	·25470	·25184	24899	$\cdot 24613$	·24329	·24044

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Percentage natural mortality, C

					· P	ercentage	natural m	ortality, O	,			
<u>Y</u>	Q/Z	0	1	2	3	4	· 5	6	7	8	9	10
6.6	0.4940	·23753	·23502	·23251	·23001	·22751	·22501	·22251	·22001	·21752	·21503	·21255
6.7	0.4739	·20774	·20556	·20339	·20122	19905	19689	·19473	·19256	·19041	18825	·18609
6.8	0.4551	.17994	17808	.17621	·17435	.17249	.17063	.16877	·16691	16506	16320	16135
6.9	0.4376	·15436	·15277	.15118	14960	·14801	·14643	.14484	-14326	·14168	+14010	13852
7.0	0.4214	·13112	·12977	·12843	·12709	·12575	·12442	· ·12308	$\cdot 12174$	12040	·11907	·11773
7.1	0.4062	·11026	·10914	·10802	·10689	. 10577	10465	·10353	·10241	·10129	·10017	-09905
7.2	0.3919	•09179	+09086	·08993	·08900	.08807	·08714	-08621	·08528	·08435	.08342	08249
7.3	0.3786	·07564	07487	·07411	·07334	·07258	·07181	07105	.07029	·06952	.06876	.06800
7.4	0.3661	+06168	.06106	·06044	·05982	05920	.05858	.05795	05733	·05671	.05609	•05547
7.2	0.3543	04979	•04929	·04879	.04828	·04778	·04728	·04678	·04628	04578	·04528	.04478
7.6	0.3432	-03977	•03937	·03897	·03857	·03817	·03777	·03737	·03697	·03657	·03617	·03577
7.7	0.3327	·03143	·03112	·03080	·03048	·03017	02985	02954	02922	·02891	02859	·02828
7.8	0.3228	·02458	·02434	02409	.02385	·02360	.02335	·02311	·02286	.02261	·02237	·02212
7.9	0.3134	·01903	·01883	·01864	·01845	·01826	·01807	01788	.01769	·01750	.01731	-01712
8∙0	0·3046	·01457	·01442	·01428	·01413	·01399	01384	.01369	.01355	·01340	·01326	·01311
8-1	0.2962	·01104	·01093	·01082	·01071	·01060	·01049	·01038	·01027	·01016	·01005	·00993
8.2	0.2882	.00828	.00819	·00811	·00803	.00795	.00786	.00778	.00770	.00762	.00753	·00745
8.3	0.2806	+00614	·00608	·00602	.00596	.00590	·00583	.00577	.00571	·00565	.00559	+00553
8∙4	0.2734	.00451	.00446	·00442	·00437	·00433	.00428	.00424	.00419	·00415	·00410	•00406
8∙5	0.2666	.00327	·00324	·00321	·00318	·00314	.00311	.00308	.00305	.00301	.00298	.00295
8.6	0.2600	.00235	·00233	·00231	·00228	·00226	·00224	·00221	·00219	·00217	·00214	·00212
8.7	0.2538	.00167	.00166	·00164	.00162	·00161	.00159	.00157	.00156	.00154	00152	·00212
8.8	0.2478	+00118	.00117	.00116	·00114	-00113	00112	.00111	·00110	-00104	00102	·00106
8.9	0.2421	00082	.00081	-00080	.00080	.00079	00078	+00077	·00076	·00076	+00075	+00108
9·0	0.2367	.00056	·00056	·00055	.00055	·00054	·00054	.00053	·00053	.00052	00051	·00074

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¥	Q/Z	11	12	13	14	15	16	17	18	19	20
1.1	5034		_				_	-			
1.2	3425	i								—	
1.3	2354	- 1						_			_
1.4	1634	- I						_		_	
1.5	1146	•00001	·00001	·00001	-				-		_
1.6	811-2	-00001	·00001	-00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001
1.7	580-2	•00002	·00002	·00002	·00002	·00002	$\cdot 00002$	·00001	·00001	·00001	·00001
1.8	419-1	•00005	·00004	·00004	·00003	·00003	·00003	·00003	·00003	·00002	$\cdot 00002$
1.9	305-8	•00009	·00008	·00007	·00007	·00006	·00006	·00005	-00005	·00005	·00004
2 ·0	225-3	·00016	·00014	·00013	·00012	·00011	·00010	·00010	·00009	•00008	·00008
2.1	167-69	•00028	·00026	.00023	·00022	·00020	·00018	·00017	·00016	·00015	·00014
2.2	126-02	·00050	·00045	·00041	·00038	·00035	·00033	·00030	·00028	·00026	·00025
2.3	95-63.	-00086	·00078	·00071	·00066	·00061	.00056	.00052	·00049	·00046	·00043
2.4	73-28	·00145	·00131	·00120	·00111	·00102	.00095	.00088	·00083	·00077	.00073
2.5	56.70	·00238	·00217	·00199	·00183	-00169	.00157	·00147	·00137	·00128	00121
2.6	44-288	·00384	·00350	·00321	.00296	·00274	.00255	·00237	·00222	·00208	·00196
2.7	34.923	·00604	·00551	·00506	·00467	·00433	·00403	.00376	·00352	·00331	·00311
2.8	27.797	·00928	·00849	·00781	·00722	.00870	·00624	.00583	·00547	·00514	·00484
2.9	22.330	·01392	·01277	·01177	·01090	·01014	·00945	.00885	-00830	·00780	.00735
3.0	18-101	•02038	·01875	·01732	·01608	·01497	·01399	·01311	·01231	·01159	·01094
3 ·1	14-802	02910	-02685	·02488	·02315	.02160	·02022	·01898	-01786	·01684	·01590
3.2	12.211	•04053	·03753	·03488	·03254	·03044	.02856	.02686	02531	02390	.02261
8.3	10-159	.05505	.05117	·04772	04465	·04188	.03939	·03712	.03506	.03317	·03143
3.4	8.521	·07297	.06809	·06374	·05982	.05628	.05307	·05014	.04745	·04498	·04271
3.5	7.205	-09441	·08848	·08313	07829	·07389	.06987	.06618	·06278	.05965	·05674
3.6	6.1394	·11934	·11232	·10595	·10014	·09481	-08991	·08540	.08122	·07734	·07373
8.7	5-2705	·14753	·13945	·13205	·12525	11898	·11318	·10780	.10279	.09812	.09376
3.8 .	4.5571	·17854	·16947	·16111	·15336	.14616	·13946	·13321	·12736	12187	·11672
3.9	3.9676	·21179	·20185	·19260	·18398	·17591	·16836	·16127	·15460	·14831	·14237
4.0	3.4770	·24656	·23589	·22589	·21649	·20766	·19933	·19146	·18402	.17698	.17029

TABLE II (cont.)

Percentage natural mortality, O

Y	Q Z	11	12	13	14	15	16	17	18	19	20
4.1	3.0665	·28204	·27081	·26020	·25017	·24068	·23168	·22314	·21501	·20728	·19991
4.2	2.7206	.31742	.30578	·29473	·28421	·27420	·26465	·25554	·24684	$\cdot 23852$	·23055
4.3	2.4276	.35186	.33998	32864	·31779	·30740	·29744	·28789	·27873	·26992	·26145
4.4	2.1780	•38457	·37261	·36112	.35008	·33945	·32922	·31937	·30986	·30069	·29184
4.5	1.9640	41482	•40292	.39142	·38033	·36960	$\cdot 35922$	·34919	·33947	·33006	•32094
4.6	1.7797	•44198	-43025	•41887	·40784	·39713	-38674	37664	·36683	·35729	·34802
4.7	1 6202	·46551	·45405	·44289	·43202	·42144	-41113	·40109	·39129	·38174	·37242
4.8	1.4814	·48496	·47385	·46299	·45239	·44202	· 43190	·42199	•41231	•40284	·39357
4.9	1.3599	·50001	-48931	·47883	·46855	45849	·44862	·43894	·42945	·42015	$\cdot 41102$
5.0	1.2533	-51044	.50020	·49014	48026	·47054	·46100	-45162	44240	· 4 3333	•42441
5-1	1.1593	·51614	.50639	•49680	·48735	·47804	·46887	·45984	45094	-44217	·43354
5.2	1.0759	.51709	.50787	49876	·48978	·48091	·47216	·46353	·45500	·44658	·43827
5.3	1-0018	·51340	·50471	·49612	·48762	·47923	·47092	·46271	•45459	·44657	•43863
5.4	0.9357	.50524	·49709	·48903	·48104	·47313	·46529	·45754	•44985	·44224	•43470
5.5	0.8764	•49289	48529	·47775	·47028	·46286	·45551	·44822	•44099	•43382	·42671
5.6	0.8230	·47669	·46963	·46262	·45567	·44876	·44190	·43509	42832	·42161	·41494
5.7	0.7749	•45706	•45054	·44406	·43761	·43120	·42484	·41851	$\cdot 41222$	·40597	·39975
5.8	0.7313	+43447	·42847	·42250	·41656	·41066	·40478	·39893	·39311	·38733	·38157
59	0.6917	•40942	·40393	·39846	·39302	·38761	·38221	·37684	·37149	·36617	·36087
6.0	0.6557	·38245	.37745	·37248	·36752	·36258	·35766	·35275	•34787	·34300	·33815
6-1	0.6227	.35410	·34958	·34508	·34059	·33611	·33165	·32720	·32276	·31834	·31393
6.2	0.5926	·32493	32087	·31681	·31277	·30874	·30472	·30071	$\cdot 29671$	$\cdot 29272$	·28874
6.3	0.5649	·29546	·29183	·28821	-28460	·28099	·27739	·27380	·27022	·26664	·26308
6.4	0.5394	· ·26620	-26298	·25977	·25656	·25335	·25016	·24696	·24378	·24060	·23742
6.5	0.5158	·23760	·23476	·23193	22910	·22628	-22346	·22064	·21783	·21502	·21222

6.6	0.4940	·21007	·20759	·20511	·20264	-20016	·19770	·19523	·19277	·19031	
► 6·7	0.4739	·18394	·18179	·17964	·17749	$\cdot 17535$	$\cdot 17320$	·17106	$\cdot 16892$	·16679	
6.8	0.4551	·15950	·15765	·15580	·15395	$\cdot 15210$	·15026	·14841	·14657	·14473	
6-9	0.4376	·13694	·13536	$\cdot 13378$	$\cdot 13220$	·13063	-12905	·12748·	·12591	$\cdot 12433$	
7.0	0-4214	·11640	-11506	·11373	·11239	·11106	·10973	·10840	·10707	· •10574	•
7.1	0.4062	·09794	·09682	.09570	09458	·09347	.09235	·09123	·09012	·08900	
7.2	0.3919	•08157	·08064	07971	-07878	·07786	·07693	·07600	·07508	·07415	
7.3	0.3786	·06724	·06647	-06571	$\cdot 06495$.06419	·06342	·06266	•06190	·06114	
7.4	0.3661	·05485	·05423	·05361	·05299	.05237	· ·05175	·05113	·05051	·04989	
7.5	0.3543	·04428	·04378	·04328	-04278	·04228	·04178	·04128	·04078	·04028	
7.6	0.3432	.03537	·03498	·03458	·03418	·03378	·03338	·03298/	·03258	·03218	
7.7	0.3327	•02796	·02765	·02733	·02702	·02670	•02639	·02607	·02576	·02544	
7.8	0.3228	-02187	·02163	·02138	·02114	·02089	·02064	·02040	·02015	·01990	
7.9	0.3134	•01693	-01674	·01655	·01636	·01617	-01598	·01579	·01560	·01541	
8.0	0.3046	•01297	•01282	·01267	·01253	01238	·01224	·01209	·01194	·01180	
	0.0000	.00982	·00971	·00960	·00949	.00938	·00927	00070	00005	00004	
8-1	0.2962	·00982 ·00737	.00971	·00980	·00949	·00938	·00927 ·00695	·00916	·00905	·00894	
8.2	0.2882	+00737	.00728	00534	00528	00522	·00516	·00687	·00679	·00670	
8.3	0·2806 0·2734	•00347	·00340	00392	·00328	·00322	·00318	·00510 ·00374	·00504	·00497	
8.4	0.2666	•00401	00397	00392	·00388	00385	00379	·00374 ·00272	·00370	•00365	
8.2	0.2000	.00291	.00400	.00260	.00284	.00278	.00475	·00272	·00269	·00265	
8.6	0.2600	-00209	·00207	·00205	·00202	·00200	·00198	·00195	·00193	·00191	
8.7	0.2538	•00149	·00147	·00145	·00144	·00142	·00140	·00139	•00137	·00135	
8.8	0.2478	•00105	·00104	·00103	·00101	•00100	•00099	·00098	·00097	·00096	
. 8.9	0.2421	•00073	·00072	·00071	·00071	•00070	·00069	·00068	·00067	·00067	
9 ·0	0.2367	·00050	·00050	·00049	·00049	·00048	·00047	·00047	·00046	·00046	
N SE			•								

TABLE	II (cont.)
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Percentage natural mortality, C

¥	.Q/Z	21	22	23	24	25	26	27	28	29	30
1.1	5034						-	_	-		
$1 \cdot 2$	3425	· -			_				—	—	—
1.3	2354	i	-	-	—		_		. —		-
1-4	1634	I									
1.2	1146	— ·			_	—	-				
1.6	811-2	.00001	·00001	·00001		_					
1.7	580-2	+00001	·00001	·00001	·00001	-00001	·00001	+00001	·00001	·00001	·00001
1.8	419-1	·00002	00002	·00002	·00002	·00002	·00002	·00002	•00001	·00001	+00001
1.9	305-8	•00004	·00004	·00004	·00003	·00003	·00003	·00003	·00003	·00003	·00002
2.0	225.8	·00007	·00007	·00007	·00006	·00006	·00006	·00005	·00005	·00005	·00005
2·1 、	167-69	.00013	.00013	·00012	·00011	·00011	•00010	.00010	•00009	•00009	·00008
2.2	126.02	•00023	·00022	·00021	·00020	+00019	·00018	00017	+00016	·00015	·00015
2.3	95-63	·00040	+00038	00036	·00084	·00032	·00031	·00029	·00028	·00026	·00025
2.4	73-28	.00069	-00065	·00061	·00058	.00055	.00052	·00049	·00047	+00045	·00043
2.2	56.70	•00114	.00107	.00101	·00096	-00091	•00086	·00082	·00078	·00075	·00071
2.6	44-288	-00185	·00174	.00165	.00156	·00148	·00141	·00134	·00127	.00121	·00116
2.7	84.923	·00293 ′	.00277	·00262	·00248	·00236	·00224	·00213	·00203	·00194	·00185
2.8	27.797	.00456	·00431	·00408	·00387	·00368	·00349	·00333	·00317	·00302	·00288
2.9	22.330	·00694	·00657	·00622	·00590	·00561	·00533	·00508	·00484	·00462	·00441
3 ∙0	18-101	•01034	·00979	·00928	·00881	·00838	•00797	·00760	·00725	·00692	•00661
8-1	14.802	.01505	01426	·01354	·01287	·01224	·01166	·01112	-01061	·01014	·00969
3 ∙2	12-211	·02143	·02033	·01932	·01838	·01751	·01669	·01593	·01522	·01455	$\cdot 01392$
3.3	10.159	.02983	·02834	.02697	·02569	·02450	·02338	·02234	·02136	·02044	·01957
3.4	8.521	·04060	·03864	·03682	·03512	03354	.03205	·03065	·02934	·02810	·02693
3.2	7.205	•05404	.05153	.04918	·04698	·04492	·04299	-04117	·03945	·03782	·03629

	3.6	6-1394	.07037	.06722	.06427	.06150	·05889	·05644	·05412	·05193	·04985	.04788
	3.7	5.2705	08966	08582	·08221	·07881	.07559	·07255	.06967	.06695	·06435	·06189
	3.8	4.5571	·11187	10730	10298	.09890	·09503	·09136	·08787	·08455	·08139	.07838
	3.9	3.9676	·13676	·13145	·12641	·12163	·11708	.11275	·10862	·10468	10091	.09732
	4.0	8.4770	·16394	·15791	·15216	·14668	.14145	·13645	·13167	·12710	·12271	·11851
												11501
	4-1	3.0665	·19288	·18616	·17974	·17360	·16771	·16207	·15665	·15145	·14645	·14164
	4.2	2.7206	·22291	·21559	·20855	·20180	·19531	·18906	·18304	.17725	+17166	·16626
	4.3	2.4276	·25331	·24546	·23790	·23061	·22358	·21679	·21023	·20389	·19776	19182
	4.4	2.1780	·28329	·27503	·26704	·25930	·25181	·24456	·23753	·23072	·22411	21769
	4.5	1.9640	·31210	·30352	29520	·28712	27927	·27165	·26424	·25703	·25002	·24319
	••		01210			20112		21100		10100	20002	#1010
	4.6	1.7797	·33900	·33022	32167	·31335	·30524	·29734	·28963	·28212	·27479	·26764
	4.7	1.6202	·36332	·35444	.34578	·33731	·32904	·32095	·31305	·30533	·29777	-29038
	4.8	1.4814	38450	·37562	·36693	·35841	·35007	·34190	·33390	·32605	·31836	·31082
	4.9	1.3599	.40206	·39327	·38464	·37617	.36785	·35968	·35166	·34378	·33604	·32843
	5.0	1.2533	-41564	·40702	·39853	-39019	·38197	·37389	·36593	-35810	·35039	·34279
								0.000			00000	01210
	5.1	1.1593	·42502	·41663	·40836	·40020	·39216	·38423	-37641	·36870	·36109	·35359
	5.2	1.0759	43007	·42196	·41396	40606	·39825	·39054	38292	·37540	·36796	·36062
	5.3	1.0018	43077	·42300	·41532	.40772	·40020	·39276	·38540	·37812	·37091	·36378
	5.4	0.9357	·42724	·41984	·41252	40526	·39807	·39094	·38388	.37689	.36996	·36309
	5.5	0.8764	·41966	41266	·40572	·39884	·39201	·38524	·37852	.37185	-36524	·35868
						00002	00201	00011	0.000	0.100	00022	00000
	5.6	0.8230	·40832	-40174	·39521	·38873	·38229	·37590	·36954	·36324	·35697	·35075
	5.7	0.7749	39357	·38743	38133	·37526	·36923	·36323	·35727	.35134	·34545	·33959
	5.8	0.7313	·37584	·37014	·36447	·35883	·35322	·34763	·34207	·33655	·33104	·32557
	5.9	0.6917	35559	·35033	·34510	.33989	·33470	·32954	·32439	·31927	·31417	·30909
	6 ∙0	0.6557	·33332	·32850	·32370	·31892	·31416	·30941	·30469	·29997	29528	·29060
		•				01002	01110		00100	20001	20020	20000
	6·1	0.6227	·30954	·30516	·30079	·29643	·29209	·28776	·28344	·27914	·27485	·27057
	6.2	0.5926	·28477	·28081	·27686	·27292	·26899	·26507	·26116	·25726	·25337	-24949
•	6.3	0.5649	·25952	25596	·25242	·24888	·24535	·24182	·23831	·23480	·23130	22780
	6.4	0.5394	·23425	·23109	·22793	·22477	22163	·21848	·21535	·21221	·20909	20597
	6.2	0.5158	·20942	·20662	·20383	·20104	·19825	·19547	·19270	·18992	18715	18439
		•		- ,			20010					10100

18-3

TABLE II (cont.)

					rerce	епсаде пасо	Let moreau	.uy, U			
y	Q Z	21	22	23	24	25	26	27	28	29	30
	0.4940	·18540	·18294	.18049	.17805	·17561	·17317	·17073	·16829	·16586	·16343
6.6	0.4739	16252	·16039	15826	·15613	$\cdot 15401$	·15188	·14976	·14764	$\cdot 14552$	·14341
6.7		•14105	.13921	·13738	·13554	13371	·13188	$\cdot 13005$	$\cdot 12822$	$\cdot 12639$	·12457
6.8	0.4551	-12119	·11962	.11805	-11649	.11492	·11336	·11179	$\cdot 11023$	·10866	·10710
6.9	0.4376			·10042	-09909	-09777	.09644	·09512	.09379	·09247	·09114
7.0	0.4214	·10308	$\cdot 10175$	10042	-03308	00111	00011	00012			•
	0.4000	-08677	·08566	·08455	·08343	·08232	·08121	·08010	·07899	•07787	·07676
7.1	0.4062	-07230	.07137	-07045	·06953	-06860	.06768	.06675	-06583	·06491	·06398
7.2	0.3919		.05886	-05809	.05733	•05657	05581	.05505	.05429	·05353	·05277
7.3	0.3786	·05962		-04741	00133	-04617	-04555	·04493	·04431	·04369	·04307
7.4	0.3661	·04865	·04803		04079	-03728	-03678	·03628	.03578	.03528	.03479
7.5	0.3543	·03928	-03878	·03828	.03778	.03120	-03010	00020	00010	00010	
- a ·		00100	.03099	·03059	·03019	·02979	·02939	.02899	·02860	·02820	·02780
7.6	0.3432	·03139		·02418	+02387	-02355	.02324	.02292	-02261	02229	·02198
.7.7	0.3327	·02481	-02450		·02387	·01843	·01818	-01793	·01769	·01744	·01720
7.8	0.3228	·01941	·01917	·01892			·01407	·01388	·01369	·01350	·01331
7.9	0.3134	·01502	·01483	·01464	·01445	·01426		·01063	-01049	·01034	·01019
8.0	0.3046	•01151	·01136	$\cdot 01122$	·01107	$\cdot 01092$	·01078	.01003	-01040	01001	
		00070	-00861	.00850	·00839	.00828	-00817	·00806	·00795	-00784	·00773
8-1	0.2962	·00872	+00646	·00637	+00629	.00621	.00612	·00604	.00596	·00588	·00579
8.2	· 0·2882	·00654		00037	·00467	.00461	·00454	.00448	·00442	·00436	·00430
8.3	0.2806	·00485	·00479		00407	.00338	.00334	.00329	.00325	.00320	·00316
8.4	0.2734	·00356	·00352	·00347			·00334	.00239	.00236	.00232	·00229
8.5	0.2666	·00259	·00255	$\cdot 00252$	·00249	·00246	-00242	-00200	00200	00202	
	0.0000	·00186	·00184	·00181	·00179	·00176	.00174	.00172	·00169	·00167	·00165
8.6	0.2600		·00134	-00129	.00127	·00125	·00124	-00122	·00120	·00119	·00117
8.7	0.2538	·00132		00128	-00090	.00088	.00087	.00086	.00085	·00084	·00083
8-8	0.2478	·00093	·00092		·00090	·00088	-00061	-00060	.00059	-00058	.00057
8-9	0.2421	•00065	·00064	·00063		·00042	.00042	+00041	·00041	·00040	·00040
9.0	0.2367	·00045	·00044	·00044	·00043	-00042	00042	00011			- / •

Percentage natural mortality, C

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Y	Q/Z	31	32	33	34	35	36	37	38	39	40
1.1	5034										_
1.2	3425	- 1				—					
1.3	2354		—				-				
1.4	1634							-		-	
1.5	1146	-		-	 -		. —			-	
1.6	811-2	_								_	-
1.7	580-2	.00001	·00001	·00001	-00001	·00001	00001	·00001	·00001	<u> </u>	
1.9	419-1	.00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001
1·8 1·9	305-8	-00002	·00002	·00002	$\cdot 00002$	·00002	·00002	·00002	·00002	·00002	·00002
1.9 2.0	225.3	·00004	·00004	·00004)004	·00004	·00003	·00003	·00003	·0000 3	·0000 3
2.0	220.3	.00004	00001	00001							
	107 00	.00008	.00008	·00007	·00007	·00007	·00006	·00006	·00006	·00006	·00005
2.1	167-69	00014	-00013	·00013	.00012	.00012	·00011	·00011	·00010	·00010	·00009
2.2	126.02		·00013	.00022	-00021	.00020	.00019	.00018	·00018	·00017	·00016
2.3	95-63	•00024	·00023	000022	·00036	·00034	.00033	·00031	·00030	.00029	·00028
2.4	73.28	•00041		-00062	·00059	.00057	·00054	.00052	.00050	.00048	·00046
2.5	56.70	•00068	·00065	.00062	•00059	-00007	-00034	00002	00000	00010	
2.6	44.288	-00111	·00106	·00101	·00097	·00092	·00089	·00085	·00081	·00078	·00075
2.0	34.923	+00176	·00169	00161	·00154	·00148	.00142	·00136	·00130	.00125	·00120
	27.797	+00276	.00263	.00252	·00241	.00231	.00221	·00212	·00204	·00195	·00188
2.8		-00422	·00403	.00386	.00370	·00354	·00339	$\cdot 00325$	00312	·00300	·00288
2.9	22.330		·00403	.00579	·00555	.00532	.00510	·00489	·00469	-00451	·00433
3 ∙0	18-101	·00632	.00009	00019	-00000	00002	00010				
3.1	14.802	.00927	·00888	·00850	.00815	.00782	.00750	·00720	·00691	·00664	·00637
3·1 3·2	12.211	-01333	-01276	·01223	·01173	·01126	.01080	·01037	·00996	·00957	·00920
	12.211	-01875	01797	·01724	·01654	·01588	·01525	·01465	·01408	·01354	·01302
3.3		.02582	.02478	.02378	·02284	-02194	·02109	·02027	·01·949	.01875	·01804
3.4	8.521		.03345	·03214	-03089	·02970	.02856	·02748	·02645	·02546	·02451
3.2	7.205	•03483	.09940	-03414	00000	02010	22000				

					rerce	ntage natu	rai mortali	ty, O			
Y	Q/Z	81	82	83	84	85	86	87	88	89	40
8.6	6.1394	•04601	·04423	04254	•04093	·03938	·03791	·03651	·03516	.03387	·03263
8.7	5.2705	05954	·05731	.05517	05313	.05118	.04932	.04753	.04581	.04417	04259
8.8	4.5571	07551	.07276	•07013	.06761	06520	06289	.06067	05853	-05648	05451
8.9	8.9676	09387	.09057	·08741	.08437	08145	07865	.07595	+07835	07085	•06844
4.0	8.4770	•11447	11059	10687	10328	09988	.09650	.09329	09020	08721	08432
4-1	8.0665	.13701	·13255	·12825	.12410	·12010	·11628	·11249	·10888	.10538	·10200
4.2	2.7206	+16106	·15603	·15116	14646	·14191	·18751	·18324	12910	.12509	·12120
4.8	2.4276	18608	18051	.17512	·16989	·16481	15989	·15511	15046	.14595	.14156
4.4	2.1780	·21146	·20541	19953	.19382	·18826	18285	.17758	.17246	16747	16261
4.5	1.9640	·23655	·23008	·22377	·21763	·21164	·20580	20010	.19454	18911	18382
4-6	1.7797	·26066	·25384	·24719	·24069	·23433	·22812	·22205	·21611	·21031	·20462
4.7	1.6202	·28315	·27607	·26914	·26236	·25572	·24921	·24283	·23659	.23046	-22446
4.8	1.4814	·30342	·29617	·28905	·28206	27521	·26848	-26188	·25539	-24902	24276
4.9	1.8599	·82096	·81361	·30638	·29928	·29229	·28542	·27866	·27201	26547	•25904
5.0	1.2533	·83532	·82795	·82070	·81356	·30652	·29958	29275	-28602	27938	27284
5-1	1.1593	•34619	•33889	·33168	·32457	·31756	·81063	·30380	·29705	29039	·28381
5.2	1.0759	·85336	·84618	·83909	·83209	82516	81832	-81155	.30486	·29824	29170
5.3	1.0018	.85672	·84974	·84282	83598	·82921	·82250	-81587	.30930	.80279	·29635
5.4	0.9357	·35629	·34954	.84286	-33624	·32967	.32317	·31672	·31032	.30399	·29770
5.5	0.8764	·85217	·34571	·33930	·33294	·32663	·82037	·31416	·30799	·30187	-29580
5.6	0.8230	·84457	·33843	.33233	·32628	·32026	· 31428	·30835	·30245	·29659	-29077
5.7	0.7749	·83377	.32798	.32222	·81650	·81081	.30515	·29953	-29393	-28837	-28284
5.8	0.7318	·82012	·81470	.80931	-30394	29860	·29329	·28800	·28273	-27750	27229
5.9	0.6917	·30403	·29899	-29398	·28898	·28401	·27906	+27413	-26921	26432	25945
6.0	0.6557	·28594	28129	27666	27205	26746	-26288	25831	·25377	-24923	·24472

TABLE II (cont.)

Percentage natural mortality. O

6.1	0.6227 1	·26631	·26206	·25782	·25359	·24938	·24518	·24099	·23681	·23265	$\cdot 22850$
6.2	0.5926	·24561	·24175	·23790	·23406	·23022	·22640	·22259	·21878	·21499	·21120
6.3	0.5649	·22431	·22083	·21736	21390	·21044	20699	·20354	·20010	·19667	$\cdot 19325$
6.4	0.5394	20285	·19974	·19663	·19353	·19044	·18735	·18426	-18119	.17811	·17504
6.5	0.5158	·18163	.17887	·17612	·17337	·17062	·16788	·16514	·16240	·15967	·15695
		10100									
6.6	0.4940	·16101	·15858	·15616	·15374	·15133	·14891	·14650	·14410	·14169	·13929
6.7	0.4739	·14129	·13918	·13707	·13496	·13286	·13075	$\cdot 12865$	·12655	$\cdot 12445$	·12236
6.8	0.4551	·12274	·12092	·11910	· ·11728	·11546	·11364	·11182	·11001	·10819	·10638
6.9	0.4376	·10554	·10398	·10242	·10086	·09931	.09775	·09619	·09464	·09309	·0915 3
7.0	0.4214	·08982	·08850	·08718	·08586	·08454	·08322	·08190	·08058	·07926	·07794
		••••									
7.1	0.4062	-07565	·07454	·07343	$\cdot 07232$	-07122	·07011	•06900	·06789	·06679	·06568
7.2	0.3919	·06306	·06214	.06121	·06029	·05937	·05845	·05753	·05661	·05569	·05477
7.3	0.3786	.05201	·05125	·05049	·04974	·04898	·04822	·04746	·04670	·04594	·04519
7.4	0.3661	-04245	·04183	·04122	·04060	·03998	·03936	·03874	·03812	·03751	·03689
7.5	0.3543	.03429	·03379	·03329	·03279	·03229	·03179	·03129	·03079	·03030	·02980
		•••									
7.6	0.3432	·02740	·02700	·02660	·02620	·02581	·02541	·02501	·02461	·02421	·02382
7.7	0.3327	.02166	02135	·02103	·02072	·02041	·02009	·01978	· ·01946	-01915	·01883
7.8	0.3228	·01695	·01670	·01646	·01621	·01597	-01572	·01547	·01523	·01498	·01474
7.9	0.3134	-01312	·01293	·01274	01255	·01236	·01217	·01198	·01179	·01160 .	·01141
8.0	0.3046	.01005	.00990	·00976	.00961	·00947	·00932	·00917	·00903	+00888	·00874
	••••	•••••									
8.1	0.2962	·00762	·00750	·00739°	·00728	·00717	-00706	·00695	·00684	·00673	·00662
8.2	0.2882	.00571	.00563	·00554	·00546	·00538	·00530	·00521	·00513	.00505	·00497
8.3	0.2806	.00424	·00418	·00411	·00405	.00399	·00393	·00387	00381	00375	.00368
8.4	0.2734	+00311	00307	.00302	.00298	.00293	.00289	.00284	.00279	.00275	.00270
8.5	0.2666 .	-00226	.00223	·00219	·00216	00213	.00210	.00206	.00203	.00200	·00196
	0 2000	00220	00220	00-10		00110					
8.6	0.2600	·00162	·00160	·00158	-00155	·00153	·00151	·00148	·00146	·00144	·00141
8.7	0.2538	+00115	00114	·00112	00110	+00109	.00107	.00105	·00104	.00102	·00100
8.8	0.2478	.00081	.00080	.00079	.00078	00077	.00075	.00074	.00073	.00072	·00071
8.9	0.2421	+00057	.00056	.00055	.00054	.00053	.00053	+00052	.00051	.00050	.00049
9.0	0.2367	.00039	+00038	.00038	.00037	.00037	.00036	·00036	.00035	.00034	·00034
	0 2001 1	00000	00000	00000						0.001	

I am indebted to the Editors of the Annals of Applied Biology for permission to reproduce the first two sections of this table.

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		Percentage natural mortality, C												
Y	Q/Z	41	42	43	44	45	46	47	48	49	50			
1.8	419.1	•00001	•00001	·00001	·00001	•00001	·00001	·00001	.00001	·00001	·00001			
1.9	305-8	·00002	·00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001	·00001			
2.0	225·3	·00003	·0000 3	·00003	·00002	·00002	·00002	·00002	·00002	·00002	·00002			
2.1	167-69	·00005	·00005	.00005	-00005	·00004	·00004	·00004	·00004	·00004	·00004			
2.2	126.02	·00009	·00009	·00008	·00008	•00008 ·	·00007	·00007	·00007	·00007	·00006			
2.3	95-63	·00016	·00015	·00014	·00014	·00013	·00013	·00012	·00012	·00011	·00011			
2.4	73-28	·00026	·00025	·00024	·00023	·00023	·00022	·00021	·00020	·00019	·00018			
2.5	56.70	·00044	00042	·00041	·00039	·00038	·00036	·00035	·00033	·00032	·000 31			
2.6	44-288	.00072	·00069	·00066	·00064	·00061	·00059	·00056	·00054	$\cdot 00052$	·00050			
2.7	34 923	·00115	·00110	·00106	·00102	·00098	·00094	·00090	.00087	·00083	·00080			
2.8	27.797	·00180	·00173	·00166	·00160	-00153	·00147	$\cdot 00142$	·00136	·001 31	·00126			
2.9	22.330	.00276	.00265	-00255	·00245	·00236	·00226	·00218	-00209	·00201	·00194			
3 ·0	18-101	·00 416	·00399	·00384	·00369	·00355	·00341	·00328	·00315	·00303	·00292			
3-1	14.802	-00613	·00589	·00566	·00544	·00523	·00503	·00484	·00466	·00448	·00431			
3·2	12.211	.00885	·00851	·00818	·00787	·00757	·00728	·00701	·00674	·00649	·00624			
3.3	10-159	.01252	·01204	·01159	-01115	·01073	·01033	·00994	·00957	·00921	-00886			
3∙4	8.521	.01736	·01671	·01609	·01549	·01491	·01436	$\cdot 01382$	·01331	+01282	·01234			
3.2	7.205	.02360	·02273	·02189	·02109	·02031	·01957	·01885	·01816	·01749	·01685			
36	6-1394	·03144	·03030	·02920	·02815	.02713	·02615	·02521	·02429	·02342	·02257			
3·7	5.2705	·04107	·03961	·03820	·03684	·03554	·03427	·03306	·03188	·03074	·02964			
3.8	· 4·5571	·05261	·05078	·04901	.04730	·04566	·04407	·04253	·04105	·03961	·03821			
8.9	3-9676	·06611	·06386	·06169	·05959	·05757	·05560	·05370	.05186	·05008	·04835			
4 ·0	3.4770	-08153	·07883	·07622	07369	·07124	-06887	·06657	·06433	·06217	·06006			

TABLE II (cont.)

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4-1	3.0665	·09872	·09554	·09246	·08948	·08658	·08376	·08103	-07838	·07579	-07328
4.2	2.7206	-11743	·11376	·11020	·10674	·10337	·10010	$\cdot 09692$	$\cdot 09382$	·09080	·08786
4.3	2.4276	·13729	·13314	·12910	·12516	·12133	·11759	·11395	·11040	·10694	·10357
4.4	2.1780	·15787	$\cdot 15325$	·14874	·14434	·14005	·13587	$\cdot 13178$	$\cdot 12778$	·12388	·12007
4.5	1.9640	·17864	·17359	·16865	$\cdot 16382$	·15910	·15448	·14997	·14555	·14122	·13699
4.6	1.7797	·19906	·19362	·18829	·18307	·17796	·17295	·16804	·16323	$\cdot 15852$	·15390
4.7	1.6202	·21857	·21280	·20713	·20158	$\cdot 19612$	-19077	$\cdot 18552$	·18036	·17530	$\cdot 17032$
4.8	1.4814	-23662	·23058	·22464	·21881	$\cdot 21307$	·20744	·20189	·19644	·19108	·18581
4.9	1.3599	·25270	·24647	·24033	·23428	·22833	·22247	·21670	·21102	·20542	·19990
5.0	1.2533	·26639	·26003	·25376	·24757	·24148	·23546	·22953	·22368	·21790	·21221
0.0	1.7033	20000	20000								
e 1	1.1593	·27732	·27091	·26458	$\cdot 25832$	$\cdot 25215$	·24605	$\cdot 24002$	·23407	·22819	·22237
5.1	1.0759	-28524	·27884	·27252	·26626	·26008	·25396	·24790	$\cdot 24192$	·23599	•2301 3
5.2	1.0018	·28998	-28366	·27741	.27122	·26509	·25901	·25300	·24704	·24114	·23530
5.3	0.9357	·29148	·28530	·27918	•27311	·26709	·26113	·25521	·24935	·24353	·23776
5.4		·28977	·28379	-27785	-27196	·26611	·26031	·25454	·24882	·24314	·23751
5.2	0.8764	.29911	.70212	-21100	21100		20001	2010-			
~ ~	0.0000	·28499	·27925	·27354	·26787	·26224	·25664	·25108	·24555	·24006	·23461
5.6 .	0.8230	·28499	·27925 ·27187	·26644	·26103	-25565	-25030	·24499	·23970	·23444	·22921
5.7	0.7749		·27187 ·26194	25680	-25169	·24660	·24154	·23650	-23149	·22650	·22153
5.8	0.7313	·26710		·23080	·24017	-23539	·23064	·22591	·22120	·21650	·21183
5.9	0.6917	-25460	·24977	·24490 ·23127	·24017	·22238	·21795	·21355	·20916	·20478	-20042
6.0	0.6557	·24022	·23573	·23127	•22081	-22230	-21130	-21555	20010	20110	20012
				01011	01001	·20792	·20384	·19977	·19572	·19167	·18764
6-1	0.6227	·22436	·22023	·21611	·21201	·20792 ·19241	18868	·18496	·18125	·17755	·17385
6-2	0.5926	·20742	·20366	·19990	·19615		-17286	·16949	16612	.16276	-15940
6.3	0.5649	18984	·18643	$\cdot 18302$	-17963	·17624		15371	·15068	.14766	·14464
6.4	0.5394	·17198	$\cdot 16892$	·16587	·16282	·15978	·15674		·13527	·13257	12989
6-5	0.5158	·15422	·15150	·14879	·14608	·14337	·14066	·13796	.13921	.19701	-12909
1							30404	10055	10017	·11780	·11542
6.6	0.4940	·13689	·13450	·13210	$\cdot 12971$	$\cdot 12732$	·12494	·12255	12017	·10358	·10150
6.7	0.4739	·12026	·11817	·11608	·11399	·11191	·10982	·10774	10566		
6.8	0.4551	·10457	· 10276	$\cdot 10095$	·09914	·09734	·09553	:09373	·09193	·09013	·08833
6-9	0.4376	·08998	·08843	·08688	·08533	·08378	-08224	08069	·07914	·07760	·07606
7.0	0.4214	·07663	•07531	·07400	·07268	·07137	·07005	·06874	-06743	·06611	.06480
1	,										

•				•	Percen	tage nature	l mortality	7, C			
Y	Q/Z	41	42	43	44	45	46	47	48	49	50
7.1	0.4062	·06457	·06347	·06236	·06126	·06015	·05905	·05794	·05684	·05574	05463
7.2	0.3919	·05384	-05292	·05201	·05109	·05017	·04925	·04833	·04741	·04649	·04557
7.3	0.3786	·04443	·04367	.04291	.04215	·04140	·04064	·03988	·03913	03837	·03761
7.4	0.3661	.03267	·03565	·03504	·03442	·03380	·03318	·03257	.03195	.03133	$\cdot 03072$
7.5	0.3543	•02930	·02880	·02830	·02780	·02731	-02681	·02631	.02581	·02531	·02482
7.6	0.3432	·02342	.02302	·02262	·02222	·02183	·02143	·02103	·02063	.02023	·01984
7.7	0.3327	·01852	·01820	·01789	.01757	·01726	·01695	·01663	.01632	·01600	·01569
7.8	0.3228	·01449	·01424	·01400	·01375	·01351	·01326	·01301	·01277	01252	·01228
7.9	0.3134	.01122	·01103	·01084	01065	·01046	·01026	·01007	·00988	·00969	·00950
8.0	0.3046	•00859	·00845	·00830	·00815	· 0 0801	·00786	$\cdot 00772$	·00757	·007 43	-00728
8-1	0.2962	•00651	·00640	·00629	00618	·00607	·00596	·00585	·00574	·00563	·00552
8.2	0.2882	·00488	·00480	·00472	·00463	·00455	·00447	·00439	·00430	-00422	·00414
8.3	0.2806	.00362	·00356	-00350	·00344	·00338	·00332	·00325	·00319	·00313	·00307
8.4	0.2734	·00266	·00261	·00257	$\cdot 00252$	·00248	·00243	·00239	$\cdot 00234$	·00230	$\cdot 00225$
8.5	0.2666	·00193	·00190	·00187	·00183	·00180	·00177	•00174	-00170	·00167	·00164
8.6	0.2600	·00139 ·	·00136	·00134	·00132	·00129	·00127	·00125	·00122	·00120	·00118
8.7	0.2538	·00099	·00097	·00095	·00094	$\cdot 00092$	·00090	·00089	·00087	·00085	·00084
8.8	0.2478	.00070	·00068	·00067	.00066	·00065	·00064	$\cdot 00062$	·00061	·00060 '	·00059
8.9	0.2421	.00048	·00048	·00047	·00046	·00045	·00044	-00044	·00043	·00042	·00041
9.0	0.2367	·00033	·00033	$\cdot 00032$	·00032	·00031	·00031	·00030	·00029	.00029	·00028

TABLE II (cont.)

Y	Q/Z	51	52	53	54	55	56	57	58	59	60
1.8	419.1	·00001	•00001	·00001			-			-	
1.9	305.8	+00001	·00001	·00001	·00001	·00001	- 0 0001	·00001	·00001	·00001	·00001
2.0	225.3	.00002	·00002	$\cdot 00002$	$\cdot 00002$	-00002	·00002	·00001	·00001	·00001	·00001
2-1	167-69	.00003	·00003	·00003	·00003	·0000 3	•0000 3	·00003	·00003	·00002	·00002
2.2	126-02	-00006	·00006	·00006	·00005	·00005	·00005	·00005	·00005	·00004	·00004
2.3	95.63	-00010	·00010	·00010	·00009	·00009	·00009	·00008	·00008	·00008	·00007
-4	73.28	-00018	·00017	·00016	·00016	·00015	·00015	·00014	·00013	·00013	·00012
-5	56.70	.00030	.00028	.00027	·00026	·00025	·00024	·00023	·00022	·00021	·00021
9.6	44.288	.00048	·00046	·00045	·00043	·00041	·00039	·00038	·00036	·00035	·00034
.7	34.923	.00077	·00074	·00071	·00068	·00066	·00063	·00061	·00058	·00056	·00054
.8	27.797	.00121	·00116	·00112	•00107	·00103	•00099	·00095	•00091	·00088	·00084
.9	22.330	-00186	·00179	$\cdot 00172$	·00165	·00159	·00153	·00147	·00141	·00135	·00130
0	18-101	·00280	·00270	·00259	·00249	·00240	·00230	·00221	00213	·00204	·00196
1 1	14.802	·00414	·00399	·00383	·00369	·00354	·00341 .	·00327	·00314	·00302	· 0 0290
2	12.211	.00600	·00578	·00556	·00534	00514	·00494	·00475	·00456	·00438	00421
.3	10-159	.00853	·00821	.00790	.00760	·00731	·00703	·00676	·00649	·00624	·00599
-4	8.521	-01188	·01144	·01101	·01059	·01019	·00981	·00943	·00907	·00871	·00837
-5	7.205	·01623	·01563	·01505	·01449	·01395	·01342	•01291	·01242	·01194	·01147
•6	6·1394	·02174	·02095	·02018	·01944	01872	·01802	·01734	·01668	·01605	·01543
.7	5.2705	·02858	·02755	·02655	02559	·02465	·02374	·02286	·02200	·02117	·02036
.8	4.5571	.03686	.03556	·03429	.03306	03186	·03070	·02958	·02848	·02742	·02638
.9	3.9676	.04667	·04504	.04346	·04193	-04044	·03899	·03758	·03620	·03487	·03357
0	3.4770	+05802	05603	·05410	05222	·05040	·04862	·04689	·04520	·04356	·04196
.1	3.0665	•07084	·06847	·06615	·06390	·06170	·05956	·05748	·05545	·05346	·05153
.2	2.7206	-08500	·08221	·07949	·07684	07425	+07172	·06926	·06685	·06450	·06220
.3	2.4276	+10027	09705	.09391	·09084	.08785	·08492	·08206	.07926	·07652	·07384
.4	2.1780	-11634	.11270	·10914	·10565	·10224	·09890	.09563	·09244	·08930	·08623
-5	1.9640	13285	.12879	-12481	·12092	·11710	·11336	·10970	·10610	·10258	·09912
0 1	1.9040	10200	10010	101							• •

TABLE	II	(cont.)
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Percentage natural mortality, C

Y	Q Z	51	52	53	54	55	56	57	58	59	60
4.6	1.7797	·14936	.14491	·14055	·13627	·13207	·12794	·12390	·11992	+11602	·11218
4.7	1.6202	·16544	·16061	15592	$\cdot 15129$	·14673	·14225	·13785	$\cdot 13352$	12926	·12507
4.8	1.4814	·18062	$\cdot 17551$	·17049	·16534.	·16067	·15588	.15116	$\cdot 14652$	·14194	·13744
4.9	1.3599	·19447	·18911	18383	17863	·17350	·16844	·16346	15854	·15369	-14891
5-0	1.2533	•20658	·20104	19556	-19016	18482	17956	.17436	·16923	·16416	·15915
5-1	1.1593	·21663	·21096	·20335	·19981	19433	·18891	·18356	·17827	.17304	·16787
5-2	1.0759	·22434	·21860	$\cdot 21292$	·20730	·20175	·19624	19080	·18541	·18007	.17479
5.3	1.0018	·22951	. 22377	·21809	·21246	·20688	20135	19588	·19045	-18508	+17975
5.4	0.9357	·23204	·22636	·22074	·21516	·20962	20413	·19868	·19328	18792	·18261
5-5	0.8764	+23191	·22636	-22084	·21536	·20993	20453	·19917	19385	18856	·18332
5.6	0.8230	·22919	·22380	·21843	·21313	·20783	·20259	•19737	19218	·18703	·18191
ō·7	0.7749	•22401	·21884	$\cdot 21369$	·20858	·20349	·19843	·19340	·18839	·18341	·17846
5.8	0.7313	·21658	·21166	·20677	·20189	·19704	19221	18741	·18262	.17786	.17312
5-9	0.6917	·20717	·20253	·19791	·19331	·18873	·18417	·17962	·17509	17059	16609
8-0	0.6557	+19607	$\cdot 19174$	18742	·18312	·17884	17456	·17030	16606	16183	$\cdot 15762$
6-1	0.6227	.18362	·17961	·17362	·17163	·16766	·16370	·15974	·15580	·15188	·14796
6-2	0.5926	+17017	·16649	·16282	$\cdot 15917$	$\cdot 15552$	+15188	$\cdot 14825$	·14462	·14101	·13740
6-3	0.5649	·15606	·15272	$\cdot 14938$	·14606	+14274	·13943	·13612	$\cdot 13282$	12953	·12624
6-4	0.5394	·14163	-13862	-13562	·13262	·12963	$\cdot 12665$	·12366	·12069	+11771	+11475
6.5	0.5158	·12720	$\cdot 12452$	+12184	+11916	·11649	·11383	·11116	$\cdot 10850$	·10385	·10319
6-6	0.4940	+11305	·11068	·10831	·10595	·10339	·10123	•09887	·09652	·09417	·09182
6.7	0.4739	·09942	+09735	·09528	+09321	+09114	·08908	·08701	+08495	.08289	·0808 3
6-8	0.4551	+08653	+08473	08294	·08114	·07933	.07736	·07577	07398	07219	·07040
6.9	0.4376	+07451	07297	+07143	+06989	+06835	+06681	+06528	.06374	06220	-06067
7-0	0.4214	+06349	+06218	-06087	.02926	0.05826	·05693	·05564	·05434	·05303	+03172

7.1	0.4062	∣ ∙05353	05243	·05133	·05023	·04913	·04803	·04693	·04583	·04473	•
7.2	0.3919	•04466	·04374	·04282	·04190	·04099	·04007	·03916	·03824	·03732	•
7.3	0.3786	·03686	·03610	·03535	·03459	·03383	·03308	·03232	·03157	·03081	•
7.4	0.3661	•03010	$\cdot 02948$	·02887	$\cdot 02825$	·02763	$\cdot 02702$	·02640	$\cdot 02578$	·02517	•
7.5	0.3543	·02432	·02382	·02332	·0228 3	·02233	·02183	·02133	·0208 4	•02034 ·	•
7.6	0.3432	·01944	·01904	·01864	·01825	·01785	·01745	·01705	·01666	·01626	
7.7	0.3327	•01537	·01506	·01475	·01443	·01412	·01380	·01349	·01317	·01286	•
7.8	0.3228	·01203	·01179	·01154	·01129	·01105	·01080	·01056	·01031	·01006	•
7.9	0.3134	.00931	.00912	·00893	·00874	·00855	·00836	·00817 '	:00798	·00779	•
8.0	. 0.3046	+00713	•00699	·00684	·00670	·00655	·00641	·00626	·00611	·00597	•
8-1	0.2962	+00541	•00530	·00519	·00508	·00497	·00485	·00474	00463	·00452	
8.2	· 0·2882	.00405	·00397	·00389	·00381	$\cdot 00372$	·00364	·00356	·00348	·00339	
8.3	0.2806	·00301	·00295	·00289	·00282	·00276	-00270	·00264	·00258	·00252	
8.4	0.2734	.00221	·00216	$\cdot 00212$	·00207	·00203	·00198	·00194	-00189	·00185	•
8.5	0.2666	·00160	·00157	•00154	·00151	·00147	·00144	·00141	•00138	·00134	
8.6	0.2600	.00115	·00113	·00111	·00108	·00106	·00104	·00101	·00099	·00096	
8.7	0.2538	-00082	·00080	·00079	·00077	·00075	·00074	00072	·00070	·00069	
8.8	0.2478	-00058	·00057	00055	·00054	·00053	$\cdot 00052$	00051	·00050	·00048	
8.9	0.2421	.00040	·00039	·00039	·00038	·00037	·00036	·00035 ·	·00034	·00034	
9.0	0.2367	.00028	·00027	·00027	·00026	·00025	·00025	·00024	·00024	·00023	
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TABLE	Π	(cont.)
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Percentage natural mortality, C

Y	Q/Z	61	62	63	64	65	66	67	68	69	70
1·9 2·0	305·8 225·3	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001 ·00001	·00001	·00001
$2 \cdot 1$	167·69	·00002	·00002	•00002	·00002	•00002	•00002	·00002	•00002	•00002	•00002
$2 \cdot 2$	126·02	·00004	·00004	•00004	·00004	•00003	•00003	·00003	•00003	•00003	•00003
2·3	95·63	+00007	·00007	+00006	·00006	·00006	-00006	-00005	-00005	+00005	-00005
2·4	73·28	+00012	·00011	+00011	·00010	·00010	-00010	-00009	-00009	+00008	-00008
2·5	56·70	+00020	·00019	+00018	·00017	·00017	-00016	-00015	-00015	+00014	-00013
2·6	44·288	·00032	·00031	-00030	•00028	·00027	·00026	·00025	·00024	·00023	·00022
2·7	34·923	·00052	·00049	-00047	•00045	·00043	·00042	·00040	·00038	·00036	·00035
2∙8	27·797	·00081	·00078	·00074	·00071	·00068	·00065	·00062	·00060	·00057	·00054
2∙9	22·330	·00125	·00119	·00114	·00110	·00105	·00101	·00096	·00092	·00088	·00084
3∙0	18·101	·00188	·00180	·00173	·00166	·00159	·00152	·00145	·00139	·00133	·00127
3·1	14·802	·00278	•00267	·00256	·00245	·00235	·00225	·00215	•00206	·00197	·00188
3·2	12·211	·00404	•00388	·00372	·00356	·00342	·00327	·00313	•00299	·00286	·00273
3∙3 3∙4	10-159 8-521	·00575 ·00804	·00552 ·00772	+00530 +00741	-00508 -00710	·00487 ·00681	·00466 ·00652 ·00895	·00446 ·00624	·00427 ·00597 ·00820	·00408 ·00571 ·00784	+00389 +00545 +00749
3∙5 3∙6	7·205 6·1394	·01102 ·01483	·01058 ·01424	·01016 ·01367	·00975 ·01312	·00934 ·01258	-01206	·00857 ·01155	-01106	·01057	-01010
3·7	5·2705	·01958	·01881	·01807	·01735	·01664	·01595	·01529	·01463	·01400	·01338
3·8	4·5571	·02538	·02440	·02344	·02251	·02161	·02072	·01986	·01902	·01820	·01740
3·9	3·9676	·03230	·03107	·02987	·02870	·02755	·02644	·02535	·02429	·02325	·02224
4.0	3.4770	•04039	.03887	.03739	·03594	.03452	.03314	03179	·03047	·02919	·02793

4-1	3.0665	•04964	·04779	·04599	·04423	$\cdot 04251$	·04083	·03919	·03758	·03601	·03447	
4.2	2.7206	•05996	·05776	·05562	·05352	·05146	·04946	·04749	·04557	·04368	·04184	
4.3	2.4276	·07122	·06865	·06614	·06368	·06128	05892	·05661	·05434	·05212	·04995	
4.4	2.1780	.08323	·08028	·07739	·07456	·07178	·06906	·06639	·06377	·06120	·05867	
4.5	1.9640	.09572	·09240	·08913	·08592	-08277	·07968	·07664	·07366	• 07073	•06785	
10	2 0010											
4.6	1.7797	·10841	·10471	·10107	·09750	·09398	·09053	·08713	·08379	·08050	·07727	
4.7	1.6202	·12095	·11690	·11291	·10899	·10513	$\cdot 10132$	·09758	·09389	·09026	•08669	
4.8	1.4814	.13300	·12863	·12432	$\cdot 12007$	·11589	·11177	·10770	·10370	·09975	·09585	
4.9	1.3599	-14420	·13954	·13496	·13043	·12596	·12155	11720	·11291	·10867	·10449	
5.0	1.2533	-15421	·14933	·14451	·13975	·13504	·13039	·12580	·12126	·11678	·11234	
0.0	1.2099	10421	.14200	14451	10010	10002	10000	12000				
5.1	1.1593	.16275	·15769	·15269	·14775	·14285	·13802	·13323	·12850	·12381	·11918	
5.2	1.0759	.16957	16439	·15927	·15419	·14917	·14420	·13927	·13440	$\cdot 12957$	$\cdot 12478$	
5.3	1.0018	•17447	·16923	16405	-15891	·15381	·14876	·14375	·13879	·13387	·12899	
		•17733	·17210	·16691	·16176	·15665	·15158	·14655	·14156	-13661	·13169	
5.4	0.9357			·16780	·1627C	·15763	·15260	.14760	·14264	.13771	·13281	
5.5	0.8764	•17811	·17293	•10/80	·10270	•10703	15200	14100	14204	10111	10201	
5-6	0.8230	-17681	·17175	·16672	·16172	-15675	·15181	·14690	·14202	·13717	$\cdot 13235$	
5.7	0.7749	·17353	·16863	·16376	·15891	·15409	·14929	·14452	·13977	·13505	·13035	
5.8	0.7313	16840	·16371	·15903	.15438	·14975	·14514	·14055	·13598	·13143	·12690	
5.9	0.6917	·16162	.15716	.15273	·14831	·14390	·13952	$\cdot 13515$	·13080	·12646	·12214	•
6.0	0.6557	·15342	·14923	14506	·14090	·13676	·13263	·12851	$\cdot 12441$.12032	·11624	
0.0	0.0001	10342	14723	14000	14030	10070	10000	12001				
6-1	0.6227	·14405	·14016	·13628	·13240	·12854	·12469	·12085	·11702	·11321	·10940	
6.2	0.5926	·13381	·13022	·12664	$\cdot 12307$	·11951	·11595	·11241	$\cdot 10887$	$\cdot 10534$	$\cdot 10182$	
6.3	0.5649	.12296	·11969	·11642	·11316	·10991	·10666	$\cdot 10342$	·10018	·09696	·09374	
6.4	0.5394	.11179	·10883	·10588	·10293	.09999	·09705	.09412	·09119	-08827	·08535	
6.5	0.5158	-10055	.09790	·09526	·09262	:08999	.08736	.08473	.08211	-07948	·07687	
	0.0100	-10000	00100	00020	00202			00210	•			
6.6	0.4940	-08947	·08713	08479	.08245	·08012	-07779	·07546	·07313	·07080	·06848	
6.7	0.4739	.07878	.07672	·07467	-07262	·07057	.06852	·06648	·06443	·06239	·06035	
6.8	0.4551	.06862	. 06683	-06505	.06327	·06149	·05971	·05793	·05616	·05438	·05261	
6.9	0.4376	+05913	-05760	.05607	·05454	.05301	·05148	·04995	.04842	·04690	04537	
7.0	0.4214	.05042	·04912	-04781	-04651	.04521	·04390	-04260	·04130	·04000	·03870	287
	V.4414		·UT014	OTIOL	-OTOOT	VIUAL	V 300 V	01800				0

					2 01001	ntage natur	ar 11101 (mill)	, , 0			
Y	· Q/Z	61	62	63	64	65	66	67	68	69	70
7.1	0.4062	•04253	•04143	04033	03924	-03814	.03704	.03595	•03485	+03376	·03266
7.2	0.3919 .	·03549	·03458	03366	03275	03183	03092	.03001	·02909	02818	.02727
7.8	0.8786	.02930	.02855	·02780	02704	02629	02553	02478	02403	.02327	02252
7.4	0.8661	+02394	.02332	.02271	.02209	02147	.02086	.02024	·01963	-01901	·01840
7.5	0.8548	+01934	.01885,	01835	.01785	.01736	01686	01636	-01586	01537	-01487
7.6	0.3432	+01547	·01507	·01467	·01427	·01388	·01348	01308	.01269	.01229	·01189
7.7	0.8327	.01223	·01192	·01160	01129	+01098	-01066	.01035	·01003	00972	+00941
7.8	0.8228	-00957	.00933	·00908	00884	.00859	.00834	00810	.00785	00761	00736
7.9	0.8134	00741	.00722	.00703	.00684	.00665	.00646	00627	.00608	00589	+00570
8.0	0.8046	+00568	00553	00539	·00524	.00509	00495	00480	•00466	00451	-00437
8-1	0.2962	+00430	00419	.00408	00397	.00386	00375	.00364	·00353	00342	·00331
8.2	0.2882	·00323	·00314	.00306	·00298	.00290	00281	00273	.00265	00256	+00248
8.3	0.2806	.00239	00233	·00227	.00221	.00215	.00209	.00203	+00196	.00190	+00184
8-4	0.2734	.00176	+00171	00167	.00162	.00158	00153	.00149	·00144	·00140	+00135
8.5	0.2666	.00128	00124	.00121	·00118	.00115	00111	00108	00105	.00102	-00008
8.6	0.2600	.00092	·00089	·00087	-00085	.00082	.00080	·00078	·00075	·00073	-00071
8.7	0.2538	+00065	.00084	.00062	+00060	+00059	.00057	.00055	00054	.00052	-00050
8.8	0.2478	.00046	·00045	.00044	.00042	·00041	·00040	+00039	00038	+00037	-00035
8.9	0.2421	.00032	.00031	.00030	.00030	00029	-00028	.00027	00026	00025	+00035
9.0	0.2367	.00022	.00021	·00021	.00020	.00020	.00019	00019	00018	00018	+00017

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 TABLE II (cont.)

 Percentage natural mortality. C

	Y	Q Z	71	72	73	74	75	76	77	78	79	80
	2.0	225.3	•00001	•00001	·00001	·00001	•00001	·00001	·00001	·00001	·00001	_
4	2.1	167-69	.00001	·00001	·00001	·00001	·00001	-00001	·00001	·00001	·00001	·00001
•	2.2	126-02	·00003	$\cdot 00002$	·00002	·00002	·00002	·00002	·00002	$\cdot 00002$	·00002	·00002
	2.3	95.63	·00004	·00004	·00004	·00004	·00004	·0000 3	·00003	·0000 3	·0000 3	·00003
	2.4	73-28	·00008	·00007	·00007	·00007	·00006	·00006	·00006	·00005	.00002	·00005
	2.5	56.70	·0001 3	·00012	· 0 0011	·00011_	·00010	·00010	·00009	·00009	·00008	·00008
	2 ·6	44-288	.00021	·00020	-00019	. •00018	·00017	.00016	·00015	·00014	·0001 3	·00013
	2.7	34.923	·00033	·00031	·00030	·00028	·00027	·00026	·00024	·0002 3	·00022	·00020
	2.8	27.797	.00052	·00049	·00047	·00045	$\cdot 00042$	·00040	·00038	·00036	·00034	$\cdot 00032$
	2.9	22.330	.00080	·00076	$\cdot 00072$	·00069	·00065	$\cdot 00062$	·00059	·00055	·00052	·00049
	3.0	18-101	··00121	·00115	·00109	·00104	·00099	·00094	·00088	·00084	·00079	·00074
	3.1	14.802	·00179	·00170	·00162	·00154	·00146	·00139	·00131	·00124	·00117	·00110
	3.2	12.211	.00260	. 00248	·00236	·00224	00213	·00202	•00191	·00181	·00170	·00160
	3.3	10.159	·00371	·00354	·00337	·00320	·00304	$\cdot 00288$	·00273	00258	·00243	·00229
	3.4	8.521	00520	·00496	$\cdot 00472$	·00449	·00426	·00404	·00383	.00362	·00341	·00321
	3.5	7.205	.00715	·00681	·00649	·00617	·00586	·00556	-00526	·00498	·00469	·00442
	3.6	6.1394	·0096 4	·00920	·00876	·00833	·00792	·00751	·00711	·00673	·00635	·00598
	3.7	5.2705	·01278	·01219	-01161	·01105	-01050	·00996	·00944	·00893	·00843	·00794
	3.8	4.5571	·01662	·01586	.01512	·01439	·01368	·01298	·01231	·01164	·01099	·01035
	3.9	3.9676	·02125	·02028	· •01934	01841	·01751	·01663	·01576	·01492	·01409	$\cdot 01328$
	4 ·0	3.4770	02669	·02549	·02431	·02316	·0220 3	·02093	·01985	·01879	·01775	·01673
	4 ·1	3.0665	.03296	·03149	·03005	-02864	-02725	·02590	02457	·02327	·02199	·02074
	4.2	2.7206	·04003	·03826	-03652	03482	·03315	·03152	·02991	·02834	·02680	·02528
	4-3	2.4276	·04781	·04572	-04367	·04165	·03968	·03774	·03583	·03396	·0321 3	·03032
5	4.4	2.1780	.05620	·05377	·05138	04903	04673	·04446	·04224	·04006	·03791	·03580
	4.5	1.9640	·06502	06224	·05951	·05682	·05418	·05158	·04903	·04651	·04404	·04161

t.)
t.)

					Percer	ntage natur	al mortalit	у, С			
Y	Q/Z	71	72	73	74	75	76	77	78	79	80
4.6	1.7797	•07409	.07096	-06788	.06485	·06187	+05893	.05604	·05319	•05039	•04763
4.7	1.6202	+08317	·07970	·07629	·07292	·06960	·06633	·06311	·05994	·05681	+05372
4.8	1.4814	•09201	08822	·08449	·08081	·07717	·07359	·07005	·06656	·06311	+05971
4.9	1.8599	10036	·09628	·09226	·08828	·08436	·08048	.07665	.07287	.06913	-06544
5 ∙0	1.2533	·10796	·10364	·09936	•09513	•09095	·08681	·08272	·07868	07469	•07074
5.1	1-1593	·11459	-11006	·10557	-10118	·09673	09238	·08808	·08382	·07960	•07543
5.2	1.0759	·12005	-11535	.11071	.10610	·10154	.09702	09255	.08811	.08372	.07937
5.8	1.0018	.12416	.11936	11461	·10990	·10522	·10059	.09599	·09144	.08692	.08244
5.4	0.9357	·12681	12197	.11717	·11240	·10767	·10298	.09832	.09369	08910	·08454
5.5	0.8764	.12795	12312	·11833	·11356	·10883	·10413	.09946	09482	09021	·08563
5.6	0.8230	.12756	12279	·11806	·11335	.10867	·10402	•09939	·09479	.09022	·08567
5.7	0.7749	12568	·12103	11640	·11180	·10723	·10267	·09814	.09364	08915	08469
5.8	0.7313	.12240	-11791	.11344	·10900	· 10457	·10016	:09578	09141	·08706	.08273
5-9	0.6917	·11784	11356	·10929	·10504	·10080	.09659	.09238	08820	08403	.07987
6.0	0.6557	+11218	10813	·10410	·10008	.09607	.09208	08809	08412	·08017	·07623
6-1	0.6227	10560	·10182	·0980 4	·09428	·09053	·08678	•08305	·07933	.07562	·07192
6-2	0.5926	+09831	·09481	·09132	·08783	08435	.08088	.07742	.07397	.07052	-06708
6.3	0.5649	.09052	.08731	·08411	·08092	07773	.07454	.07137	.06820	06503	-06187
6-4	0.5394	·08244	·07953	.07662	.07372	07083	·06794	.06506	06218	.05930	·05643
6.5	0.5158	•07426	·07165	·0690 4	·06644	·06384	·06124	·05865	·05606	·05348	•05090
6.6	0.4940	.06616	.06384	·06153	·05922	•05691	05460	·05230	·05000	.04770	·04540
6.7	0.4739	•05831	.05628	·05424	·05221	05018	·04815	·04612	.04410	.04207	.04005
6.8	0.4551	.05084	·04907	·04730	·04553	.04376	.04200	·04023	·03847	03671	03495
6-9	0.4376	·04384	.04232	·04080	.03927	+03775	·03623	·03471	.03319	-03168	·03016
7.0	0.4214	·03741	.03611	·03481	·03351	.03222	03092	.02963	.02833	·02704	+02574

	7-1	0.4062	·03157	·03047	·02938	·02829	·02719	·02610	·02501	·02392	·02283	·02174
	7.2	0.3919	·02636	·02544	·02453	$\cdot 02362$	·02271	·02180	·02089	·01997	·01906	·01815
	7.3	0.3786	·02177	·02101	·02026	·01951	·01876	·01800	·01725	·01650	·01575	·01500
	7.4	0.3661	·01778	·01717	·01655	·01594	·01533	•01471	·01410	·01348	·01287	·01226
	7.5	0.3543	01437	·01388	·01338	01289	•01239	·01189	·01140	•01090	·01040	·00991
	7.6	0.3432	·01149	·01110	·01070	·01030	·00991	·00951	·00911	·00872	00832	·00792
	7.7	0.3327	·00909	·00878	·00846	·00815	·00784	·00752	·00721	·00690	·00658	·00627
	7.8	0.3228	·00712	·00687	·00663	·00638	·00613	·00589	·00564	·00540	.00515	·00491
	7.9	0.3134	.00551	$\cdot 00532$	·0051 3	·00494	·00475	·00456	·00437	·00418	·00399	·00380
	8-0	0.3046	$\cdot 00422$	·00408	·00393	·00378	·00364	·00349	·00335	·00320	·00306	·00291
	8-1	0.2962	·00320	·00309	·00298	·00287	·00276	·00265	·00254	·00243	·00232	·00221
	8.2	0.2882	·00240	·00232	·00223	.00215	·00207	·00199	·00190	$\cdot 00182$	·00174	·00165
	8.3	0.2806	00178	00172	·00166	·00160	·00153	·00147	·00141	·00135	·00129	·00123
	8.4	0.2734	·00131	-00126	·00122	·00117	·00113	·00108	·00104	·00099	-00095	·00090
	8.5	0.2666	·00095	·00092	·00088	·00085	·00082	·00079	·00075	·00072	00069	·00065
	8.6	0.2600	·00068	·00066	·00064	00061	00059	•00056 ·	00054	·00052	·00049	·00047
	8.7	0.2538	00048	·00047	·00045	·00043	$\cdot 00042$	·00040	·00038	·00037	·00035	·00033
	8-8	0.2478	-00034	·00033	·00032	·00031	·00029	·00028	·00027	·00026	·00025	·00024
	8-9	0.2421	·00024	·00023	-00022	·00021	·00021	·00020	·00019	·00018	·00017	·00016
	9.0	0.2367	•0001 6	·00016	·00015	·00015	·00014	·00014	·00013	·00012	·00012	· •00011
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19-2			. A.									
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		•			Percer	ntage natur	al mortalit	у, С			
Y	Q/Z .	81	82	83	84	85	86	87	88	89	90
2.1	167-69	•00001	·00001	·00001	·00001	·00001	·00001	.00001	_	_	
2.2	126.02	·00001	·00001	·00001	·00001	·00001	·00001	-00001	·00001	·00001	·00001
2.3	95.63	•00003	·00002	·00002	·00002	·00002	-00002	·00002	·00001	·00001	·00001
2.4	73.28	·00004	·00004	·00004	·00004	·00003	·00003	·00003	·00003	·00002	.00002
2.5	56.70	-00007	·00007	•00006	·00006	·00005	·00005	·00005	·00004	·00004	·00003
2.6	44 ·288	.00012	·00011	·00010	·00010	•00009	-00008	·00008	·00007	·00006	•00006
2.7	34.923	•00019	·00018	·00017	·00015	·00014	·00013	·00012	·00011	·00010	•00009
2.8	27.797	-00030	·00028	·00026	·00024	·00022	·00021	·00019	·00017	·00016	·00014
2.9	22.330	·00046	·00043	·00040	·00037	·00035	-00032	·00029	·00027	·00024	·00022
3.0	18.101	•00070	•00065	·00061	·00057	$\cdot 00052$	·00048	·00044	-00041	·00037	·00033
3.1	14.802	·0010 3	•00097 [′]	·00090	·00084	·00078	·00072	·00066	•00060	·00055	·00049
3.2	12.211	·00150	·00141	·00131	.00122	·00113	·00105	·00096	·00088	·00080	·00072
3.3	10.159	·00215	.00201	·00188	·00175	.00162	.00150	-00137	.00125	·00114	·00102
3.4	8.521	·00301	·00282	·00264	00245	·00228	·00210	·00193	·00176	·00160	·00144
3 ·5	7.205	·00 4 15	·00389	·00363	·00338	·00314	·00289	·00266	·00243	.00220	·00198
3.6	6.1394	•00561	·00526	·00491	·00457	·00424	·00392	·00360	·00329	·00298	·00269
3.7	5.2705	.00746	·00699	·00653	·00608	·00564	·00521	·00479	·00438	·00397	·00357
3.8	4.5371	·00973	.00912	·00853	·00794	·00737	·00681	·00626	$\cdot 00572$	·00519	-00467
3.9	3.9676	-01248	·01170	·01094	·01020	·00946	-00875	00804	·00735	·00667	·00601
4 ·0	3.4770	•01574	·01476	·01380	·01287	·01195	·01104	·01016	·00929	·00844	·00760
4.1	3 ·0665	•01951	·01831	·01713	·01597	·01483	·01371	·01262	·01154	·01049	·00945
4.2	2.7206	.02379	$\cdot 02233$.02090	·01950	·01811	·01676	.01542	·01411	.01282	·01156
4.3	$2 \cdot 4276$	·02855	·02681	.02510	02342	-02177	.02015	·01855	·01698	·01544	·01392
4.4	2.1780	.03372	·03168	·02967	·02770	·02575	·02384	.02196	·02011	-01829	·01650
4.5	1.9640	·03921	·03685	·03453	03225	·03000	0.02779	02561	·02346	.02134	01926

TABLE II (cont.)

4.6	1.7797	ı •04491	·04223	·03959	.03699	$\cdot 03442$	·03190	·02941	·02695	·02453	·02214	
4.7	1.6202	-05068	·04767	$\cdot 04472$	-04180	·03892	·03608	03328	·03051	·02778	.02509	
4.8	1.4814	·05636	·05305	·04978	·04655	·04337	·04022	·03711	·03404	·03101	·02802	
4.9	1.3599	.06180	.05820	·05464	$\cdot 05112$	·04764	•04421	·04081	·03745	·03414	·03085	
5.0	1.2533	-06683	·06296	·05914	·05536	05162	·04792	·04426	·04064	·03705	·03351	
5-1	1.1593	.07129	·06720	·06315	·05914	·05517	·05124	·04735	·04349	·03967	·03589	
5.2	1.0759	.07505	-07078	·06654	.06235	·05819	·05407	. •04998	·04593	·04192	·03794	
5.3	1.0018	07799	.07358	·06921	·06488	·06057	.05631	·05208	·04788	·04371	·03958	
5.4	0.9357	-08002	.07553	·07107	06665	·06226	·05790	·05357	·04927	·04500	·04076	
5.5	0.8764	-08108	.07656	07208	.06762	·06318	·05878	·05441	·05006	·04574	·04145	
	0.8230	-08116	-07666	·07219	.06775	·06334	·05895	·05458	·05024	·04592	·04163	
5.6	0.8230	-08025	07584	.07144	.06707	.06272	05839	·05409	·04980	·04554	·04130	
5.7	0.7313	.07842	.07413	·06986	•06560	.06137	·05715	.05295	.04877	·04461	·04047	
5.8		.07573	07161	·06751	.06341	.05934	·05528	05123	·04720	·04319	·03919	
5.9	0.6917		06838	·06448	•06058	-05671	·05284	·04899	·04514	·04132	·03750	
6.0	0.6557	•07230	.00939	.00440	.00030	-03071	00201					
6-1	0-6227	.06823	·06454	·06087	·03721	·05356	$\cdot 04992$.04629	$\cdot 04267$	·03906	·03546	
6.2	0.5926	+06365	·06023	-05682	05342	$\cdot 05002$	• ·04663	·04325	·03987	·03651	.03315	
6.3	0.5649	.05872	·05558	·05244	·04930	·04618	·04306	·03994	·03683	·03373	·03063	
6.4	0.5394	05357	·05071	·04785	·04500	·04215	·03931	·03647	·03364	·03081	·02799	
6.5	0.5158	·04832	·04575	·04318	-04061	-03805	·03548	·03293	·03037	·02782	·02528	
6.6	0.4940	.04311	-04082	·03853	·03624	·03396	·03168	·02940	·02712	·02485	·02258	
6.7	0.4739	-03803	·03601	·03400	.03198	·02997	02796	·02595	·02395	·02194	·01994	
6.8	0.4551	.03319	·03143	·02967	.02792	·02616	.02441	·02266	.02091	·01916	·01741	
	0.4376	+02864	02713	.02561	.02410	·02259	02107	·01956	·01805	·01654	.01504	
6-9		-02445	·02316	02187	·02058	·01929	·01800	-01671	·01542	·01413	01284	
7.0	0.4214	·02445	.02310	-02107	02000							
7.1	0.4062	·02065	01956	-01847	01738	·01629	.01520	·01411	$\cdot 01302$	·01194	·01085	
7.2	0.3919	.01724	·01633	.01542	01451	.01361	.01270	-01179	·01088	·00997	·00906	
7.3	0.3786	·01425	01349	·01274	·01199	·01124	·01049	·00974	-00899	·00824	·00749	
7.4	0.3661	·01164	·01103	·01041	·00980	·00919	·00857	·00796	·00735	·00674	.00612	
7.5	0.3543	.00941	.00892	·00842	.00792	·00743	·00693	·00644	·00594	·00545	00495	č
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					Percer	itage natul	ai mortant	y, C			
Y	Q/Z	81	82	83	84	85	86	87	88	89	90
7.6	0.3432	-00753	·00713	·00673	·00634	·00594	·00555	·00515	·00475	·00436	•00396
7.7	0.3327	·00596	·00564	·00533	.00501	·00470	·00439	·00407	·00376	·00345	·00313
7.8	0.3228	·00466	.00442	·00417	·00393	·00368	·00343	·00319	·00294	-00270	·00245
7.9	0.3134	·00361	·00342	·00323	·00304	.00285	-00266	.00247	·00228	.00209	·00190
8.0	0.3046	.00277	·00262	-00247	·00233	-00218	·00204	•00189	·00174	-00160	·00146
8-1	0.2962	.00210	·00199	·00188	·00176	·00165	·00154	00143	·00132	·00121	·00110
8.2	0.2882	.00157	·00149	·00141	·00132	00124	·00116	·00108	·00099	·00091	·00083
8.3	0.2806	.00117	·00110	·00104	·00098	·00092	.00086	.00080	·00074	·00068	·00061
8.4	0.2734	·00086	.00081	·00077	·00072	·00068	·00063	·00059	·00054	·00050	-00045
8.5	0.2666	·00062	·00059	.00056	.00052	·00049	·00046	00043	·00039	·00036	·00033
8.6	0.2600	00045	·00042	·00040	·00038	.00035	·00033	·00031	·00028	·00026	·00024
8.7	0.2538	·00032	·00030	.00028	.00027	.00025	·00023	·00022	·00020	·00018	.00017
8.8	0.2478	·00022	.00021	·00020	.00019	-00018	·00017	+00015	·00014	·00013	·00012
8.9	0.2421	·00016	·00015	·00014	·00013	.00012	·00011	·00011	·00010	-00009	.00008
9.0	0.2367	-00011	·00010	00010	.00009	.00008	.00008	-00007	.00007	-00006	-00006

TABLE II (cont.)

Percentage natural mortality, C

TABLE III. Maxim	um and Minimum	Working	Probits and	l Range
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	orking probits	1 1	Maximum wor	
Expected		Banga		Expected
probit Y	$y_0 = Y - P/Z$	Range 1/Z	$y_{100} = Y + Q/Z$	probit Y
1.1	0.8579	5034	9.1421	8.9
1-2	0.9522	3425	9-0478	8-8
1.3	1.0462	2354	8.9538	8.7
1.4	1.1400	1634	8-8600	8-6
1.5	1.2334	1146	8.7666	8∙5
1.6	1.3266	811-5	8.6734	8.4
1.7	1.4194	580-5	8.5806	. 8.3
1.8	1.5118	419·4 306·1	8·4882 8·3962	8·2 8·1
1-9 2-0	1·6038 1·6954	225.6	8.3046	8.0
2.1	1.7866	168-00	8 2134	7.9
2.2	1.8772	126-34	8.1228	- 7.8
2.3	1.9673	95-96	8-0327	7.7
2.4	2.0568	73.62	7.9432 •	7.6
2.5	2.1457	57.05	7.8543	7.5
2.6	2.2339	44.654	7.7661	7.4
2.7	2.3214	35-302	7.6786	7.3
2.8	2.4081	28.189	7.5919	7.2
2.9	2.4938	22.736	7.5062	7.1
3-0	2.5786	18.5216	7-4214	7-0
3.1	2-6624	15-2402	7.3376	6-9
3.2	2.7449	12.6662	7.2551	6-8
3.3	$2 \cdot 8261$	10.6327	7.1739	6.7
3.4	2.9060	9-0154	7-0940	6-6
3.5	2.9842	7.7210	7.0158	6.5
3.6	3.0606	6-6788	6.9394	6.4
3.7	3.1351	5.8354	6-8649	6.3
3.8	3.2074	5.1497	6.7926	6.2
3∙9 4•0	3·2773 3·3443	4·5903 4·1327	6·7227 6·6557	6·1 6·0
4.1				
4.2	3.4083	3.7582	6.5917	5.9
4.3	3∙4687 3•5251	3·4519 3·2025	6.5313	5·8 5·7
4.4	3\5770	3.0010	6·4749 6·4230	5.6
4.5	3-6236	2.8404	6.3764	5.5
4-6	3.6643	2.7154	6.3357	5.4
4-7	3-6982	2.6220	6.3018	5.3
4.8	3.7241	2.5573	6-2759	5.2
4-9	3.7407	2.5192	6.2593	5.1
5.0	3.7467	2.5066	6.2533	5.0
5.1	3 ·7401	2.5192	6-2599	4.9
5.2	3.7186	2.5573	6-2814	4.8
5.3	3.6798	2.6220	6.3202	4.7
5.4	3-6203	2.7154	6.3797	4 ·6
5.2	3.5360	2.8404	6·4640	4.5
5-6	3.4220	3-0010	6.5780	4.4
5.7	3 ·2724	3.2025	6.7276	4.3
5.8	3.0794	3.4519	6.9206	4.2
5.9	2.8335	3.7582	7.1665	4.1
6-0	2.5230	4.1327	7.4770	4 ∙0
6.1	2.1324	4-5903	7-8676	3.9
6·2	1.6429	5.1497	8.3571	3.8
6·3	1-0295	5.8354	8-9705	3.7
6.4	0.2606	6.6788	9 ·7394	3-6

The working probit, y, may be obtained as y = (Y - P/Z) + p/Z or y = (Y + Q/Z) - q/Z, whichever is the more convenient, where p(-1-q) is the observed proportion killed.

TABLE IV. Working Probits

(Y = 2-0-2-9; 0-50 % kill)

%				Ex	pected pr	obi t, P				
kill	2-0	2.1	2.2	2.3	2.4	2.5	2-6	2.7	2.8	2.9
0	1-695	1.787	1.877	1.967	2.057	2.146	2.234	2.321		
v	1.020	1.101	1.011	1.901	4001	2.140	4.794	Z·321	2.408	2.494
1	3-951	3-467	3.141	2.927	2 ·793	2.716	2-681	2-674	2.690	2 ·721
2	6-207	5.147	4-404	3-886	3.529	3.287	3.127	3-027	-972	-949
3	8-463	6-827	5-667	4-846	4-265	-857	-574	-380	3.254	3.176
4	_	8.507	6-931	5-806	5.002	4.428	4-020	-733	-536	-403
5		_	8.194	6.765	-738	-998	-467	4-086	-818	-631
				-						
6	·	—	9-458	7.725	6-474	5 569	4.913	4.440	4-099	3·858
7	_		—	8-684	7.210	6-139	5-360	-793	·381	4-085
8	—	_		9-644	-946	·710	-806	5.146	-663	·313
9	—	—	—	—	8-683	7.280	6.253	- 4 9 9	-945	-540
10			-		9-419	·851	-699	-852	5.227	•767
11		_	—	_	_	8-421	7.146	6.205	5.509	4-995
12	_		_			-992	-592	-558	-791	5.222
13		—		_	. —	9.562	8-039	-911	6-073	•449
14	_		—	-			-486	7.264	-355	-677
15	-	—	—	-	—		·932	-617	-636	·904
16	_	_	_		_		9 ·379	7.970	6-918	6-132
17				_		_	•825	8.323	7.200	-359
18	_		_	_			-020	-676	-482	-586
19	_	_	_	_		_	_	9-029	-462	-814
20	_	_	· <u>-</u>	Ξ		_	_	-382	8-046	7-041
20								-004	0.040	1.041
21	—		_	_	—	_	—	9.735	8-328	7.268
22	—							—	·610	-496
23		_	—	—		—		_	·892	·723
24	—				—				9 ·173	-950
25	—		—	—	—		—		-455	8.178
26	—	—		—		_		_	9 ·737	8-405
27	<u> </u>		_							-633
28		_		—			—	_		-860
29	—	_	—	_		—		_		9-087
30		-	-		—	_		 .	—	·315
31		_		_	_		_			9.542
32		_				_				.769
33	_	_					_	_		-997
34	•	_	_		_	_		_	_	
35		_	_		_				_	_
			_			-			_	

296

(Y = 3.0-3.9; 0-50% kill)

%				Exp	ected pro	obit, P				
kill	3-0	3.1	3.2	3.3	3.4	3.5	3.6	3.7	3.8	3.9
0	2.579	2.662	2.745	2.826	2.906	2.984	3.061	3.135	3.207	3·277
		0.015	0 070	0.020	9 002	3-061	3.127	3 ·193	3 ·259	3.323
1	2.764	2.815	2.872	2·932 3·039	2·996 3·086	•139	·194	-252 -252	·310	•369
23	•949 3∙134	.∙967 3·120	-998 3·125	·145	·176	-216	-261	-252	·362	-415
4	·319	·272	252	-251	-267	·293	-328	-369	-413	-461
5	-505	•424	·378	-358	-357	·370	-395	-427	-465	-507
6	3-690	3.577	3.505	3.464	3.447	3.447	3.461	3.485	3 ·516	3.553
ž	-875	-729	-632	-570	-537	·525	-528	-544	-568	·599
8	4-060	·882	•758	-677	·627	-602	•595	-602	-619	-645
9	-246	4.034	-885	-783	-717	-679	•662	-660	-671	-690 -736
10	•431	•186	4-012	-889	-808	-756	•728	•719	-722	-
11	4.616	4.339	4.138	3.996	3 ·898	3.834	3 ·795	3.777	3.774	3.782
12	-801	•491	-265	4.102	-988	-911	-862	·835	•825 •877 -	-828 -874
13	-986	-644	·391 ·518	-208 -315	4-078 -168	-988 4-065	-929 -996	-894 -952	-928	-920
14 15	5·172 ·357	•796 •948	·518 ·645	·315 ·421	·108 ·258	·142	4.062	4.010	·980	-966
19	100	-940								
16	5.542	5.101	4.771	4.527	4.348	4.220	4.129	4.069	4-031	4-012
17	.727	·253	-898	-634	-439	·297	·196	·127	-083	-058
18	-913	•406	5-025	·740	•529	-374	•263 •330	·185	·134 ·186	·104 ·149
19 20	6-098 -283	-558 -710	·151 ·278	•846 •953	·619 ·709	·451 ·528	·396	·244 ·302	-237	·149
21	6-468	5.863	5.405	5-059	4 ·799	4-606	4.463	4.361	4.289	4.241
22	-653	6.015	-531	·165	·889	-683	-530	-419	·340	-287
23	-839	·168	-658	·272	-979	•760	•597	-477	·392 ·443	·333 ·379
24	7-024 •209	·320	·785	·378 ·484	5·070 160	-837 -914	•664 •730 -	•536 •594	•445 •495	-425
25		-472	•911							
26	7.394	6 ·625	6.038	5.591	5.250	4.992	4-797	4.652	4.546	4.471
27	·580	·777	·165	-697	·340	5-069	·864	.711	-598	·517
28	-765	-930	-291	-803	·430	·146 ·223	•931 •997	·769	·649 ·701	-563 -608
29 30	-950 8-135	7∙082 ∙234	•418 •545	·910 6·016	-520 -610	·225 ·300	5·064	-827 -886	•752	-654
31	8.320	7.387	6.671	6.122	5.701	5.378	5.131	4.944	4.804	4.700
32	-506	-539	•798	·229	-791	·455	·198	5-002	·855	·746
33 34	-691 -876	-692 -844	-925 7-051	·335 ·441	-881 -971	•532 •609	•265 •331	-061 -119	-907 -958	-792 -838
34 35	9-061	-996	-178	-548		-687	-398	·119	5-010	-884
					0 001					
36	9.247	8.149	7.305	6.654	6.151	.5.764	5.465	5.236	5-061	4.930
37 38	-432 -617	·301 ·454	-431 -558	-760 -867	·242 ·332	-841 -918	•532 •599	·294 ·353	·113 ·164	-976 5-022
39	-802	-606	•685	-973	·332 ·422	-995	-665	•411	·216	-068
40	-987	-758	-811	7.079	•512	6-073	.732	469	-267	·113
							-		-	
41	_	8-911	7.938	7.186	6-602	6.150	5.799	5.528	5.319	5.159
42 43		9-063 -216	8-065 -191	·292 ·398	-692 -782	·227 ·304	•866 •932	•586 •644	·370 ·422	·205 ·251
45 44	_	-210	-318	•505	-182	·304 ·381	·932 ·999	•044 •703	•422 •473	-297
45	_	-520	-445	-611	-963	-459	6.066	-761	-525	-343
46	_	9-673	8.571	7.717	7-053	6.536	6 ·133	5 ·819	5 ·576	5.389
47	-	·825	-698	-824	.143	-613	·200	-878	-628	435
48	—	-978	-825	-930	·233	-690	·266	-936	·679	-481
49		—	-951	8-036	·323	-767	·333	-994	·731	·527
50	-	—	9-078	-143	-414	-845	-400	6-053	•782	•572
				•					•	

(X = 4-0-4-9; 0-50% kill)

%				Ext	ected pr	obit, Y				
kill	4-0	4.1	4.2	4·3	4.4	4.5	4-6	4.7	4-8	4.9
0	3.344	3-408	3.469	3.525	3.577	3-624	3.664	3.698	3.724	3.741
1	3-386	3-446	3.503	3.557	3-607	3-652	3-691	3.724	3 ·750	3.766
2	-427	-487	-538	-589	-637	-680	-719	•751	.175	-791
3	-468	·521	·572	-621	-667	-709	·746	.777	-801	-816
4	-510	-559	-607	-653	-697	-737	-773	-803	-826	-841
5	•551	-596	-641	-685	-727	•766	-800	-829	·852	-867
6	3.592	3-634	3-676	3.717	3.757	3.794	3.827	3-856	3-878	3-892
7	-634	-671	-710	•749	-787	·822	-854	-882	-903	-917
8	-675	•709	·745	•781	-817	·851	-882 -909	•908 •934	-929 -954	-942 -967
9 10	-716 -758	-747	•779	•813 •845	-847 -877	-879 -908	-936	-954 -960	-980	-993
		·784	-814	•						
11	3.799	3.822	3.848	3.877	3.907	3 ·936	3.963	3.987	4-005	4-018
12	·840	-859	-883	-909	-937	·964	-990	4-013	-031 -057	-043 -068
13 14	-882 -923	·897 ·934	-917 -952	-941 -973	•967 •997	-993 4-021	4-017 -044	-039 -065	-082	-093
14	-925 -964	•934 •972	·952 ·986	4-005	4-027	-050	-072	-092	-108	·119
16	4.006	4-010	4-021	4.038	4-057	4-078	4-099	4.118	4.133	4.144
17	-047	-047	-056	-070	-087	•106	·126	·144	-159	·169 ·194
18	-088	-085	-090	-102	-117	·135 ·163	·153 ·180	·170	·184 ·210	-219
19	-130 -171	-122 -160	•125 _ •159	-134 -166	•147 •177	·105	207	-196 -223	-236	·245
20										
21	4.212	4-198	4 ·194	4 ·198	4.207	4.220	4.235	4.249	4.261	4-270
22	-253	·235	·228	·230	-237	·248	·262	-275	-287 -312	·295 ·320
23	-295	·273	·263	•262 •294	-267 -297	-277 -305	·289 ·316	•301 •327	·314	-345
24 25	-336 -377	·310 ·348	•297 •332	-326	·297 ·327	•305 •334	-343	·354	•363	·370
26	4-419	4.385	4.366	4.358	4.357	4.362	4.370	4.380	4.389	4·396 ·421
27	-460	-423	-401	·390	-387	-391	·397 ·425	-406 -432	-415 -440	•421 •446
28	-501	-461	-435 -470	-422 -454	-417 -447	-419 -447	-420 -452	•452 •459	-466	-471
29 30	-543 -584	498 536	•470 •504	-486	-477	-476	-479	-485	-491	-496
31	4-625	4.573	4.539	4.518	4 507	4.504	4.506	4.511	4.517	4·522 •547
32	667	-611	·573	·550	-537	•533	-533	•537 •563	•542 •568	•041 •572
33	·708	-649 690	-608	-582 -614	∙567 •597	-561 -589	•560 •588	-590	·508	-597
34 35	·749 ·791	-686 -724	-642 -677	-646	627	-618	-615	-616	-619	622
36	4.832	4.761	4.711	4.678	4-657	4.646	4.642	4-642	4.645	4·648 -673
37	·873	•799	-746	-710	-687	-675	-669 -696	-668 -695	-670 -696	-698
38	·915	·836	•780 •815	·742 ·774	-717 -747	·703 ·731	-723	.721	.721	-723
39 40	-956 -997	·874 ·912	-849	-806	-777	.760	750	747	-747	.748
								-		
41	5-039	4.949	4.884	4.838	4.807	4.788	4.778	4.773	4·773 ·798	4·774 ·799
42	-080	-987	-918	-870	·837	-817	•805 •832	•79 9 •826	-196	-824
43	·121 ·163	5·024 ·062	-953 -988	-902 -934	-867 -897	-845 -873	·852 ·859	•820 •852	•849	-849
44	·105 •204	-899	5.022	-966	-927	-902	-886	-878	·875	·874
45										
46	5.245	5.137	5-057	4.998	4.957	4.930	4.913	4.904	4.900	4.900
47	-287	-175	-091	5-030	-987	-959	-941	·931	-926 -952	-925 -950
48	·328	-212	-126	-062	5-017	·987	-968 -995	-957 -983	•952 •977	-975
49	·369 -411	·250 ·287	·160 ·195	-094 -126	-047 -078	5-015 -044	-995 5-022	-965 5-009	5-003	5 000
50	* £11 .	-201	-199	-120	-010	111	0 944	0.000	0.000	

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(Y = 5-0-5-9; 0-50 % kill)

•				(_ ···	e ee, e		.			
%		~ .		-	ected pro	-	5.6	5.7	5.8	5.9
kill	5-0	5.1	5.2	5.3	5.4	5.5	•••			
0	3.747	3.740	3.719	3.680	3.620	3.536	3-422	3.272	3.079	2.834
1	3.772	3.765	3.744	3.706	3.647	3.564	3.452	3.304	3.114	2.871
2	.797	-790	·770	-732	-675	-593	-482	•336	·148	•909
3 4	-822 -847	·816 ·841	-795 -821	•758 •785	•70 2 •729	-621 -650	•512 •542	-368 -400	·183 ·217	-946 -984
5	-872	-866	-846	-811	-756	-678	-572	-433	-252	3-021
6	3.897	3.891	3.872	3·837 •863	3.783	3.706	3-602 -632	3-465 -497	3·287 •321	3-059 -097
7 8	-922 -947	·916 ·942	·898 ·923	•890	-810 -838	•735 •763	•662	-529	-356	-134
ğ	-972	-967	-949	-916	•865	.792	-692	-561	•390	·172
10	-997	-992	-974	-942	-892	·820	-722	•593	-425	-209
11	4-022	4-017	4-000	3-968	3 ·919	3.848	3.752	3.625	3.459	3.247
12	-047	-042	-025	•994	•946	.877	.782	-657	-494	-284
13	-073	-068	-051	4-021	-973	·905	-812	-689	·528	·322
14	-098	•093	-077	-047	4.000	·934	-842	-721	-563	-360
15	-123	•118	•102	-073	-028	-962	-872	-753	-597	-397
16	4.148	4.143	4.128	4.099	4.055	3.990	3.902	3 ·785	3.632	3.435
17	-173	·168	153	·126`	-082	4.019	.932	-817	-666	-472
18 19	·198 ·223	·194 ·219	•179 •204	·152	-109	-047	-962 -992	-849	.701	-510
20	-248	-219	-230	·178 ·204	·136 ·163	-076 -104	4.022	·881 ·913	·735 ·770	•548 •585
		-	-						•	
21 22	4.273	4.269	4.256	4.230	4.191	4.132	4.052	3-945	3.804	3.623
23	·298 ·323	·294 ·320	·281 ·307	·257 ·283	·218 ·245	·161 ·189	-082 -112	·977 4·009	-839 -873	-660 -698
24	-348	·345	-332	·309	·272	-218	·142	-041	-908	-735
25	·373	·370	-358	-335	-299	·246	-172	-073	-942	-773
26	4.398	4.395	4.383	4.362	4.326	4.275	4.202	4 ·105	3.977	3-811
27	423	•420	409	-388	•353	-303	232	-137	4.011	-848
28	-449	-445	•435	•414	-381	·331	·262	-169	-046	·886
29 30	-474 -499	·471 ·496	-460 -486	•440	-408	·360	•292 •322	·201	-080	·923
30	438	•4590	*400	-466	-435	·388	•322	·233	-115	-961
31	4.524	4.521	4.511	4.493	4.462	4.417	4.352	4.265	4.149	3-999
32	-549	·546	·537	·519	-489	•445	·382	·297	·184	4.036
33 34	·574 ·599	·571 ·597	•563 J88	•545 •571	•516 •544	•473 •502	-412 -442	•329 •361	·219 ·253	-074 -111
35	-624	-622	-614	-598	-571	-530	472	-393	-288	·149
~~										
36 37	4·649 •674	4·647 •672	4-639 -665	4·624 ·650	4·598 -625	4.559	4·502 •532	4.425	4·322 ·357	4.186
38	-699	-697	-690	·676	-652	-587 -615	•562	-457 -489	·391	·224 ·262
39	·724	·723	.716	-702	-679	-644	-592	.521	-426	-299
40	·749	•748	-742	·729	-706	-672	-622	-553	-460	·337
41	4.774	4.773	4.767	4.755	4.734	4.701	4-652	4.585	4.495	4.374
42	•799	-798	.793	-781	•761	-729	-682	-617	-529	412
43	-825	-823	-818	-807	•788	-757	.712	-649	-564	•450
44 45	-850 -875	·849 ·874	-844 -869	-833 -860	-815 -842	·786 ·814	·742 ·772	-682 -714	·598 ·633	-487 -525
				-000	-042	-014	-114	.114	-000	-040
46	4.900	4.899	4.895	4.886	4.869	4.843	4-802	4.746	4.667	4.562
47 48	·925 ·950	-924 -949	-921 -946	-912 -938	-897 -924	-871	-832	-778	•702 •736	•600 •637
49	-975	-975	-972	-965	-924 -951	-899 -928	-862 -892	·810 ·842	•771	-675
50	5.000	5.000	-997	-991	-978	-956	-922	-874	-805	.713
	•				-					

(X = 6-0-6-9; 0-50% kill)

o.,				F	· · · · · · · · · · · · · · · · · · ·		•			
%				-	ected pro	•				
ĿШ	6-0	6-1	6∙2	6.3	6·4	6∙5	6-6	6-7	6∙8	6-9
0	2.523	2.132	1-643	1-030	0-261		-			—
1	2.564	2.178	1-694	1-088	0-327	-	_ `	_	_	_
2	-606	-224	-746	-146	-394	_		_	_	_
3	-647	·270	-797	·205	-461					
4	-688	-316	-849	•263	-528		-			-
5	-730	-362	-900	-321	-595		-	-		_
6	2.771	2.408	1-952	1.380	0-661	_		-		<u> </u>
7	-812	-454	2.003	-438	-728	-				
8	-854	-500	-055	-496	•795		—		-	_
9 10	-895 -936	-546 -591	•106 •158	•555 •613	-862 -928	0-067			—	-
10	-900	-091	-100	-010	-940	0.001		-		
11	2.978	2-637	2.209	1-671	0-995	0.144	_		_	_
12	3-019	-683	·261	·730	1.062	·221		—		—
13	-060	·729	-312	-788	·129	·299	-			
14 15	-102 -143	·775 -821	·364 •415	·846 ·905	·196 ·262	·376 •453			_	—
10	-1-90	-021	-110	-300	-202	-100	_			-
16	3 ·18 4	-2-867	2.467	1-963	1.329	0-530	—		_	-
17	-226	-913	· •518	2.022	•398	-607		—	_	-
18	-267	-959	-570	-060	-463	-685	—			
19 20	·308 ·350	3-005 -050	-621 -673	·138 ·197	•530 •596	•762 •839	_	_		_
20	-500	-000	-010	-101	-030	-009	_			. –
21	3-391	3-096	2.724	$2 \cdot 255$	1.663	0-916		—	-	-
22	-432	-142	-776	·313	•730	-993	0-062		Ξ	
23 24	-474	-188 -234	-827 -879	·372 ·430	-797 -864	1-071 -148	·152 ·243	_	_	
24	-515 -556	·234 ·280	-930	-488	-930	-225	·245 ·333	_	_	_
20	-000	-200	300	200	200		-000	_		_
26	3-598	3-326	2.982	2.547	1-997	1.302	0-423			
27	-639	-372	3-033	-605	2-064	-379	-513	—	-	—
28	-680	-418	-085	-663 -722	-131	-457 -534	-603	—		
29 30	·721 ·763	-464 -509	·136 ·188	-780	-197 -264	-611	-693 -784	_	_	
	-100	000	100	100	201	V11	101			
31	3-804	3.555	3-239	2.838	2.331	1-688	0-874	—	_	-
32	-845	-601	·291	-897	-398	•766	-964	0.050		-
33 34	-887 -928	-647 -693	·342 ·394	-955 3-014	-465 -531	-843 -920	1-054 -144	0-050 -156	_	
35	-969	-739	-445	-072	-598	-920	-234	-262	_	_
	000	100								
36	4-011	3.785	3.497	3.130	2-665	2.074	1.324	0-369		
37	-052	-831	-548	-189	-732	-152 -229	-415	-475		—
38 39	-093 -135	-877 -923	-600 -651	·247 ·305	-799 -865	·229 ·306	-505 -595	-581 -688	_	_
40	-176	·969	-703	-364	-932	-383	-685	•794	_	_
			-							
41	4-217	4-014	3.754	3.422	2.999	2.460	1.775	0.900		_
42 43	·259	-060	-806	-480	3-066 -132	•538 •615	•865 •955	1-007 -113	0-035	
43 44	·300 ·341	·106 ·152	-857 -909	•539 •597	-132 -199	-613	-955 2-046	·113 ·219	162	_
45	-383	-198	-960	-655	-266	-769	-136	-326	-289	
46	4.424	4.244	4-012	3.714	3.333	2-846 -924	2·226 ·316	1·432 ·538	0·415 ·542	
47 48	-465 -507	·290 ·336	-063 -115	· •772 •830	-400 -466	-924 3-001	·310 ·406	·əsə -645	•042 •669	_
40 49	·507 ·548	·330 ·382	·115 ·166	-889	-533	-078	-496	-751	.795	_
50	-589	-428	-218	-947	-600	·155	-586	-857	.922	
		-		-						

(Y = 3-0-3-9; 51-100 % kill)

%				Ex	pected pr	obit, Y				
kill	3-0	3-1	3.2	3.3	3.4	3.5	3-6	3.7	3.8	3·9
51	_		9 ·205	8.249	7.504	6.922	6-467	6-111	5.834	5-618
52			-331	-355	-594	-999	-534	-170	-885	-664
53	—	<u> </u>	-458	-462	-684	7-076	-600	-228	-937	·710
54	-		-585	-568	.774	·154	-667	·286	-988	·756
55	-	 .	-711	-674	-864	·231	•734	·345	6-040	-802
56			9-838	8 ·781	7.954	7.308	6 ·801	6-403	6-091	5.848
57			-965	-887	8-045	-385	-868	-461	·143	·894
58		—		-993	·135	-462	-934	-520	-194	-940
59 60		—		9.100	-225 -315	-540 -617	7-001	-578	·246	-986
00	-		<u> </u>	·206	-519	-017	-068	-636	-297	6-031
61	—	. .	<u> </u>	9.312	8-405	7-694	7.135	6-695	6.349	6-077
62 63		-	_	-419 -525	-495	-771	·201	•753	-400	·123
63 64			_	•525 •631	•585 •676	-848 -926	•268 •335	-811 -870	-452 -503	·169
65	_	=	_	-738	-766	8-003	-402	-928	-505	·215 ·261
						0.000	-104	-920	000	-201
66			<u> </u>	9.844	8.856	8-080	7-469	6-986	6-606	6-307
67	-	_		-950	-946	·157	-535	7 0 4 5	-658	•353
68 69	-	-		—	9·036 ·126	-234	-602	·103 ·162	-709	·399
70	_		_	_	·126 ·216	·312 ·389	-669 •736	·102 ·220	·761 ·812	-445 -491
	_		_	_	-210	-209	.190	-220	-012	491
71		—		—	9·3 07	8.466	7.803	7.278	6 ∙864	6.536
72	-		—		·397	-543	-869	·337	915	-582
73 74	_			_	-487	-621 -698	-936	·395	-967	-628
75	_	_	_	_	-577 -667	-775	8-003 -070	-453 -512	7-018 -070	-674 -720
76	-		—		9.757	8.852	8-136	7-570	7.121	6.766
77 78	_	_	*****	_	-848 -938	-929 9-007	-203 -270	-628 -687	•173	·812
79	_	_	=	_	-930	-084	·270 ·337	-745	-224 -276	-858 -904
80				_	_	-161	-404	-803	-327	-950
81 82				—	—	9·238	8-470	7.862	7.379	6-995
83	_	_	_	_		·315 ·393	•537 •604	-920 -978	-430 -482	7-041
84	_	_	=	=	_	-395 -470	-671	8-037	-402	-087 -133
85	_	_		_	_	-547	-738	-095	-585	179
60									•	
86 87	_	—	-			9-624	8-804	8.154	7-636	7.225
88	=	_	-	_	_	-701 -779	-871 -938	·212 ·270	-688 -739	·271 ·317
89	_	_	=	=	_	-856	9.005	-329	-791	·363
90	_					-933	072	-387	-842	-409
01										
91 92	_			—	—	—	9.138	8.445	7.894	7.454
93	_	_	_			—	·205	-504 -562	-945 -997	-500
94		_	_	_	_		·272 ·339	•302 •620	8-048	-546 -592
95				_	_	_	-405	-679	•100	-638
96				•						
97	_		_	_	_	_	9·472 ·539	8·737 ·795	8·151 ·203	7·684 ·730
98	_	_	_			_	-559 -606	•195 •854	-205 -254	776
99	—	_					-673	-912	-306	-822
100		-	-		_	_	-739	-970	-357	-868

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(Y-4-0-4-9; 51-100% kill)

%				Exp	ected pr	obit, <i>Y</i>				
kill	4-0	4.1	4 ·2	4 ·3	4 ·4	4.5	4-6	4.7	4-8	4.9
51	5.452	5.325	5.229	5-158	5.108	5.072	5-049	5-035	5-028	5-025
52	• 4 9 3	·363	·264	·190	·138	·101	-076	-062	-054	-051
53	-535	-400	-298	·222	·168	·129	·103	-088	-079	-076
54	-576	•438	·333	·254	·198	·157	·131	·114	·105	·101
55	-617	-475	-367	-286	-228	·186	·158	·140	•131	•126
56	5.659	5·513	5.402	5.318	5.258	5·214	5.185	5 ·167	5 ·158	5.151
57	•700	•550	•436	·351	·288	·243	·212	·193	·182	·177
58	·741	-588	-471	·383	·318	·271	·239	·219	·207	·202
59	-783	-626	•505	-415	-348	·299	·266	·245	·233	·227
60	·824	-663	·540	•447	-378	·328	-294	·271	·258	·252
61	5.865	5.701	5.574	5.479	5-408	5.356	5.321	5.298	5.284	5.277
62	·907	-738	-609	-511	-438	-385	-348	·324	·310	·303
63	·948	·776	-643	·543	•468	•413	·375	·350	·335	·328
64	-989	·814	-678	-575	-498	•441	-402	·376	-361	·353
65	6-031	-851	$\cdot 712$	-607	-528	-470	•429	-402	•386	-378
66	6 0 7 2	5.889	5.747	5.639	5.558	5-498	5.456	5.429	5.412	5.403
67	-113	·926	-781	-671	-588	-527	-484	-455	-437	-429
68	·155	·964	-816	·703	-618	·555	-511	-481	-463	-454
69	·196	6-001	·851	•735	-648	·583	·538	-507	-489	•479
70	-237	-039	-885	•767	-678	-612	•565	•534	·514	•504
71	6.279	6-077	5-920	5.799	5.708	5.640	5.592	5.560	5.540	5.529
72	·320	·114	·954	·831	·738	·669	-619	·586	•565	•55 5
73	·361	-152	-989	·863	•768	-697	-647	-612	•591	·580
74	·402	-189	6.023	·895	-798	·725	-674	-638	-617	-605
75	-444	·227	-058	-927	-828	·754	·701	-665	-642	-630
76	6.485	6.265	6.092	5.959	5.858	5.782	5.728	5.691	5-668	5-655
77	-526	·302	-127	·991	·888	·811	·755	•717	-693	·680
78	•568	·340	-161	6-023	·918	-839	•782	•743	•719	•706
79	-609	-377	·196	-055	-948	·868	-809	·770	-744	·731
80	·650	•415	-230	-087	•978	·896	-837	-796	-770	•756
81	6-692	6.452	6.265	6·119	6-008	5 924	5.864	5.822	5.796	5.781
82	·733	•490	-299	-151	-038	·953	-891	-848	-821	·806
83	.774	-528	·334	·183	-068	-981	-918	·874	·847	·832
84	-816	-565	·368	·215	-098	6.010	·945	·901	·872	• 857
85	-857	-603	-403	·247	·128	-038	-972	-927	-898	-882
86	6.898	6.640	6.437	6.279	6·158	6-066	6-000	5-953	5 ·923	5.907
87	·940	-678	·472	-311	·188	-095	-027	-979	·949	·932
88	-981	•716	•506	·343	·218	·123	-054	6-006	·975	-958
89	7.022	·753	·541	·375	-248	·152	-081	-032	6-000	-983
90	·064	•791	-575	-407	·278	·180	·108	-058	-026	6-008
91	7.105	6.828	6.610	6.439	6-308	6-208	6.135	6-084	6-051	6-033
92	•146	·866	•644	-471	-338	·237	·162	·110	-077	-058
93	·188	·903	-679	·503	·368	-265	·190	·137	.102	-084
94	$\cdot 229$	·941	·713	-535	·398	·294	·217	·163	·128	109
95	-270	-979	·748	-567	· 4 28	·322	·244	·189	·154	-134
96	7.312	7-016	6.783	6-600	6.458	6.350	6.271	6.215	6-179	6.159
97	-353	-054	-817	-632	·488	-379	·298	·242	·205	·184
98	·394	-091	-852	-664	-518	-407	-325	-268	-230	·210 ·235
99	-436	-129	·886	-696	·548	-436	•353	·294	·256 ·281	·255 ·260
100	-477	•166	-921	-728	•578	-464	-380	·320	-201	-200

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(Y = 5-0-5-9; 51-100 % kill)

%				Ехр	ected pro	bit, F				
kill	5-0	5-1	5-2	5-3	5-4	5.5	5-6	5-7	5.8	5-9
51	5-025	5-025	5 023	5-017	5-005	4-985	4-953	4-906	4.840	4.750
52	-050	-050	-048	-043	-032	5-013	-983	-938	-874	-788
53	-075	-075	-074	-069	-059	-041	5-013	-970	-909	-825
54 55	·100 ·125	·100 ·126	-100 -125	-096 -122	-087 -114	-070 -098	-043 -073	5-002 -034	-943 -978	-863 -901
00	-120	-120	-140	-140	-114	-080	-013	-00-2	-910	-901
56	5 ·150	5 ·151	5-151	5.148	5.141	5.127	5.103	5-066	5-012	4-938
57	-175	-176	-176	-174	·168	·155	-133	-098	-047	-976
58 59	-201 -226	·201 ·226	·202 ·227	-201 -227	·195 ·222	-183 -212	-163 -193	-130 -162	-082 -116	5-013 -051
60	·251	-252	253	-253	·250	·240	·223	-194	-151	-068
61	5.276	5.277	5.279	5.279	5.277	5.269	5.253	5.228	5.185	5.128
62 63	·301 ·326	·302 ·327	-304 -330	•305 •332	-304 -331	-297 -325	·283 ·313	-258 -290	-220 -254	-164 -201
64	-351	-352	-355	-358	•358	·354	-343	-322	-289	-239
65	-376	-378	-381	-384	-385	-382	·373	-354	-323	·276
66 67	5-401 -426	5-403 -428	5-406 -432	5-410 -437	5·412 -440	5-411 -439	5-403 -433	5·386 -418	5-358 -392	5·314 ·351
68	451	-453	-458	-463	-467	-467	-463	-450	-427	-389
69	-476	-478	-483	-489	-494	-496	-493	-482	-461	-427
70	-501	-504	-509	-515	-521	-524	-523	-514	-496	-464
71	5-526	5 ·529	5-534	5.541	5 ·548	5 ·553	5.553	5.546	5-530	5.502
72	-551	-554	-560	-568	-575	-581	-583	-578	-565	-539
73	-577	-579	-585	-594	-603	-609	-613	-610	-599	-577
74 75	-602 -627	-604 -630	-611 -637	-620 -646	-630 -657	-638 -666	-643 -673	-642 -674	-63 <u>4</u> -668	-615 -652
-	-						010	VII		
76	5-652	5-655	5-662	5-673	5-684	5-695	5.703	5-706	5.703	5-690
77 78	-677 -702	-680 -705	-688 -713	-699 -725	-711	-723	-733	-738	-737	•727 •765
79	-727	-730	-739	•751	•738 •765	-752 -780	-763 -793	-770 -802	-772 -806	-802
80	-752	-755	-764	.777	.793	-808	-823	-834	-841	-840
81	5.777	5 ·781	5.790	E 004	F 000	E 000	F 059	F 000	EOTE	E 070
82	•802	-806	-816	5-804 -830	5-820 -847	5-837 -865	5-853 -883	5·866 -898	5-875 -910	5·878 -915
83	-827	-831	-841	-856	-874	-694	-913	-930	-944	-953
84	·852	-856	-867	-882	-901	-922	-943	-962	-979	-990
85	-877	·881	-892	-968	-928	-950	-973	-995	6-014	6-028
86	5-902	5-907	5-918	5-935	5-956	5-979	6-003	6-027	6-048	6-066
87	-927	-932	-943	-961	-983	6-007	-033	-059	-083	-103
88 89	-953 -978	-957 -982	-969	-987	6-010	-036	-063	-091	-117	-141
90	6-003	6-007	-995 6-020	6-013 -040	-037 -064	-064 -092	-093 -123	-123 -155	-152 -186	-178 -216
~ 1										
91 92	6-028 -053	6-033 -058	6-046 -071	6-066 -092	6-091	6.121	6·153	6.187	6·221 •255	6·253 ·291
93	-078	-063	-097	-118	·118 ·146	•149 •178	·183 ·213	·219 ·251	·255 ·290	·291 ·329
94	·103	·108	-122	-144	-173	-206	-243	-283	-324	-366
95	-128	-133	-148	-171	-200	-234	-273	-315	359	-404
96	6 ·153	6 ·159	6.174	6 ·197	6.227	6-263	6-303	6·347	6-393	6.441
97	-178	·184	·199	-223	-254	·291	·333	-379	-428	-479
98 99	·203 ·228	-209	·225	·249	·281	-320	-363	-411	-462	-517
100	·253	-234 -259	·250 ·276	-276 -302	-309 -336	•348 •376	·393 -423	-443	-497 -531	-554 -592
			-210	-004	.000	-376	* 1 65	-475	160-	-074

(Y = 6-0-6-9; 51-100 % kill)

	%				Ern	ected pro	hit V				
	∕o kill	6-0	6-1	6·2	6·3 ·	6·4	6.5	6-6	6.7	6-8	6.9
	51	4-631		4.269		3-667	3.233	2-677	1.964		
	51 52	4-031	4·473 •519	4·209 •321	4.006	-734	3·235 •310	-767	2.070	1-049 ·175	0-022
	53	-713	-565	-372	.122	-800	387	-857	-176	-302	.175
	54	-755	-611	-424	·181	-867	-464	-947	-283	-429	-327
	55	-796	-657	-475	·239	-934	•541	3.037	-389	•555	-480
	56.	4.837	4.703	4.527	4.297	4.001	3-619	3 ·127	2-495	1.682	0.632
	57	-879	-749	-578	-356	-068	-696	-218	602	-809	-784
	58	-920	.795	-630	-414	-134	·773	308	•708	-935	·784 ·937
	59	-961	-841	-681	·472	-201	-850	·398	814	2.062	1.089
	60	5.003	-887	·733	·531	·268	-927	•488	-921	·189	·242
	61	5-044	4∙932 -978	4.784	4.589	4.335	4-005	3.578	3-027	2 ·315	1.394
	62	-085	-978	·836	-647	•401	-082	-668	-133	·442	·546
	63	·127	5-024	·887	-706	-468	·159	-758	·240	•569	-699
	64	·168	-070	·939	·764	-535	·236	·849	-346	-695	•851
	65	-209	·116	-990	·823	-602	·31 3	·939	-452	-822	2-004
	66	5 ·251	5.162	5-042	4.881	. 4 ·669	4.391	4-029	3.559	2 ·949	2.156
	67	·292	-208	-093	·939	·735	-468	·119	-665	3-075	·308
	68	·333	-254	-145	-998	-302	·545	·209	•771	·202	-461
	69	·375	·300	·196	5-056	·869	-622	·299	-878	-329	-61 3
	70	·416	·346	-248	-114	-936	-700	•390	·984	•455	•766
•	71	5-457	5-392	5 ·299	5.173	5.003	4.777	4.480	4-090	3 ·582	2.918
	72	-499	-437	·351	·231	-069	·854	·570	-197	·709	3.070
	73 74	·540	-483	-402	·289	-136	·931	-660	·303	•835	·223
	74	-581	-529	•454	•348	·203	5 008	-750	-409	·962	·375
	75	-623	•575	•505	-406	·270	-086	·840	•516	4-089	·528
	76	5.664	5.621	5 ·557	5.464	5.336	5.163	4-930	4.622	4 ·215	3.680
	77	·705	-667	-608	-523	•403	·240	5.021	-728	·342	·832
	78	·747	-713	-660	-581	-470	-317	·111	·835	-469	·985
	79	•788	•759	•711	-639	-537	-394	·201	-941	·595	4.137
	80	·829	·805	•763	-698	·604	•472	·291	5-047	·722	·290
	81	5.870	5.851	5-814	5.756	5.670	5.549	5-381	5.154	4·849	4.442
	82	·912	·896	-866	·815 ·873	-737	-626	•471	·260	·975	·594
	83	·953	-942	·917	·873	·804	·703	-561	·366	5.102	·747
	84	·994	-988	·969	-931	-871	·780	652	-473	·229	-899
	85	6-036	6-034	6-020	·990	-938	·858	·742	•579	·355	5-052
	86	6.077	6-080	6.072	6-048	6-004	5-935	5.832	5-685	5.482	5.204
	87	-118	-126	·123	·106 ·	-071	6-012	.922	·792	-609	-356
	88	·160	·172	•175 ·	·165 ·	·138	-089	6-012	-898	·735	-509
	89	·201	·218	·226	·223 ·	-205	·166	·102	6.004	·862	-661
	90	·242	·264	-278	-281	-272	·244	-192	-111	-988	·814
	91	6-284	6-310	6-329	6-340	6.338	6.321	6.283	6-217	6.115	5.966
	92	·325	-355	-381	. •398	-405	•398	·373	·323	-242	6.118
	93	·366	-401	•452	-400	-472	-475	·463	-430	·368	·271
	94	•408	-447	-484	·515	-539	·553	-553	•536	•495	·423
	95	-4-19	-493	-535	-573	-605	-630	-643	-642	-622	•576
	96	6-490	6-539	6 ·587	6-631	6-672	6·707	6.733	6.749	6.748	6-728
•	97	-532	-585	-638	-690	-739	-784	·824	-855	•875	-880
	98	-573	-631	-690	·748	-806	·861	·914	·961	7.002	7-033
·	99	-614	-677	·741	-807	·873	-939	7.004	7.068	-128	-185 -338
	100	-656	-723	•793	-865	-939	7-016	-094	-174	-255	-000

(Y = 7.0-7.9; 51-100% kill)

%				Ex	pected pr	obi t, Y				
kill	7-0	7.1	7.2	7.3	7-4	7.5	7-6	7.7	-7-8	7.9
51	_		_	_	_	_	_	_		
52	_	-						—		-
53		—	-	-	—	—	—	—	_	<u> </u>
54 55	_	-	-	_			-		—	
99		-	-	-	—	_		-	—	—
56			_	_	_	_		—		
57		—	—	<u> </u>			—	-		_
58	-	-		-				-		
59 60	0-013	_	•_	_	_		_	_	Ξ	_
					_					
61	0.198	_		_				-	<u> </u>	
62	·383	—	· —		<u> </u>	Ξ	—			—
63 64	•568 •753					_		<u> </u>	—	
65	-939	=	_	=	=	_	_	_	_	_
66	1.124				<u> </u>		- '	—	—	
67 68	·309 ·494	0-003 -231	—	—	-	_	-	-	—	_
69	-680	·251 ·458	_	_	_			_		
70	-865	-685		_	_	_	_	=	_	=
	•									
71	2.050	0.913	-	<u> </u>				—	—	—
72 73	·235 ·420	1·140 ·367			—		_		-	-
74	-606	-595	0.263	_	_	<u> </u>	—	=	_	
75	•791	-822	·545			_	_		_	
76	2.976	2.050	0.007							
77	2·970 3·161	-277	0·827 1·108			-	_	·		-
78	-347	-504	390	_	_	=	_		_	_
79	-532	.732	672	0.265	_		<u> </u>			
80	-717	·9 59	·9 5 4	-618		—	_	—		
81	3.902	3.186	$2 \cdot 236$	0.971		_	_		_	_
82	4-087	-414	-518	1.324	. —		_	_		
83	·273	-641	•800	-677	0.175	_	—	—	_	<u> </u>
84 85	-458 -643	-868 4-096	3.082	2.030	-621	<u> </u>	—		—	
00	-045	4090	·364	•383	1-068	-				
86	4.828	4.323	3.645	2 ·736	1.514	_		_		·
87	5-014	•551	-927	3-089	-961	0.438	<u> </u>		<u> </u>	_
88 89	·199 ·384	-778	4.209	-442	2-408	1.008				_
90	•569	5-005 -233	·491 ·773	-795 4-148	•854 3∙301	·579 2·149	0.581	-		
				1 110	0.001	4·1·13	0.001		_	
91	5.754	5.460	5-055	4.501	3.747	2.720	1.317	—		
92 93	-940 6·125	-687	·337	·854	4.194	3.290	2.054	0.356	_	-
93 94	·125 ·310	-915 6-142	-619 -901	5·207 •560	·640	.861	-790	1.316	0.540	
95	-495	•369	6.182	·200 ·914	5·087 ·533	4·431 5·002	3·526 4·262	2·275 3·235	0·542 1·806	_
						0 000	* 808	0.000	1.000	—
96 97	6-681	6.597	6.464	6-267	5.980	5.572	4-998	4.194	3.069	1.493
91 98	-866 7-051	-824 7-051	•746 7•028	-620 -973	6.426	6.143	5.735	5.154	4.333	3.173
99	-236	-279	-310	7.326	·873 7·319	•713 7•284	6·471 7·207	6·114 7·073	5-596 6-859	4·853 6·533
100	-421	-506	-592	-679	•766	-854	-943	8.033	8.123	0.555 8.213
				•	.•					
	FPA								20	

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TABLE V. The Probability, P, the Ordinate, Z, and Z^2

Y	P	Z	Z ²
5.0	0-50000	0.39894	0.1591
5-1	0.53983	0.39695	0.15757
5.2	0.57926	0.39104	0.12291
5.3	0.61791	0.38139	0.14546
5.4	0.65542	0.36827	0.13562
5.5	0.69146	0·35207	0.12395
5.6	0.72575	0.33322	0.11104
5.7	0.75804	0.31225	0.09750
5.8	0.78814	0.28969	0.08392
5-9	0.81594	0.26609	0.02080
6·0	0.84134	0.24197	0-05858
6·1	0.86433	0.21785	0.0474
6·2	0.88493	0-19419	0.0377
6·3	0.903200	0.17137	0.0293
6.4	0.919243	0.14973	0.02242
6 ∙5	0.933193	0.12952	0.0167
6·6	0.945201	0.11092	0.0123
6·7	0.955435	0.094049	0.0088
6-8	0.964070	0.078950	0.0062
6.9	0.971283	0.065616	0.0043
7.0	0.977250	0.053991	0.00292
7.1	0.982136	0.043984	0.0019
7.2	0.986097	0.035475	0.0012
7.3	0.989276	0.028327	0.0008
7.4	0.9918025	0.022395	0.0005
7.5	0.9937903	0.017528	0.0003
7.6	0.9953388	0.013583	0.0001
7.7	0.9965330	0.010421	0.0001
7.8	0.9974449	0.0079155	0.0000
7.9	0.9981342	0.0059525	0.00004
8 ∙0	0.9986501	0.0044318	0.0000
8 ∙1	0.99903240	0.0032668	0.0000
8.2	0.99931286	0.0023841	0.0000
8.3	0.99951658	0.0017226	0.0000
8.4	0.99966307	0.0012322	0.00000
8.2	0.99976737	0.00087268	0.0000
8∙6	0.99984089	0.00061190	0.0000
8.7	0.99989220	0.00042478	0.00000
8-8	0.999927652	0.00029195	0.00000
8-9	0.999951904	······································	0.00000
9.0	0-999968329	0.00013383	0.00000

Values of P and Z are tabulated above, to 5 significant digits, in order to aid the formation of functions that may be required in any new developments of probit methods; Z^2 is the weighting coefficient introduced in §47. When Y is less than 5.0, read the entries for (10-Y) as Q, Z, Z^2 .

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Degrees of				P	robability	y.			
freedom	•90	•70	·50	•30	·10	-05	•02	-01	· 0 01
1	·016	·15	•45	1.1	2.7	3.8	5-4	6.6	10-8
2	•21	•71	1.4	2.4	4 ·6	6.0	7.8	9 ·2	13.8
1 2 3 4 5	•58	1.4	2.4	3.7	6.3	7.8	9.8	11.3	16.3
4	1.1	2.2	3.4	4 ·9	7.8	9.5	11.7	13.3	18-5
5	1.6	3.0	4.4	6.1	9 ∙2	11-1	13.4	15-1	20.5
6 7 8 9	2.2	3.8	5.3	7.2	10.6	12.6	15.0	16.8	22.5
7	2.8	4.7	6.3	8.4	12.0	14.1	16.6	18.5	24.3
8	3.5	5.5	7.3	9.5	13.4	15.5	18.2	20.1	26.1
9	4.2	6.4	8.3	10.7	14.7	16.9	19.7	21.7	27.9
10	4.9	7.3	9.3	11.8	16-0	18.3	21.2	23.2	29.6
12	6.3	9-0	11.3	14.0	18.5	21.0	24.1	26.2	32.9
14	7.8	10.8	13.3	16.2	21.1	23.7	26.9	29.1	36-1
16	9.3	12.6	15.3	18.4	23.5	26.3	29.6	32.0	39.3
18	10.9	14.4	17.3	20.6	26.0	28.9	32.3	34.8	42.3
20	12.4	16.3	19-3	22.8	28·4	31.4	35.0	37.6	4 5·3
22	14.0	18-1	21.3	24.9	30.8	33.9	37.7	40·3	48 ·3
24	15.7	19.9	23.3	27.1	33.2	36.4	40.3	43.0	51.2
26	17.3	21.8	25.3	29.2	35.6	38.9	42.9	45.6	54·1
28	18.9	23.6	27.3	31.4	37.9	41.3	45.4	48.3	56.9
30	20.6	25.5	29.3	33.5	40.3	43.8	48.0	50.9	59.7

TABLE VI. Distribution of χ^2

When χ^s is based on more than 30 degrees of freedom, the quantity $\sqrt{(2\chi^s)} - \sqrt{(2f-1)}$ (where f is the number of degrees of freedom) has approximately the following distribution:

>30 | -1.28 - 0.52 0.00 0.52 1.28 1.64 2.05 2.33 3.09

I am indebted to Professor R. A. Fisher and Dr F. Yates, and also to Messrs Oliver and Boyd, Ltd., of Edinburgh, for permission to print Table VI as an abridgement of Table IV of their book *Statistical Tables for Biological*, *Agricultural and Medical Research*.

Degrees of	Probability								
freedom	-90	-70	-50	•30	·10	~ ∙05	-02	•01	-001
1	•16	- <u>-</u>	1.00	1.96	6.31	12.7	31.8	63.7	637.
2 3 4 5	•14	-44	-82	1.39	2.92	4.30	6.96	9.92	31.6
3	•14	·42	•76	1.25	2.35	3.18	4.54	5-84	12.9
4	-13	-41	-74	1.19	2.13	2.78	3 ·75	4.60	8-61
5	•13	-41	-73	1.16	2.02	2.57	3 ∙36	4 ·03	6 ·86
6	-13	-40	-72	1.13	1.94	2.45	3.14	3.71	5.96
7	-13	•40	•71	1.12	1.90	2.36	3 ⋅00	3.50	5.40
. 8	·13	·40	•71	1.11	1.86	2.31	2.90	3 ·36	5.04
9	·13	• 4 0	•70	1.10	1.83	2.26	2.82	3.25	4.78
10	-13	-40	•70	1.09	1.81	2.23	2.76	3.17	4.59
12	13	•40	•70	1.08	1.78	2.18	2.68	3.06	4.32
14	•13	•39	-69	1.08	1.76	2.14	2.62	2.98	4.14
16	•13	·39	•69	1.07	1.75	2.12	2.58	2.92	4.02
18	•13	-39	-69	1.07	1.73	2.10	2.55	2.88	3 ·92
20	•13	•39	•69	1.06	1.72	2·09	2.53	2·84	3 ·85
22	-13	•39	-69	1.06	1.72	2.07	2.51	2 ·82	3.79
24	•13	-39	•68	1.06	1.71	2.06	2.49	2.80	3.74
26	•13	•39	•68	1.06	1.71	2.06	2.48	2.78	3.71
28	-13	•39	-68	1.06	1.70	2.05	2.47	2.76	3.67
30	•13	·39	•68	1.06	1.70	2.04	2·46	2.75	3.65
40	·13	-39	-68	1.05	1.68	2.02	2.42	2.70	3 ·55
60	-13	·39	-68	1.05	1.67	2.00	2.39	2.66	3 ∙46
120	·13	-39	-68	1.04	1.66	1-98	2.36	2.62	3.37
Ø	·126	·385	-674	1.036	1.645	1.960	2·3 26	2.576	3 ·291

TABLE VII. Distribution of t

I am indebted to Professor R. A. Fisher and Dr F. Yates, and also to Messrs Oliver and Boyd, Ltd., of Edinburgh, for permission to print Table VII as an abridgement of Table III of their book *Statistical Tables for Biological*, *Agricultural and Medical Research*.

Expected probit Y	Weighting coofficient w	Expected probit Y	Weighting coefficient w	
<u> </u>	0.00000 004	5.1	0.34242	
1.2	0.00000 009	5.2	0.36344	
1.3	0.0000002	5.3	0.38069	
1.4	0.00000 04	5.4	0.39359	
1.5	0.00000 08	5-5	0.40173	
1.6	0.00000 2	5.6	0.40488	
1.7	0·00000 3	5.7	0.40296	
1.8	0.00000 6	5-8	0.39612	
1.9	0.00001	5.9	0.38466	
2.0	0.00002	6.0	0.36904	
2.1	0-00004	6-1	0.34983	
2.2	0.00006	6.2	0.32770	
2.3	0.00011	6.3	0·30338	
2.4	0.00019	6.4	0.27760	
2.5	0.00031	6.5	0.25109	
2.6	0-00051	6.6	0.22452	
2.7	0.00081	6.7	0.19848	
2.8	0.00128	6.8	0.17348	
2.9	0.00197	6-9	0.14993	
3.0	0.00298	7.0	0.12813	
3.1	0.00443	7.1	0.10829	
3.2	0.00647	7.2	0.09051	
3.3	0.00926	7.3	0.07482	
3.4	0.01302	7.4	0.06118	
3 ∙5	0.01798	7.5	0.04948	
3.6	0.02439	7-6	0.03958	
3.7	0.03251	7.7	0.03132	
3.8	0.04261	7.8	0.02452	
3.9	0.05491	7.9	0.01899	
4 ∙0	0.06959	8.0	0.01455	
4.1	0.08677	8-1	0.01103	
4.2	0.10648	8.2	0.00827	
4.3	0.12863	8.3	0.00614	
4.4	0.15300	8.4	0.00421	
4 ∙5	0.17926	8.5	0.00327	
4.6	0.20692	8.6	0.00235	
4.7	0.23540	8.7	0.00167	
4.8	0.26398	8.8	0.00118	
4.9	0.29189	8.9	0.00082	
5.0	0.31831	9·0	0.00057	

TABLE VIII. The Weighting Coefficient in Wadley's Problem

Expected probit Y	$\begin{array}{c} \text{Minimum} \\ \text{working probit} \\ Y - Q/Z \end{array}$	Range Q [*] /Z	Weighting coefficient Z ² /PQ ³
5-0 ,	3.7467	0-62666	1.2732
5-1	3.9407	0.53346	1.3784
5.2	4.1241	0-45269	1.4912
5.3	4.2982	0-38279	1.6124
5.4	4.4643	0-32241	1.7428
5.2	4-6236	0.27039	1.8831
5.6	4.7770	0.22572	2.0342
5.7	4.9251	0.18750	2.1970
5.8	5.0687	0.15493	2·3724
5.9	5·2083	0.12732	2·5613
6-0	5.3443	0.10403	2.7647
6-1	5.4773	0.084485	2.9833
6.2	5-6074	0.068187	3-2181
6.3	5.7351	0.054679	3.4700
6.4	5.8606	0.043557	3.7395
6.2	5.9842	0.034460	4.0275
6.6	6 ·1060	0·027073	4.3346
6·7	6 ∙2261	0-021117	4.6613
6-8	6 ∙3449	0.016352	5.0081
6-9	6.4624	0.012568	5·3753
7.0	6.5786	0.0095862 .	5.7633
7.1	6-6938	0.0072558	6 1721
7.2	6.8081	0.0054491	6-6019
7.3	6.9214	0.0040600	7.0528
7.4	7.0339	0.0030007	7.5247
7-5	7.1457	0.0021999	8.0177
7.6	7.2568	0.0015996	8·5 315
7.7	7.3673	0.0011534	9.0661
7.8	7.4772	0.00082480	9.6214
7.9	7 ·5866	0-00058484	10-1971
8 ∙0	7.6954	0.00041117	10.7933
8.1	7.8038	0.00028660	11-4097
8·2	7.9118	0.00019802	12.0463
8.3	8.0194	0.00013567	12.7030
8.4	8-1266	0.000092128	13.3797
8.5	8·2334	0.000062011	14.0762
8.6	8.3400	0.000041372	14-7926
8.7	8.4462	0.000027357	15.5289
8.8	8.5522	0.000017929	16-2859
8.9	8.6579	0.000011645	17.0607
9.0	8.7633	0.0000074951	17.8563

TABLE IX. N	Ainimum	Working	Probit, I	Range, and	i
Weighting	Coefficien	t for Inv	erse Sam	pling	

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